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Superoxide and Singlet Oxygen Produced within the Thylakoid Membranes Both Cause Photosystem I Photoinhibition^{1[OPEN]}

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Photosystem I (PSI) photoinhibition suppresses plant photosynthesis and growth. However, the mechanism underlying PSI photoinhibition has not been fully clarified. In this study, in order to investigate the mechanism of PSI photoinhibition in higher plants, we applied repetitive short-pulse (rSP) illumination, which causes PSI-specific photoinhibition in chloroplasts isolated from spinach leaves. We found that rSP treatment caused PSI photoinhibition, but not PSII photoinhibition in isolated chloroplasts in the presence of O₂. However, chloroplastic superoxide dismutase and ascorbate peroxidase activities failed to protect PSI from its photoinhibition. Importantly, PSI photoinhibition was largely alleviated in the presence of methyl viologen, which stimulates the production of reactive oxygen species (ROS) at the stromal region by accepting electrons from PSI, even under the conditions where CuZn-superoxide dismutase and ascorbate peroxidase activities were inactivated by KCN. These results suggest that the ROS production site, but not the ROS production rate, is critical for PSI photoinhibition. Furthermore, we found that not only superoxide (O₂⁻) but also singlet oxygen (¹O₂) is involved in PSI photoinhibition induced by rSP treatment. From these results, we suggest that PSI photoinhibition is caused by both O₂⁻ and ¹O₂ produced within the thylakoid membranes when electron carriers in PSI become highly reduced. Here, we show, to our knowledge, new insight into the PSI photoinhibition in higher plants.

Higher plants need light to drive photosynthesis. However, excess light causes photoinhibition in chloroplasts (Melis, 1999). Photosystem (PS) II is very sensitive to environmental stress, such as high-light intensities or drought conditions, and such stress conditions can cause a decrease in D1 protein of the PSII reaction center (Aro et al., 1993). This phenomenon is called photoinhibition of PSII, and its occurrence greatly reduces plant productivity (Melis, 1999; Külheim et al., 2002). In previous studies, photoinhibition in PSII has been shown to be caused by singlet oxygen (¹O₂), which is produced through a charge recombination reaction between P680⁺ and the reduced secondary electron acceptor (Q_A⁻) in PSII. Next, ¹O₂ is thought to stimulate the degradation of

D1 protein (Asada and Takahashi, 1987; Krieger-Liszkay, 2005; Hideg et al., 2007; Gill and Tuteja, 2010; Vass, 2011). In addition to ¹O₂, recent studies have revealed that superoxide (O₂⁻) is also produced by PSII, which also causes photoinhibition (Bondarava et al., 2010; Zulfugarov et al., 2014). In contrast to the theory that oxidative degradation of the D1 protein causes photoinhibition in PSII, several studies now suggest that reactive oxygen species (ROS) suppress de novo D1 protein synthesis through the oxidative inactivation of the thioredoxin-regulated elongation factor G. The latter plays an important role in protein translation of the D1 protein (Kojima et al., 2007; Nishiyama et al., 2011).

In addition to PSII, PSI can also experience photoinhibition, and photoinhibition in PSI is similarly caused by ROS (Sonoike and Terashima, 1994; Terashima et al., 1994; Sonoike et al., 1995; Sonoike, 1995, 1996; Sejima et al., 2014). In PSI, the risk of ROS production increases when the photosynthetic electron transport chain is in a highly reduced state (Sonoike and Terashima, 1994; Allahverdiyeva et al., 2005; Oelze et al., 2012; Grieco et al., 2012). In fact, PSI photoinhibition occurs when the PSI electron carriers become reduced, impairing net carbon assimilation, and hence plant growth (Allahverdiyeva et al., 2005; Munekage et al., 2008; DalCorso et al., 2008; Suorsa et al., 2012; Grieco et al., 2012; Kono et al., 2014). An important difference between photoinhibition in PSI and PSII is that PSI recovers very slowly, whereas

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photoinhibited PSII recovers rapidly ($t_{1/2}$ is about 60 min; Melis, 1999; Sonoike, 2011). Therefore, PSI photoinhibition has more severe consequences than PSII photoinhibition in higher plants.

Many studies have confirmed the occurrence of PSI photoinhibition, although the underlying molecular mechanisms have not yet been clarified. For example, the production site of ROS that induce PSI photoinhibition has not been identified. Furthermore, it is difficult to analyze PSI photoinhibition, since PSI is highly resistant to photoinhibition compared with PSII (Terashima et al., 1994). In previous studies, specific materials and experimental conditions (such as chilling sensitive plants and cold temperatures) were used to induce PSI photoinhibition in vivo (Terashima et al., 1994; Sonoike et al., 1995). These experimental limitations complicate the elucidation of the molecular mechanisms underlying PSI photoinhibition, as well as the mechanisms that protect PSI from photoinhibition.

Recently, our research group successfully established a method for specifically inducing PSI photoinhibition in the leaves of higher plants, by repetitively illuminating leaves with short-pulse light under dark conditions at room temperature (Sejima et al., 2014; Zivcak et al., 2015a, 2015b). This method was named “repetitive short-pulse” (rSP) treatment. PSI photoinhibition induced by the rSP treatment requires O_2 , and rSP treatment decreases the total content of P700 chlorophyll. Based on these results, we proposed that rSP treatment stimulates the production of ROS in PSI, and that ROS decreases P700 chlorophyll levels. Furthermore, we also observed a decrease in P700 chlorophyll when we used a light intensity similar to that of sunlight ($2,000 \mu E m^{-2} s^{-1}$) in rSP treatment (Sejima et al., 2014). This result suggested that sun flecks stimulate PSI photoinhibition under natural field conditions. Therefore, the elucidation of PSI photoinhibition induced by rSP treatment would improve our understanding of the mechanism underlying photoinhibition in PSI under natural field conditions.

In this study, we focused on the mechanism underlying O_2 -dependent PSI photoinhibition induced by rSP treatment, and we investigated the relationship between the ROS production at PSI and PSI photoinhibition. We found that ROS produced within the thylakoid membranes promoted the degradation of P700 chlorophyll during rSP treatment and induced PSI photoinhibition. Importantly, we found that 1O_2 is produced in PSI, and 1O_2 also stimulates the PSI photoinhibition in addition to O_2^- although many studies are exclusively focused on O_2^- in PSI photoinhibition. On the basis of these results, we discuss the relationship between the ROS production mechanism and PSI photoinhibition.

RESULTS

rSP Treatment Specifically Induces PSI Photoinhibition in Isolated Chloroplasts

First, we applied rSP treatment to isolated chloroplasts, in order to elucidate the mechanism of PSI

photoinhibition, using the method of Sejima et al. (2014) (Fig. 1). In the rSP treatment, the light intensity of the short pulse was set to $20,000 \mu E m^{-2} s^{-1}$, the pulse duration was set at 300 ms, and the illumination frequency was every 10 s under dark conditions. Figure 1A shows the time-course for the change in quantum yields in PSII [Y(II)] and PSI [Y(I)] throughout rSP treatment. Y(II) considerably decreased during rSP treatment in the absence of electron acceptors during the reaction mixture. Y(ND) and Y(NA) represent the extent of photosynthetic electron transport limitation in PSI at the donor and acceptor sides, respectively (Klughammer and Schreiber, 1994). Y(NA) increased throughout rSP treatment (Fig. 1B). In contrast, Y(ND) was not induced during the rSP treatment (Fig. 1C). These results indicate that rSP treatment suppresses the photosynthetic electron transport reaction at the acceptor side of PSI in isolated chloroplasts. On the other hand, Y(II) was less affected during rSP treatment, compared to Y(I) (Fig. 1A). These results are consistent with previous results obtained from intact leaves (Sejima et al., 2014; Zivcak et al., 2015a, 2015b).

We estimated the residual activity of PSII and PSI after rSP treatment, based on the change in the maximum quantum yield of PSII (F_v/F_m) and the total amount of P700 chlorophyll (P_m) before and after rSP treatment (Fig. 1D). The rSP treatment decreased PSII activity to 80%, and PSI activity to 18%. We furthermore analyzed the effect of rSP treatment on PSII, PSI, and the whole-chain photosynthetic electron transport activities. PSII activity was estimated from DMBQ-dependent O_2 evolution rates, and no difference was observed between the control and rSP treatment

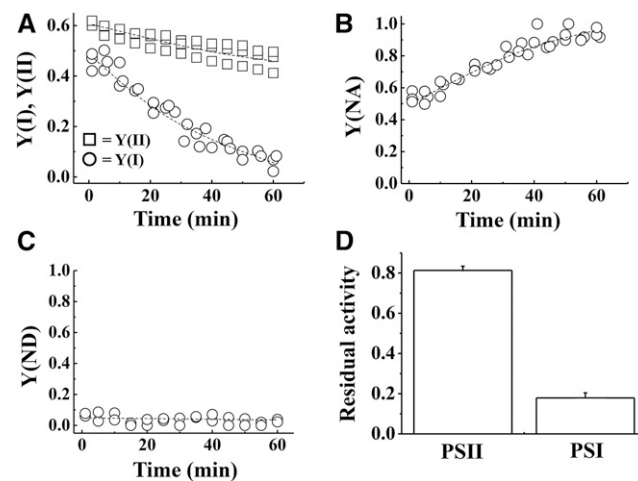


Figure 1. A to C, Time-course analysis of photosynthetic parameters [A, Y(I) and Y(II); B, Y(NA); C, Y(ND)] in isolated chloroplasts. The reaction mixture contained $30 \mu g ml^{-1}$ isolated chloroplasts. Isolated chloroplasts were illuminated every 10 s with a short-pulse (300 ms, $20,000 \mu E m^{-2} s^{-1}$) and for 1 h without AL illumination at $25^\circ C$. Experiments were repeated three times. D, After rSP treatment, the reaction mixture was kept in the dark for 30 min, and F_v/F_m and P_m were measured. Data were normalized to F_v/F_m and P_m before rSP treatment, and are represented as the residual activities of PSII and PSI after rSP treatment. Data are expressed as mean \pm SE of three independent experiments.

samples (Supplemental Fig. S1A). In contrast, PSI and the whole-chain photosynthetic electron activities, estimated from the MV-dependent O_2 absorption rates, were considerably lower in the rSP treatment sample, compared to the control (Supplemental Fig. S1, B and C). These results indicate that rSP treatment induces photoinhibition that is specific to PSI in isolated chloroplasts, which is similar to the results seen in intact leaves (Sejima et al., 2014; Zivcak et al., 2015a, 2015b).

Furthermore, we performed western-blot analyses to confirm whether PSI photoinhibition induced by rSP treatment was accompanied by PSI protein degradation. We found that the amount of PSI core protein PsaA did not decrease after rSP treatment, even though the P700 chlorophyll content was largely decreased (Supplemental Fig. S2). This result indicates that rSP treatment does not stimulate protein degradation in PSI. Likewise, the PSII core protein PsbB did not decrease after rSP treatment (Supplemental Fig. S2). Therefore, rSP treatment-dependent PSI photoinhibition does not involve reductions of PSII and PSI proteins.

Effects of O_2 and Methyl Viologen on PSI Photoinhibition in Isolated Chloroplasts

We analyzed the effect of rSP treatment under anaerobic conditions, in order to reveal whether PSI photoinhibition requires O_2 in isolated chloroplasts in a similar manner to intact leaves. Nitrogen gas was bubbled through the reaction mixture to remove the diluted O_2 , and to create anaerobic conditions. The decrease in $Y(I)$ during rSP treatment was alleviated under anaerobic conditions, compared to aerobic conditions (Fig. 2A). Furthermore, the residual activity of PSI after rSP treatment was significantly higher under anaerobic conditions, compared to aerobic conditions (Fig. 2B). Overall, these results indicate that the presence of O_2 stimulates PSI photoinhibition.

Next, we analyzed the effects of methyl viologen (MV) on the rSP treatment (Fig. 2A). MV accepts electrons from PSI, and the reduced MV subsequently donates electrons to O_2 to produce the superoxide radical (O_2^-) under aerobic conditions (Babbs et al., 1989). In the presence of MV, there was hardly any decrease in $Y(I)$ throughout rSP treatment (Fig. 2A). The residual activity of PSI was significantly higher than when MV was not added (Fig. 2B). In previous studies, PSI photoinhibition was suppressed by limiting photosynthetic electron flow from PSII (Sonoike, 1996; Sejima et al., 2014). That is, accumulation of the oxidized P700 suppresses rSP-dependent PSI photoinhibition (Sejima et al., 2014). Because MV is able to efficiently accept electrons from PSI, this means that the limiting step of the linear electron transport reaction moves from the acceptor side of PSI to plastoquinone (PQ), which is oxidized by cytochrome b_6/f turnover in the presence of MV (Tikhonov, 2013). MV is thought to have two effects on the oxidation of donor side of P700 in PSI: first, the stimulation of ΔpH formation to suppress the oxidation of reduced PQ (PQH₂) by cytochrome b_6/f complex; and

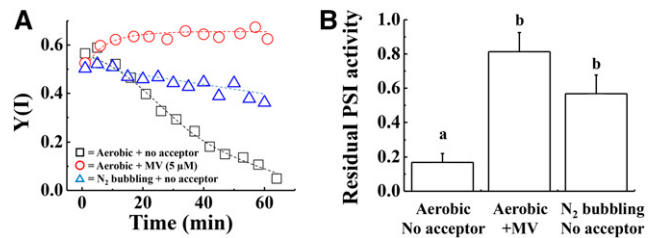


Figure 2. A, Time-course analysis of $Y(I)$ in isolated chloroplasts under different experimental conditions. The reaction mixture contained $30 \mu\text{g ml}^{-1}$ isolated chloroplasts, and the reaction mixture was maintained at 25°C . Black squares indicate $Y(I)$ in the absence of electron acceptors under aerobic conditions. Red circles indicate $Y(I)$ in the presence of MV ($5 \mu\text{M}$) under aerobic conditions. Blue triangles indicate $Y(I)$ in the absence of electron acceptors under anaerobic conditions. Experiments were repeated at least three times, and representative data are shown. B, After rSP treatment, the reaction mixture was kept in the dark for 30 min, and the P_m was measured. Data were normalized to P_m before rSP treatment, and are represented as the residual activity of PSI after rSP treatment. Data are expressed as mean \pm SE of three independent experiments. Different letters above the columns indicate a significant difference between the treatments (Tukey-Kramer HSD test, $P < 0.05$).

second, a stimulation of electron flow from P700 to O_2 to oxidize the photosynthetic electron transport chain.

Effects of the Protonophore Nigericin on PSI Photoinhibition

To examine whether MV forms a ΔpH to suppress PSI photoinhibition during rSP treatment, we applied a protonophore, nigericin, to isolated chloroplasts during rSP treatment. In the absence of MV, the addition of nigericin partially accelerated a decline in $Y(I)$ throughout rSP treatment, compared to in the absence of nigericin (Supplemental Fig. S3). This indicated that a ΔpH was generated during rSP treatment and that electron flow from the donor side of PSI to the acceptor side was limited. This would suppress the reduction of O_2 and the production of ROS in PSI. Indeed, the kinetics of oxidized P700 chlorophyll during short-pulse illumination differed considerably in the absence and presence of nigericin (Fig. 3A). In the presence of nigericin, oxidized P700 chlorophyll was more rapidly reduced during a short-pulse illumination, compared to when nigericin was not added. This result indicates that a ΔpH value was induced during the short-pulse illumination, and this would contribute to the suppression of photosynthetic electron flow from PSII to oxidized P700 chlorophyll. On the other hand, in the presence of MV, the addition of nigericin to the reaction mixture did not lead to a decrease in $Y(I)$ (Supplemental Fig. S3). This result showed that the reduction in PSI photoinhibition by MV could not be due to the stimulation of ΔpH formation. The kinetics of oxidized P700 chlorophyll during a short-pulse illumination in the presence of MV showed that the oxidized state in P700 chlorophyll was maintained during short-pulse illumination, regardless of the addition of nigericin (Fig. 3B).

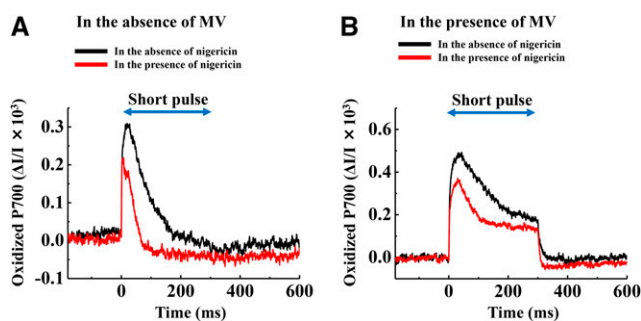


Figure 3. Kinetics of oxidized P700 induced by a short-pulse in the absence of MV (A) and in the presence of MV (B). The data were obtained after rSP treatment was applied for 5 min. The black line shows the condition in the absence of nigericin. The red line shows the condition in the presence of nigericin. Experiments were performed at least three times. Short-pulse was illuminated every 10 s, and average data over three short-pulse illuminations are shown. The reaction mixture contained $30 \mu\text{g ml}^{-1}$ isolated chloroplasts, and the reaction mixture was maintained at 25°C .

This result indicated that MV effectively accepts electrons from PSI, and keeps P700 chlorophyll oxidized during a short-pulse illumination.

Chloroplastic Superoxide Dismutase and Ascorbate Peroxidase Cannot Protect PSI from Photoinhibition Induced by rSP Treatment

Higher plants have superoxide dismutase (SOD) and ascorbate peroxidase (APX) in their chloroplasts, and they help to detoxify ROS (Miyake and Asada, 1992; Asada, 2000). First, we analyzed the effects of rSP treatment on chloroplastic SOD and APX to assess whether chloroplastic SOD and APX activities are maintained in isolated chloroplasts throughout rSP treatment. A comparison of chloroplastic SOD and APX activities before and after rSP treatment showed that their activities did not change (Supplemental Fig. S4). This result indicated that ROS detoxification activities in isolated chloroplasts were maintained throughout rSP treatment. Furthermore, this result indicated that rSP treatment does not stimulate oxidative stress at the chloroplast stroma, because chloroplast APX activities are sensitive to ROS (Miyake and Asada, 1996; Mano et al., 2001). Thus, PSI photoinhibition induced by rSP treatment is probably independent of chloroplastic SOD and APX activities. To confirm this possibility, we added KCN to the reaction mixture to inhibit chloroplastic CuZn-SOD and APX activities, and applied rSP treatment. In spinach chloroplasts, the almost all SOD activities depend on CuZn-SOD (Asada et al., 1973, 1974). The addition of KCN did not affect the decrease in $Y(I)$ during rSP treatment (Fig. 4A). Furthermore, the residual activity of PSI after rSP treatment did not show significant difference between in the absence or the presence of KCN (Fig. 4B). These results suggest that chloroplastic CuZn-SOD and APX do not protect PSI from photoinhibition induced by rSP treatment.

Surprisingly, we found that, even though KCN was present, MV suppressed PSI photoinhibition, when the production of O_2^- was enhanced during rSP treatment (Supplemental Fig. S5). From these facts, PSI photoinhibition would not be related to the production rate of ROS, and the scavenging of ROS outside the thylakoid membranes. Next, we studied the molecular species of ROS that is produced during rSP treatment.

Both O_2^- and $^1\text{O}_2$ Contribute to PSI Photoinhibition Induced by rSP Treatment

PSI is a major production site of ROS, and O_2^- has been generally recognized as a main ROS to be produced at PSI (Takahashi and Asada, 1982). However, recent reports provide evidence that PSI also produces $^1\text{O}_2$, which is related to PSI photoinhibition (Cazzaniga et al., 2012; Rutherford et al., 2012). Furthermore, we showed that inactivation of APX and CuZn-SOD does not affect PSI photoinhibition (Fig. 4). This might indicate that PSI photoinhibition was induced by $^1\text{O}_2$ but not O_2^- during rSP treatment. To examine the possibility whether $^1\text{O}_2$ relates to PSI photoinhibition induced by rSP treatment, we added a soluble α -tocopherol analog, 2, 2', 5, 7, 8-pentamethyl-6-chromanol (PMC), which is an amphipathic $^1\text{O}_2$ scavenger, to the reaction mixture, and conducted rSP treatment (Fryer, 1992; Grams and Inglett, 1972; Munné-Bosch, 2005). The addition of PMC did not affect the early decrease in $Y(I)$ during rSP treatment (Fig. 5A). In contrast, PMC alleviated the late decrease in $Y(I)$, compared to that seen in the control (Fig. 5A). The residual activity of PSI after rSP treatment for 1 h revealed that the addition of PMC significantly alleviated the inactivation of PSI, compared to control (Fig. 5B). In contrast, the residual activity of PSII was not different in the absence and the presence of PMC (Fig. 5C).

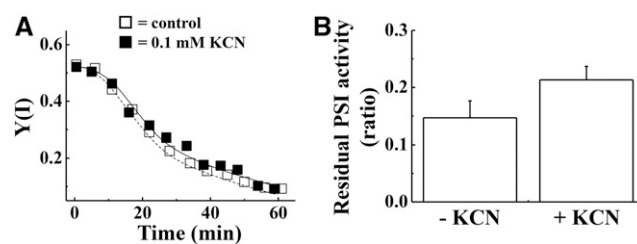


Figure 4. Effect of KCN on PSI photoinhibition induced by rSP treatment. The reaction mixture contained $30 \mu\text{g ml}^{-1}$ isolated chloroplasts, and the reaction mixture was maintained at 25°C . A, Time-course analysis of $Y(I)$ in isolated chloroplasts in the absence (white squares) and presence of KCN (0.1 mM; black squares). Experiments were repeated at least three times, and representative data are shown. B, After rSP treatment, the reaction mixture was kept in the dark for 30 min, and the P_m was measured. Data were normalized to the P_m before rSP treatment, and the residual activity of PSI after rSP treatment was calculated. Data are expressed as mean \pm SE of three independent experiments. Student's *t* test revealed that there is no significant difference in the residual activities in between the absence and the presence of KCN ($P > 0.1$).

To examine whether PMC acts as an electron acceptor like MV, we analyzed the kinetics of oxidized P700 chlorophyll during the short-pulse illumination. We found that PMC did not affect the kinetics of oxidized P700 chlorophyll during a short-pulse illumination (Supplemental Fig. S6). Furthermore, we compared the O_2 absorption rate and Y(II) in the absence and presence of PMC using an O_2 -electrode and chlorophyll fluorescence. Under continuous actinic light (AL) illumination ($500 \mu E m^{-2} s^{-1}$), PMC did not affect O_2 absorption rate and Y(II) in isolated chloroplasts (Supplemental Table S1). From these results, we revealed that PMC does not act as an electron acceptor. Furthermore, doubling the PMC concentration from $5 \mu M$ to $10 \mu M$ did not affect the residual activity of PSI after rSP treatment for 1 h [PSI residual activity = 0.35 ± 0.06 ($n = 3$)]. This result indicated that $5 \mu M$ PMC is enough to protect PSI from photoinhibition induced by 1O_2 .

DISCUSSION

In this study, we aimed to reveal the mechanisms by which rSP treatment induces photoinhibition specific to PSI by using isolated chloroplasts. We observed that rSP treatment induces PSI-specific photoinhibition in isolated chloroplasts when O_2 was present, in a similar manner to leaves (Sejima et al., 2014). In addition, rSP treatment caused a limitation of the photosynthetic electron transport reaction at the acceptor side in PSI (Fig. 1; Supplemental Fig. S1). These results suggest that

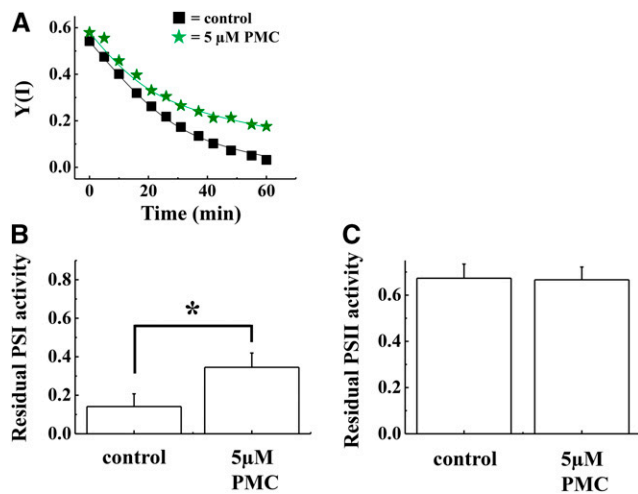


Figure 5. Effect of PMC on PSI photoinhibition induced by rSP treatment. The reaction mixture contains $30 \mu g ml^{-1}$ isolated chloroplasts, and the reaction mixture was maintained at $25^\circ C$. A, Time-course analysis of Y(II) in isolated chloroplasts in the absence (black squares) and presence of PMC ($5 \mu M$; green stars). Experiments were repeated at least three times, and representative data are shown. B and C, After rSP treatment, the reaction mixture was kept in the dark for 30 min, and the P_m and F_v/F_m values were measured. Data were normalized to the P_m and F_v/F_m values before rSP treatment, and the residual activity of PSI (B) and PSII (C) after rSP treatment were calculated. Data are expressed as mean \pm SE of three independent experiments.

rSP treatment critically affected chloroplasts, but other organelles were not involved in PSI photoinhibition.

We propose that PSI photoinhibition is triggered by ROS produced within the thylakoid membranes. Although several components of ROS production have been proposed in previous reports, the ROS production mechanism in PSI has not been clarified (Misra and Fridovich, 1971; Miyake et al., 1998; Asada, 2000; Voss et al., 2011; Kozuleva and Ivanov, 2010). ROS are known to trigger PSI photoinhibition and to degrade its components because PSI photoinhibition requires O_2 (Allahverdiyeva et al., 2005; Sonoike and Terashima, 1994; Sonoike, 1996). However, the production site of the ROS that trigger PSI photoinhibition has been not clarified. In our study, we observed that the addition of MV to isolated chloroplasts greatly alleviates PSI photoinhibition induced by rSP treatment (Fig. 2). This result is consistent with a previous report on thylakoid membranes (Sonoike, 1996). A relationship between the suppression of PSI photoinhibition and oxidation of P700 in PSI would clarify the molecular mechanism of MV to alleviate PSI photoinhibition. We have thought that an increase in $P700^+$ decreases the ratio of the ground state P700 and excited P700 against the total P700 amount, of which both can donate electrons to O_2 to produce O_2^- in the thylakoid membranes. However, even in the presence of KCN or nigericin, the presence of MV suppressed rSP-treatment-dependent PSI photoinhibition (Supplemental Figs. S3 and S5). These situations should stimulate the production of ROS in the chloroplast thylakoid membranes and stroma. Based on these results, the production rate of ROS, and the quantity of ROS should not relate to the PSI photoinhibition in chloroplasts. Therefore, we suggest that the production site of ROS, rather than the quantity of ROS, is important for causing PSI photoinhibition.

MV accepts electrons from PSI and stimulates the production of O_2^- through their radical form (MV; Babbs et al., 1989). Then, where is ROS produced by MV? According to Kozuleva et al. (2014), the midpoint redox potential (E_m) of O_2/O_2^- in the lipid phase is a lower value ($-500 mV / -600 mV$ vs. normal hydrogen electrode), compared to the E_m value of O_2/O_2^- in the aqueous phase. Since MV accepts electrons from electron carriers in PSI [F_x , F_A , or F_B ; their E_m values are higher than the E_m value of O_2/O_2^- in the lipid phase (Kozuleva et al., 2014)], MV would have a higher E_m value than the E_m value of O_2/O_2^- in the lipid phase (Michaelis and Hill, 1933; Stombaugh et al., 1976). Based on the latter and thermodynamics, MV effectively reduces O_2 to produce O_2^- at the stromal region in chloroplasts, although the reduction of O_2 within the thylakoid membranes might be insufficient. Furthermore, we used $5 \mu M$ MV in the rSP treatment. Under these conditions, MV reacts with F_A/F_B at the stromal side in PSI (Sonoike and Terashima, 1994). Based on this report, MV would stimulate the production of ROS in the chloroplast stroma, and we suggest ROS produced in the stromal region does not contribute to PSI photoinhibition. This suggestion is also supported by the

result that chloroplastic CuZn-SOD and APX activities were independent of PSI photoinhibition (Fig. 4; Supplemental Figs. S4 and S5).

In addition to the above considerations, we also suggest the possibility that MV-dependent oxidation of the electron transport chain alleviated PSI photoinhibition. A recent study showed that P700 in PSI is oxidized under electron sink-limitation conditions in the cyanobacteria *Synechococcus elongatus* PCC 7942 (Shaku et al., 2015). Shaku et al. (2015) demonstrated that an oxidized high-potential chain, including cytochrome *f* and plastocyanin-scavenged O_2^- , was produced in thylakoid membranes. This scavenging rate was comparable to the electron flow rate in linear electron transport (Tanaka et al., 1978; Takahashi et al., 1980). We observed that MV successfully oxidized the P700 during a short-pulse illumination (Fig. 3). Therefore, the stimulated oxidation of photosynthetic electron transport chain by MV could scavenge O_2^- , even though the production of ROS was stimulated (Supplemental Fig. S5).

In leaves, PSI photoinhibition induced by rSP treatment is suppressed under continuous AL illumination, compared to dark conditions (Sejima et al., 2014). This is because constant AL illumination stimulates the oxidation of PSI by activating the electron sink in photosynthesis (like the Calvin cycle and photorespiration) and the formation of ΔpH across the thylakoid membrane (Zaks et al., 2012). Indeed, we observed that ΔpH and electron donation from PSI to MV protect PSI photoinhibition in isolated chloroplasts (Fig. 3). In the study by Sejima et al. (2014), PSI photoinhibition was suppressed, in accordance with the induction of Y(ND) under AL illumination. Based on this result, in leaves, the formation of a large ΔpH across the thylakoid membranes is more critical for the protection of PSI photoinhibition, compared to the consumption of electrons in the electron sink in photosynthesis. Therefore, these results and those by Sejima et al. (2014) show that rSP treatment under dark conditions promotes the reduction of electron carriers in PSI without activating the electron sink and forming a large ΔpH , which suppresses the photosynthetic electron transport reaction from PSII to PSI in vivo. Accordingly, PSI photoinhibition would be triggered by the production of ROS within the thylakoid membranes in vivo, in the same way as in isolated chloroplasts.

We propose that PSI photoinhibition proceeds in two different ways by the different kinds of ROS produced within the thylakoid membrane (Fig. 6). First, the production of O_2^- and $OH\cdot$ at a secondary electron acceptor in PSI induces PSI photoinhibition (Takahashi and Asada 1988; Sonoike et al., 1995). PSI has four electron acceptors (A_0 as a primary acceptor, and A_1 , F_X , and F_A/F_B as secondary acceptors). It has been suggested that the ROS production site in PSI is within the thylakoid membranes and that the secondary acceptor, A_1 , is a major contributor to the Mehler reaction in vivo (Kozuleva and Ivanov, 2010; Kozuleva et al., 2014). Two types of A_1 , which have different E_m values (A_{1a} , -671 mV; A_{1b} , -844 mV), exist in PSI, and A_{1a} and A_{1b} are

coupled with PsaA and PsaB, respectively (Rutherford et al., 2012). The degradation of PsaB is more promoted during PSI photoinhibition, compared to the degradation of PsaA (Sonoike, 1996; Sonoike et al., 1997). Therefore, A_{1b} , which has a lower E_m value than A_{1a} , might be a primary ROS production site in PSI, and O_2^- produced at A_{1b} could trigger PSI photoinhibition (Sonoike and Terashima, 1994). We suggest that ROS production and the oxidative attacks by ROS in PSI occur within the thylakoid membranes. Therefore, chloroplastic SOD and APX cannot protect PSI from its photoinhibition (Fig. 4; Supplemental Fig. S4). In fact, Takahashi and Asada (1988) reported that O_2 reduction detected by O_2^- -dependent cytochrome *c* reduction was stimulated in thylakoid membranes that were disintegrated by detergent. This means that the O_2 reduction site does not exist on the thylakoid membrane surface. Furthermore, they revealed that high concentrations of NH_4Cl , which induced the protonation of O_2^- , stimulated the production of H_2O_2 in thylakoid membranes, and simultaneously NH_4Cl suppressed the O_2^- -dependent cytochrome *c* reduction (Takahashi and Asada, 1988). From these observations, they concluded that O_2^- was produced in the aprotic region of the thylakoid membranes, that is, interior regions of the thylakoid membrane. Their conclusion is consistent with our observations.

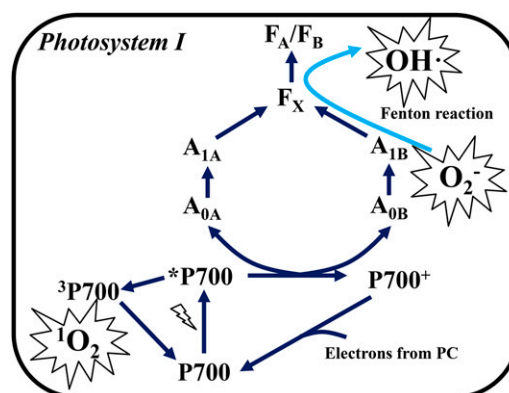


Figure 6. The scheme of ROS production and photoinhibition in PSI. When electron carriers are highly reduced in PSI, the production of O_2^- is stimulated at the A_1 site (Kozuleva and Ivanov, 2010; Kozuleva et al., 2014). O_2^- either directly attacks the peripheral component, or is converted to $OH\cdot$ via the Fenton reaction between the iron-sulfur center in F_X , F_A , or F_B in the diffusing process (Inoue et al., 1986; Sonoike et al., 1995; Takahashi and Asada, 1988). The limitation of electron transfer from P700 to downstream electron acceptors would suppress the charge separation between P700 and chlorophyll A_0 (Shuvalov et al., 1986). Furthermore, the production of the P700 triplet-state (3P700) is initiated (Rutherford et al., 2012; Shuvalov et al., 1986). 3P700 reacts with O_2 to produce singlet O_2 (1O_2). 1O_2 attacks the peripheral component in PSI, and precedes PSI photoinhibition (Cazzaniga et al., 2012). These reactions proceed within the thylakoid membranes. Therefore, a decrease in P700 chlorophyll occurs, but protein degradation does not accompany PSI photoinhibition. Blue arrows indicate the energy transfer pathway in PSI, and the light-blue arrow indicates the diffusion of O_2^- in the thylakoid membranes (Takahashi and Asada, 1988).

Second, the production of $^1\text{O}_2$ by charge recombination in PSI proceeds further PSI photoinhibition (Fig. 6). We revealed that PMC, which acts as a $^1\text{O}_2$ scavenger, suppressed the decrease in Y(I) in isolated chloroplasts, and PMC significantly protected PSI activity after rSP treatment for 1 h, compared to the control (Fig. 5). In a similar way to P680 chlorophyll, the transient state of the excited P700 chlorophyll in PSI is de-excited to the triplet state ($^3\text{P700}$) through charge recombination, and then the $^3\text{P700}$ reacts with O_2 to produce $^1\text{O}_2$ (Shuvalov et al., 1986; Rutherford et al., 2012). In a previous study, it has been reported that $^1\text{O}_2$ is not produced in PSI under photoinhibitory conditions for PSII (Hideg and Vass, 1995). However, in that study, the decrease in PSI activity was not observed under photoinhibitory conditions although PSII activity was successfully impaired (Hideg and Vass, 1995). This might indicate that electron carriers in PSI did not become reduced to produce ROS in PSI under photoinhibitory conditions for PSII. In contrast, Cazzaniga et al. (2012) reported that $^1\text{O}_2$ is produced in the PSI-LHCI complex isolated from *Arabidopsis* (*Arabidopsis thaliana*). Furthermore, they also reported that the *Arabidopsis szl1* mutant, which contains less β -carotene than the wild-type, is susceptible to PSI photoinhibition under high-light and low-temperature conditions. This means that $^1\text{O}_2$ produced in PSI caused PSI photoinhibition because β -carotene is a major $^1\text{O}_2$ quencher (Telfer, 2014). Therefore, we suggest that $^1\text{O}_2$ is actually produced in PSI, and $^1\text{O}_2$ is involved in PSI photoinhibition in higher plants. In PSI photoinhibition, $F_{A'}$, $F_{B'}$, and F_X were primarily destroyed and they lost electron transport activity (Inoue et al., 1986; Sonoike et al., 1995). Based on these reports, O_2^- produced at A_1 would diffuse in the thylakoid membrane and react with the [Fe-S] cluster in F_X , $F_{A'}$, or $F_{B'}$. Furthermore, $\text{OH}\cdot$ produced from O_2^- through the Fenton reaction would then inactivate their electron transfer activities (Takahashi and Asada, 1988; Sonoike et al., 1997). When the electron transport reaction in PSI is limited after the destruction of F_X , $F_{A'}$, and $F_{B'}$, the charge separation of P700 chlorophyll would be suppressed, and then the charge recombination between P700^+ and A_0 , or A_1 could be further accelerated (Shuvalov et al., 1986; Kozuleva and Ivanov, 2010).

We observed that PSI core protein content did not decrease after PSI photoinhibition, even though P700 chlorophyll content and PSI activity largely decreased in isolated chloroplasts as well as in leaves (Supplemental Fig. S2). Previous studies also showed that the decrease in PSI activity and P700 chlorophyll content does not correlate with the decrease in PSI protein content after PSI photoinhibition (Sonoike et al., 1997; Tjus et al., 1998; Zhang and Scheller, 2004). Therefore, these results indicate that PSI protein degradation is not the first step of PSI photoinhibition. These results support the idea that ROS produced within the thylakoid membrane specifically target electron transport carriers in PSI, and hence precede PSI photoinhibition.

We have shown that rSP treatment stimulated specific PSI photoinhibition *in vitro*. Furthermore, the kinetics of Y(I) during rSP treatment were quite similar in

the leaves and isolated chloroplasts (Fig. 1A; Sejima et al., 2014; Zivcak et al., 2015a, 2015b). To date, there has not been a common method to cause PSI photoinhibition *in vivo* and *in vitro*. This means that the rSP treatment improves PSI photoinhibition analysis efficiency. Because PSI photoinhibition is a severe phenomenon, reducing plant fitness, the identification of the protection mechanism against PSI photoinhibition would contribute to the improvement of plant fitness. We suggest that the rSP treatment should make it possible to uncover the protection mechanism against PSI photoinhibition through a forward or reverse genetic approach, such as the selection of mutagenized plants that have a higher susceptibility or tolerance to PSI photoinhibition.

MATERIALS AND METHODS

Isolation of Intact Chloroplasts from Spinach Leaves

Intact chloroplasts were isolated from spinach (*Spinacia oleracea*) leaves purchased from a local market and purified by a Percoll density gradient centrifugation, as described previously by Takagi et al. (2012). The isolated chloroplasts were suspended in a reaction mixture of HEPES-KOH [50 mM 4-(2-hydroxyethyl) piperazine-1-ethansulfonic acid-potassium hydrate], pH 7.6; 0.33 M sorbitol; 10 mM NaCl (sodium chloride); 1 mM MgCl_2 (magnesium chloride); 2 mM EDTA; and 0.5 mM KH_2PO_4 (monopotassium phosphate). The intactness of the purified chloroplasts was determined with the ferricyanide method (Heber and Santarius, 1970), and 85% to 95% of the intact chloroplasts were used in our experiment. The chloroplast chlorophyll content was determined as described previously by Arnon (1949).

Oxygen Exchange Analysis in Isolated Chloroplasts

A chloroplast suspension that had either been incubated in the dark for 1 h or subjected to repetitive short pulse (rSP) treatment for 1 h was placed in an oxygen electrode cuvette (DW 2/2; Hansatech, King's Lynn, UK) equipped with actinic red light (AL; >640 nm). Temperature was maintained at 25°C and controlled by circulating temperature-controlled water through the water jacket. The PSII, PSI, and whole-chain activities were measured as described by Miyake and Okamura (2003). PSII activity in the chloroplasts was determined by the O_2 evolution rate. To achieve this, 2, 6-dimethyl benzoquinone (500 μM) and nigericin (0.5 μM) were added to the chloroplast suspension, and illuminated with AL. PSI activity was determined by the O_2 absorption rate. For this measurement, dichlorophenolindophenol (70 μM), nigericin (0.5 μM), KCN (0.1 mM), ascorbate (1 mM), methyl viologen (100 μM), and 3-(3,4-dichlorophenyl)-1,1-dimethylurea (10 μM) were added to the chloroplast suspension, and illuminated with AL. Whole-chain activity of the photosynthetic electron transport reaction was determined by the O_2 absorption rate. For this measurement, nigericin (0.5 μM), KCN (0.1 mM), and methyl viologen (100 μM) were also added to the chloroplast suspension, and illuminated with AL.

Enzyme Assay of Chloroplastic SOD and APX

Chloroplastic APX activities were determined in a reaction mixture containing isolated chloroplasts (60 μg Chl), 50 mM potassium phosphate (pH 7.0), 0.5 mM ascorbate, and 5 mM H_2O_2 (Miyake and Asada, 1992). APX activities were determined by measuring the H_2O_2 -dependent oxidation of ascorbate. The oxidation of ascorbate was monitored by the decrease in A_{290} (the absorption coefficient was 2.8 $\text{mm}^{-1} \text{cm}^{-1}$). Chloroplastic SOD activities were determined using the xanthine oxidase/cytochrome *c* method (McCord and Fridovich, 1969). Chloroplastic SOD activities were measured in a reaction mixture containing isolated chloroplasts (60 μg Chl), 300 mM potassium phosphate (pH 8.0), 60 μM cytochrome *c*, 0.3 mM xanthine, and 120 mU xanthine oxidase. The reduction of cytochrome *c* was measured at 550 nm.

Measurement of Chlorophyll Fluorescence and P700^+

Chlorophyll fluorescence and P700^+ were simultaneously measured with a Dual-PAM-100 measuring system (Heinz Walz, Effeltrich, Germany). The

reaction mixture was maintained at 25°C by temperature control unit US-T (Heintz Walz). The chlorophyll fluorescence parameters were calculated according to Baker (2008): maximum quantum efficiency of PSII photochemistry, $F_v/F_m = (F_m - F_o)/F_m$; quantum yield of photochemical energy conversion in PSII, $Y(II) = (F_m' - F_s)/F_m'$; F_o , minimum fluorescence yield; F_m , maximum fluorescence yield; and F_s , steady-state fluorescence yield. Measuring light ($0.1 \mu\text{E m}^{-2} \text{s}^{-1}$) and saturated pulse ($20,000 \mu\text{E m}^{-2} \text{s}^{-1}$, 300 ms) were applied to determine F_o and F_m . The oxidation-reduction state of P700⁺ was determined according to the methods of Klughammer and Schreiber (1994) as follows: quantum yield of photochemical energy in PSI, $Y(I) = (P_m' - P)/P_m'$; quantum yield of nonphotochemical quenching due to the acceptor side limitation, $Y(NA) = (P_m - P_m')/P_m'$; and quantum yield of nonphotochemical quenching due to the donor side limitation, $Y(ND) = P/P_m'$. The maximum oxidation level of P700 chlorophyll (P_m) was obtained by a saturated pulse under far-red light, which reflected the maximum amount of photooxidized P700 chlorophyll. The parameter P reflects the steady-state oxidation level of P700⁺, and P_m' was obtained by a saturated pulse at a steady state.

rSP Treatment

rSP treatment was applied to chloroplasts that had been isolated from spinach leaves, as described previously by Sejima et al. (2014). Isolated chloroplasts were illuminated with short pulses ($20,000 \mu\text{E m}^{-2} \text{s}^{-1}$, 300 ms) every 10 s in the absence of AL for 1 h under the experimental conditions indicated in the figure legends. During rSP treatment, the reaction mixture containing isolated chloroplasts was maintained at 25°C by temperature control unit US-T (Heintz Walz). After rSP treatment, the photosynthetic parameters of PSII and PSI activity were determined as described in the figure legends.

Sodium Dodecyl Sulfate-Polyacrylamide Gel Electrophoresis

For the protein analysis, isolated chloroplasts suspended in the reaction mixture were collected by the centrifugation at 5000g for 1 min at 4°C. After the centrifugation, the isolated chloroplasts were resuspended in the protein-solubilizing solution [80 mM Tris-HCl (pH 6.8), 1 mM EDTA, 5% glycerol (v/v), 2% N-octyl-β-D-glycoside (w/v)]. The isolated chloroplasts were electrophoresed on 12.5% (w/v) sodium dodecyl sulfate-polyacrylamide gels containing 8 M urea, as described by Laemmli (1970).

Western Blotting

The proteins were separated by sodium dodecyl sulfate-polyacrylamide gel electrophoresis, transferred onto a polyvinylidene difluoride (PVDF) membrane (Merck Millipore, Billerica, MA), and then blocked with the commercially available Blocking One reagent (Nakalai Tesque, Kyoto, Japan) for 30 min at room temperature (25°C). The PVDF membrane was subsequently incubated with a specific peptide antibody for 1 h at room temperature (25°C). The PVDF membrane was washed three times with TBS Tween buffer [10 mM Tris-HCl (pH 7.4), 0.14 M NaCl, 0.1% (v/v) Tween 20] and incubated with ECL peroxidase-labeled secondary anti-rabbit antibody (GE Healthcare, Buckinghamshire, UK) for 1 h at room temperature (25°C). The PVDF membrane was washed three times with TBS TWEEN buffer. Proteins were detected with an alkaline phosphatase labeling kit. The protein content was quantified by ImageJ version 1.49 (National Institutes of Health, Bethesda, MD). The protein content after rSP treatment was expressed by relative value, compared to control sample.

Statistical Analysis

All data were expressed as mean ± SE of at least three independent analyses. We used ANOVAs, Student's *t* tests, and Tukey-Kramer HSD tests to detect differences. All statistical analyses were performed using Microsoft Excel 2010 (Microsoft, Redmond, WA) and JMP8 (SAS Institute, Tokyo, Japan).

Supplemental Data

The following supplemental materials are available.

Supplemental Figure S1. The light response of PSII, PSI, and whole-chain photosynthetic electron transport activity in isolated chloroplasts before and after rSP treatment.

Supplemental Figure S2. The comparison of PSII and PSI core protein content in isolated chloroplast between before and after rSP treatment.

Supplemental Figure S3. The effect of nigericin on rSP treatment in the absence and presence of MV.

Supplemental Figure S4. The change in SOD and APX activities before and after rSP treatment.

Supplemental Figure S5. The effect of KCN on short-pulse treatment in the absence and presence of MV.

Supplemental Figure S6. The kinetics of oxidized P700 induced by a short-pulse in the absence of and in the presence of PMC.

Supplemental Table S1. Oxygen evolution rate and Y(II) in the absence and in the presence of PMC.

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LITERATURE CITED

- Allahverdiyeva Y, Mamedov F, Mäenpää P, Vass I, Aro EM (2005) Modulation of photosynthetic electron transport in the absence of terminal electron acceptors: characterization of the *rbcL* deletion mutant of tobacco. *Biochim Biophys Acta* 1709: 69–83
- Arnon DI (1949) Copper enzymes in isolated chloroplasts. Polyphenoloxidase in *Beta vulgaris*. *Plant Physiol* 24: 1–15
- Aro EM, Virgin I, Andersson B (1993) Photoinhibition of Photosystem II. Inactivation, protein damage and turnover. *Biochim Biophys Acta* 1143: 113–134
- Asada K (2000) The water-water cycle as alternative photon and electron sinks. *Philos Trans R Soc Lond B Biol Sci* 355: 1419–1431
- Asada K, Takahashi M (1987) Production and scavenging of active oxygen in photosynthesis. In DJ Kyle, CB Osmond, CJ Arntzen, eds, *Photoinhibition*. Elsevier Science, Amsterdam, The Netherlands, pp 227–287
- Asada K, Takahashi M, Nagate M (1974) Assay and inhibitors of spinach superoxide dismutase. *Agric Biol Chem* 38: 471–473
- Asada K, Urano M, Takahashi M (1973) Subcellular location of superoxide dismutase in spinach leaves and preparation and properties of crystalline spinach superoxide dismutase. *Eur J Biochem* 36: 257–266
- Babbs CF, Pham JA, Coolbaugh RC (1989) Lethal hydroxyl radical production in paraquat-treated plants. *Plant Physiol* 90: 1267–1270
- Baker NR (2008) Chlorophyll fluorescence: a probe of photosynthesis *in vivo*. *Annu Rev Plant Biol* 59: 89–113
- Bondarava N, Gross CM, Mubarakshina M, Golecki JR, Johnson GN, Krieger-Liszka A (2010) Putative function of cytochrome *b559* as a plastoquinol oxidase. *Physiol Plant* 138: 463–473
- Cazzaniga S, Li Z, Niyogi KK, Bassi R, Dall'Osto L (2012) The *Arabidopsis* *szl1* mutant reveals a critical role of β-carotene in photosystem I photoprotection. *Plant Physiol* 159: 1745–1758
- DalCorso G, Pesaresi P, Masiero S, Aseeva E, Schünemann D, Finazzi G, Joliet P, Barbato R, Leister D (2008) A complex containing PGRL1 and PGR5 is involved in the switch between linear and cyclic electron flow in *Arabidopsis*. *Cell* 132: 273–285
- Fryer MJ (1992) The antioxidant effects of thylakoid vitamin E (α-tocopherol). *Plant Cell Environ* 15: 381–392
- Gill SS, Tuteja N (2010) Reactive oxygen species and antioxidant machinery in abiotic stress tolerance in crop plants. *Plant Physiol Biochem* 48: 909–930
- Grams G, Inglett GE (1972) Sensitized photooxidation of α-tocopherol and of 2, 2, 5, 7, 8-pentamethyl-6-chromanol in ethyl acetate. *Lipids* 7: 442–444
- Grieco M, Tikkanen M, Paakkanen V, Kangasjärvi S, Aro EM (2012) Steady-state phosphorylation of light-harvesting complex II proteins preserves photosystem I under fluctuating white light. *Plant Physiol* 160: 1896–1910
- Heber U, Santarius KA (1970) Direct and indirect transfer of ATP and ADP across the chloroplast envelope. *Z Naturforsch B* 25: 718–728
- Hideg E, Kós PB, Vass I (2007) Photosystem II damage induced by chemically generated singlet oxygen in tobacco leaves. *Physiol Plant* 131: 33–40

- Hideg É, Vass I (1995) Singlet oxygen is not produced in photosystem I under photoinhibitory conditions. *Photochem Photobiol* **62**: 949–952
- Inoue K, Sakurai H, Hiyama T (1986) Photoinactivation sites of photosystem I in isolated chloroplasts. *Plant Cell Physiol* **27**: 961–968
- Klughammer C, Schreiber U (1994) An improved method, using saturating light pulses, for the determination of photosystem I quantum yield via P700⁺-absorbance changes at 830 nm. *Planta* **192**: 261–268
- Kojima K, Oshita M, Nanjo Y, Kasai K, Tozawa Y, Hayashi H, Nishiyama Y (2007) Oxidation of elongation factor G inhibits the synthesis of the D1 protein of photosystem II. *Mol Microbiol* **65**: 936–947
- Kono M, Noguchi K, Terashima I (2014) Roles of the cyclic electron flow around PSI (CEF-PSI) and O₂-dependent alternative pathways in regulation of the photosynthetic electron flow in short-term fluctuating light in *Arabidopsis thaliana*. *Plant Cell Physiol* **55**: 990–1004
- Kozuleva MA, Ivanov BN (2010) Evaluation of the participation of ferredoxin in oxygen reduction in the photosynthetic electron transport chain of isolated pea thylakoids. *Photosynth Res* **105**: 51–61
- Kozuleva MA, Petrova AA, Mamedov MD, Semenov AY, Ivanov BN (2014) O₂ reduction by photosystem I involves phyloquinone under steady-state illumination. *FEBS Lett* **588**: 4364–4368
- Krieger-Liszakay A (2005) Singlet oxygen production in photosynthesis. *J Exp Bot* **56**: 337–346
- Kühlheim C, Ågren J, Jansson S (2002) Rapid regulation of light harvesting and plant fitness in the field. *Science* **297**: 91–93
- Laemmli UK (1970) Cleavage of structural proteins during the assembly of the head of bacteriophage T4. *Nature* **227**: 680–685
- Mano J, Ohno C, Domae Y, Asada K (2001) Chloroplastic ascorbate peroxidase is the primary target of methylviologen-induced photooxidative stress in spinach leaves: its relevance to monodehydroascorbate radical detected with *in vivo* ESR. *Biochim Biophys Acta* **1504**: 275–287
- McCord JM, Fridovich I (1969) Superoxide dismutase. An enzymic function for erythrocyte cytochrome c. *J Biol Chem* **244**: 6049–6055
- Melis A (1999) Photosystem-II damage and repair cycle in chloroplasts: what modulates the rate of photodamage? *Trends Plant Sci* **4**: 130–135
- Michaelis L, Hill ES (1933) The viologen indicators. *J Gen Physiol* **16**: 859–873
- Misra HP, Fridovich I (1971) The generation of superoxide radical during the autoxidation of ferredoxins. *J Biol Chem* **246**: 6886–6890
- Miyake C, Asada K (1992) Thylakoid-bound ascorbate peroxidase in spinach chloroplasts and photoreduction of its primary oxidation product monodehydroascorbate radicals in thylakoids. *Plant Cell Physiol* **33**: 541–553
- Miyake C, Asada K (1996) Inactivation mechanism of ascorbate peroxidase at low concentrations of ascorbate; hydrogen peroxide decomposes compound I of ascorbate peroxidase. *Plant Cell Physiol* **37**: 423–430
- Miyake C, Okamura M (2003) Cyclic electron flow within PSII protects PSII from its photoinhibition in thylakoid membranes from spinach chloroplasts. *Plant Cell Physiol* **44**: 457–462
- Miyake C, Schreiber U, Hormann H, Sano S, Asada K (1998) The FAD-enzyme monodehydroascorbate radical reductase mediates photoproduction of superoxide radicals in spinach thylakoid membranes. *Plant Cell Physiol* **39**: 821–829
- Munekage YN, Genty B, Peltier G (2008) Effect of PGR5 impairment on photosynthesis and growth in *Arabidopsis thaliana*. *Plant Cell Physiol* **49**: 1688–1698
- Munné-Bosch S (2005) The role of α -tocopherol in plant stress tolerance. *J Plant Physiol* **162**: 743–748
- Nishiyama Y, Allakhverdiev SI, Murata N (2011) Protein synthesis is the primary target of reactive oxygen species in the photoinhibition of photosystem II. *Physiol Plant* **142**: 35–46
- Oelze ML, Vogel MO, Alsharafa K, Kahmann U, Viehhauser A, Maurino VG, Dietz KJ (2012) Efficient acclimation of the chloroplast antioxidant defence of *Arabidopsis thaliana* leaves in response to a 10- or 100-fold light increment and the possible involvement of retrograde signals. *J Exp Bot* **63**: 1297–1313
- Rutherford AW, Osyczka A, Rappaport F (2012) Back-reactions, short-circuits, leaks and other energy wasteful reactions in biological electron transfer: redox tuning to survive life in O₂. *FEBS Lett* **586**: 603–616
- Sejima T, Takagi D, Fukayama H, Makino A, Miyake C (2014) Repetitive short-pulse light mainly inactivates photosystem I in sunflower leaves. *Plant Cell Physiol* **55**: 1184–1193
- Shaku K, Shimakawa G, Hashiguchi M, Miyake C (2015) Reduction-induced suppression of electron flow (RISE) in the photosynthetic electron transport system of *Synechococcus elongatus* PCC 7942. *Plant Cell Physiol* <http://dx.doi.org/10.1093/pcp/pcv198>
- Shuvalov VA, Nuijs AM, van Gorkom HJ, Smit HWJ, Duysens LNM (1986) Picosecond absorbance changes upon selective excitation of the primary electron donor P-700 in photosystem I. *Biochim Biophys Acta Bioenerg* **850**: 319–323
- Sonoike K (1996) Degradation of *psaB* gene product, the reaction center subunit of photosystem I, is caused during photoinhibition of photosystem I: possible involvement of active oxygen species. *Plant Sci* **115**: 157–164
- Sonoike K (2011) Photoinhibition of photosystem I. *Physiol Plant* **142**: 56–64
- Sonoike K (1995) Selective photoinhibition of photosystem I in isolated thylakoid membranes from cucumber and spinach. *Plant Cell Physiol* **36**: 825–830
- Sonoike K, Kamo M, Hihara Y, Hiyama T, Enami I (1997) The mechanism of the degradation of *psaB* gene product, one of the photosynthetic reaction center subunits of photosystem I, upon photoinhibition. *Photosynth Res* **53**: 55–63
- Sonoike K, Terashima I (1994) Mechanism of photosystem-I photoinhibition in leaves of *Cucumis sativus* L. *Planta* **194**: 287–293
- Sonoike K, Terashima I, Iwaki M, Itoh S (1995) Destruction of photosystem I iron-sulfur centers in leaves of *Cucumis sativus* L. by weak illumination at chilling temperatures. *FEBS Lett* **362**: 235–238
- Stombaugh NA, Sundquist JE, Burris RH, Orme-Johnson WH (1976) Oxidation-reduction properties of several low potential iron-sulfur proteins and of methylviologen. *Biochemistry* **15**: 2633–2641
- Suorsa M, Järvi S, Grieco M, Nurmi M, Pietrzykowska M, Rantala M, Kangasjärvi S, Paakkari V, Tikkanen M, Jansson S, et al (2012) PROTON GRADIENT REGULATION5 is essential for proper acclimation of *Arabidopsis* photosystem I to naturally and artificially fluctuating light conditions. *Plant Cell* **24**: 2934–2948
- Takagi D, Yamamoto H, Amako K, Makino A, Sugimoto T, Miyake C (2012) O₂ supports 3-phosphoglycerate-dependent O₂ evolution in chloroplasts from spinach leaves. *Soil Sci Plant Nutr* **58**: 462–468
- Takahashi M, Asada K (1982) Dependence of oxygen affinity for Mehler reaction on photochemical activity of chloroplast thylakoids. *Plant Cell Physiol* **23**: 1457–1461
- Takahashi M, Asada K (1988) Superoxide production in aprotic interior of chloroplast thylakoids. *Arch Biochem Biophys* **267**: 714–722
- Takahashi MA, Kono Y, Asada K (1980) Reduction of plastocyanin with O₂ and superoxide dismutase-dependent oxidation of plastocyanin by H₂O₂. *Plant Cell Physiol* **21**: 1431–1438
- Tanaka K, Takahashi MA, Asada K (1978) Isolation of monomeric cytochrome *f* from Japanese radish and a mechanism of autoreduction. *J Biol Chem* **253**: 7397–7403
- Telfer A (2014) Singlet oxygen production by PSII under light stress: mechanism, detection and the protective role of β -carotene. *Plant Cell Physiol* **55**: 1216–1223
- Terashima I, Funayama S, Sonoike K (1994) The site of photoinhibition in leaves of *Cucumis sativus* L. at low temperatures is photosystem I, not photosystem II. *Planta* **193**: 300–306
- Tikhonov AN (2013) pH-dependent regulation of electron transport and ATP synthesis in chloroplasts. *Photosynth Res* **116**: 511–534
- Tjus SE, Møller BL, Scheller HV (1998) Photosystem I is an early target of photoinhibition in barley illuminated at chilling temperatures. *Plant Physiol* **116**: 755–764
- Vass I (2011) Role of charge recombination processes in photodamage and photoprotection of the photosystem II complex. *Physiol Plant* **142**: 6–16
- Voss I, Goss T, Murozuka E, Altmann B, McLean KJ, Rigby SE, Munro AW, Scheibe R, Hase T, Hanke GT (2011) FdC1, a novel ferredoxin protein capable of alternative electron partitioning, increases in conditions of acceptor limitation at photosystem I. *J Biol Chem* **286**: 50–59
- Zaks J, Amarnath K, Kramer DM, Niyogi KK, Fleming GR (2012) A kinetic model of rapidly reversible nonphotochemical quenching. *Proc Natl Acad Sci USA* **109**: 15757–15762
- Zhang S, Scheller HV (2004) Photoinhibition of photosystem I at chilling temperature and subsequent recovery in *Arabidopsis thaliana*. *Plant Cell Physiol* **45**: 1595–1602
- Zivcak M, Brestic M, Kunderlikova K, Olsovska K, Allakhverdiev SI (2015a) Effect of photosystem I inactivation on chlorophyll a fluorescence induction in wheat leaves: does activity of photosystem I play any role in OJIP rise? *J Photochem Photobiol B* **152**(Pt B): 318–324
- Zivcak M, Brestic M, Kunderlikova K, Sytar O, Allakhverdiev SI (2015b) Repetitive light pulse-induced photoinhibition of photosystem I severely affects CO₂ assimilation and photoprotection in wheat leaves. *Photosynth Res* **162**: 449–463
- Zulfugarov IS, Tovuu A, Eu YJ, Dogsom B, Poudyal RS, Nath K, Hall M, Banerjee M, Yoon UC, Moon YH, et al (2014) Production of superoxide from Photosystem II in a rice (*Oryza sativa* L.) mutant lacking PsbS. *BMC Plant Biol* **14**: 242

1 **Supplementary Figures**

2

3 **Title: Superoxide and singlet oxygen produced within the thylakoid membranes both**
4 **cause photosystem I photoinhibition**

5 Running title: The photoinhibition mechanism in Photosystem I

6

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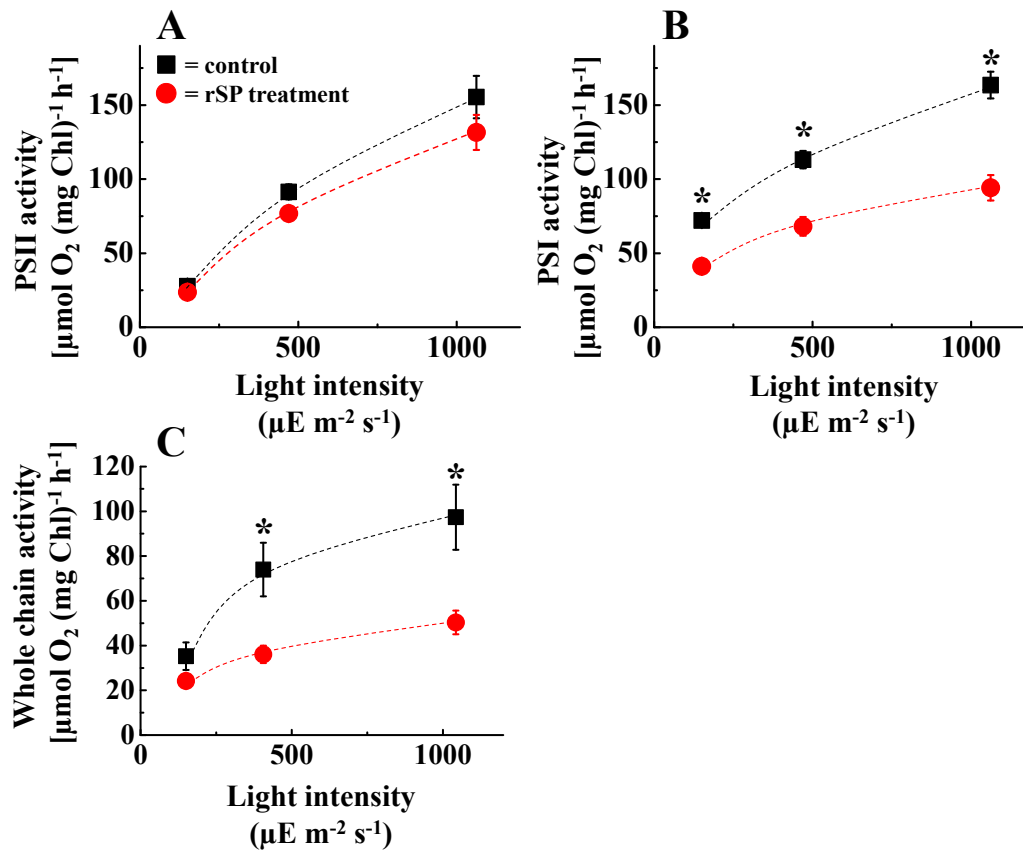
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36 **Supplemental Figure S1**

37 The light response of PSII (A), PSI (B), and whole-chain photosynthetic electron

38 transport activity (C) in isolated chloroplasts. The reaction mixture contained $30 \mu\text{g ml}^{-1}$

39 isolated chloroplasts, and the reaction mixture was maintained at 25°C . Photosynthetic

40 electron activities were measured using an O_2 electrode (see Materials and Methods).

41 Black square indicates photosynthetic electron activities of the control sample that was

42 kept in the dark for 1 h. Red circle indicates photosynthetic electron activities of

43 samples that were processed by SP treatment for 1 h. Data are expressed as the mean \pm

44 SEM of three independent experiments. Asterisks indicate a significant difference

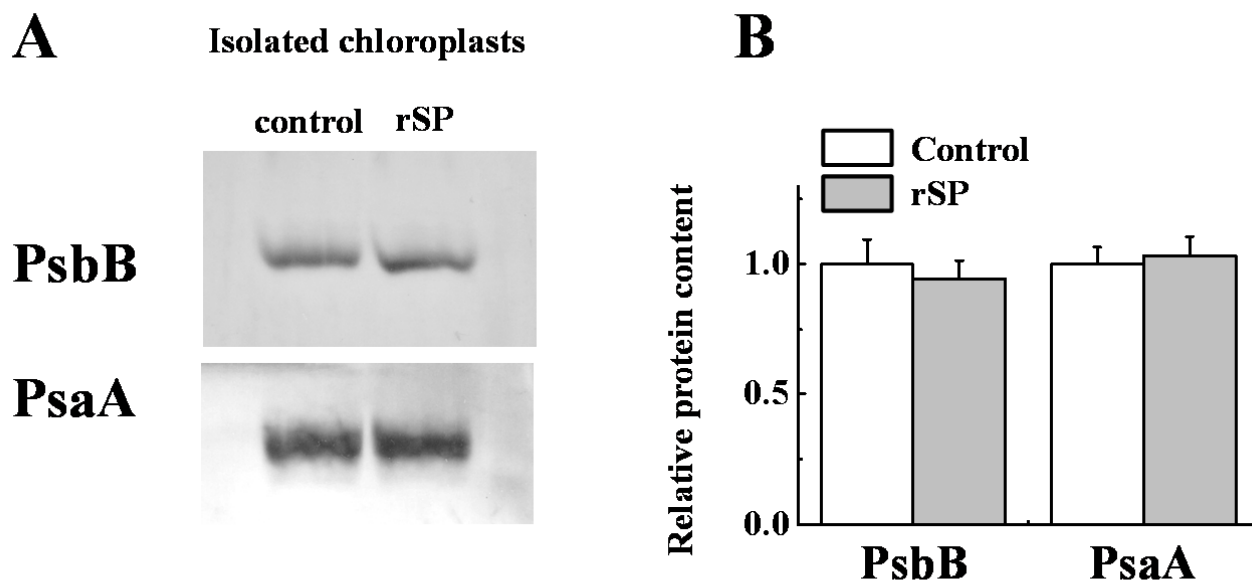
45 between the control sample and the rSP treatment sample (Student's *t*-test, $p < 0.05$).

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52 **Supplemental Figure S2**

53 The comparison of PSII and PSI core protein content in isolated chloroplast between
 54 before and after SP treatment. (A) Isolated chloroplasts before rSP treatment were used
 55 as a control, and isolated chloroplasts treated rSP treatment for 1h were used as an rSP
 56 treated sample. We used antiserum specific to PsbB and PsaA for quantifying PSII and
 57 PSI core protein content. The protein corresponding to 0.6 μg chlorophyll was loaded in
 58 each lane. The relative protein content is quantified (B). The protein content in control
 59 sample was set to 1. White bars indicate the protein content in control sample, and gray
 60 bars indicate the protein content in rSP treated sample. Data are expressed as the mean \pm
 61 SEM of three independent experiments.

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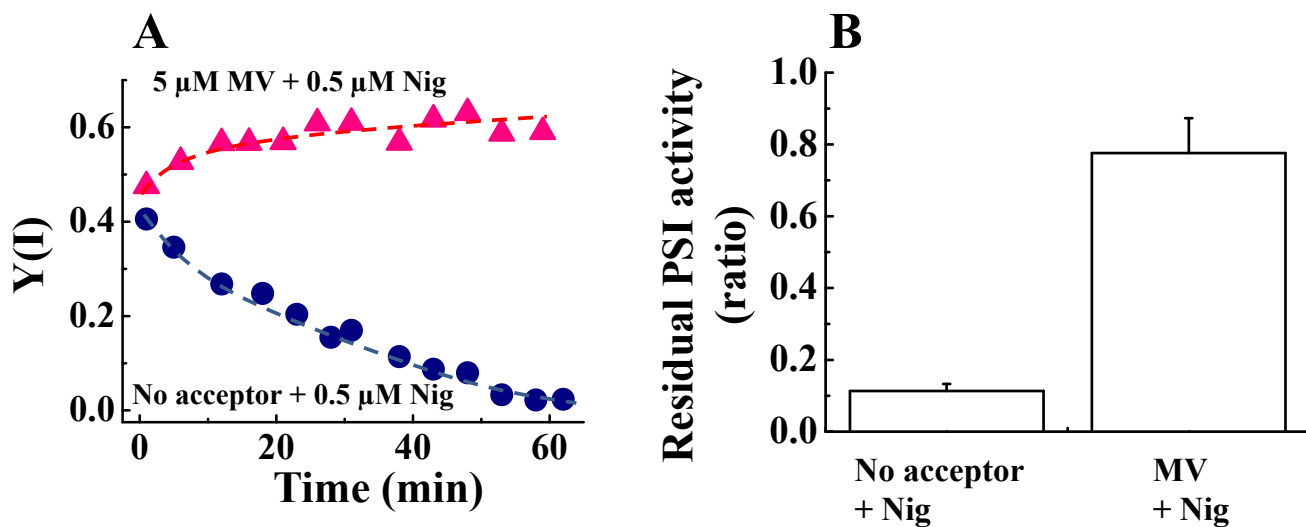
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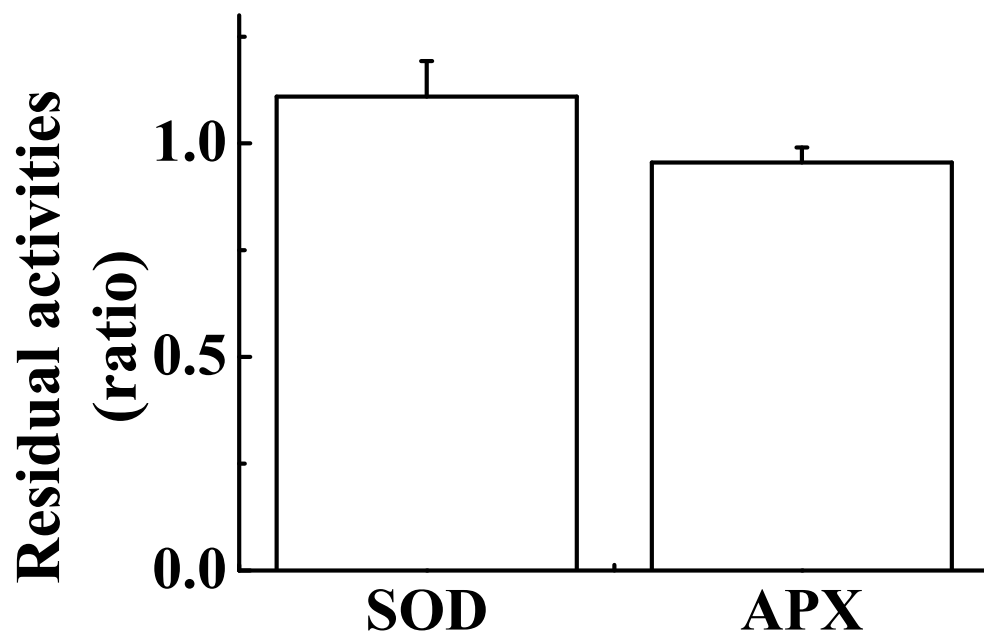


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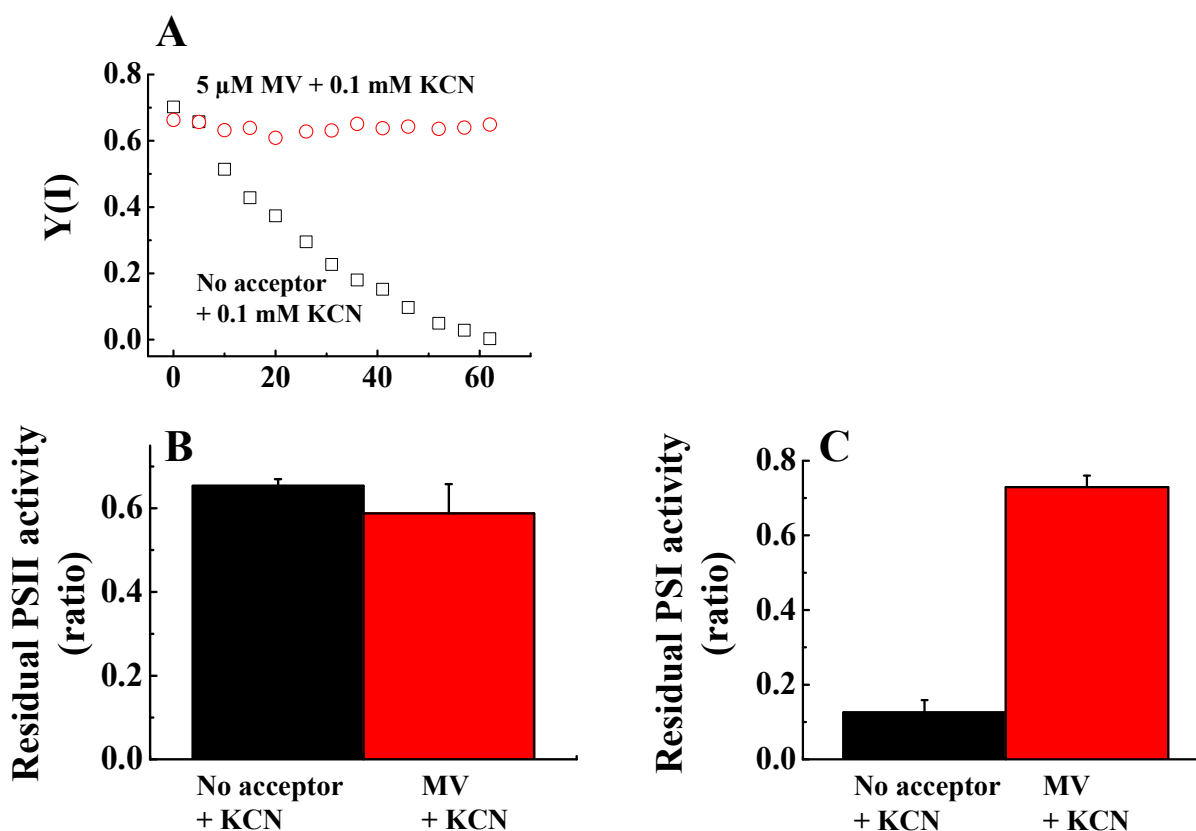
Supplemental Figure S3

The effect of nigericin on rSP treatment in the absence and presence of MV. The reaction mixture contains $30 \mu\text{g ml}^{-1}$ isolated chloroplasts and $0.5 \mu\text{M}$ nigericin, and the reaction mixture was maintained at 25°C . (A) The time-course analysis of Y(I) in isolated chloroplasts in the absence and presence of MV ($5 \mu\text{M}$). Experiments were repeated at least three times and representative data are shown. (B) The residual activity of PSI after rSP treatment in the absence and presence of MV. After rSP treatment, the reaction mixture was kept in the dark for 30 min and the Pm was measured. Data were normalized to the Pm before rSP treatment, and the data represents the residual activity of PSI after rSP treatment. Data are expressed as mean \pm SEM of three independent experiments.

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95 **Supplemental Figure S4**
96 The change in SOD and APX activities before and after rSP treatment. Chloroplastic
97 SOD and APX activities were compared before and after rSP treatment and the residual
98 activities are their ratio. Data are expressed as mean \pm SEM of at least nine independent
99 experiments. Absolute SOD activities were 233 ± 49 mU (mg Chl)⁻¹, and absolute APX
100 activities were 5.6 ± 0.7 μ mol (mg Chl)⁻¹ min⁻¹.
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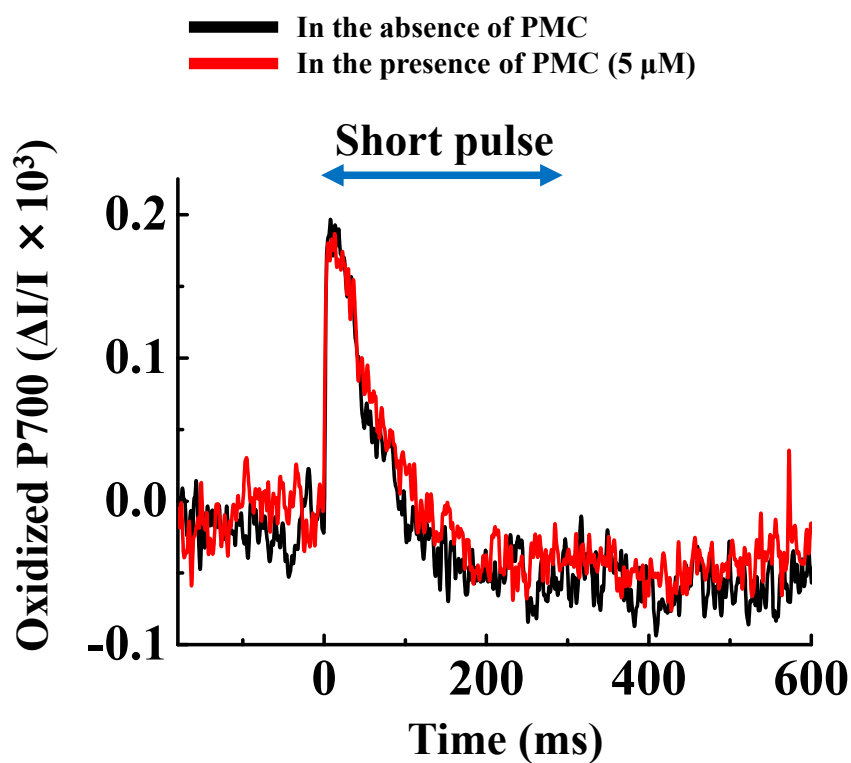
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111 | **Supplemental Figure S5**

112 The effect of KCN on rSP treatment in the absence and presence of MV. The reaction
 113 mixture contains 30 μ g ml⁻¹ isolated chloroplasts and 0.1 mM nigericin, and the reaction
 114 mixture was maintained at 25°C. (A) The time-course analysis of Y(I) in isolated
 115 chloroplasts in the absence and presence of MV (5 μ M). Experiments were repeated at
 116 least three times and representative data are shown. (B) The residual activity of PSII
 117 after rSP treatment in the absence and presence of MV. (C) The residual activity of PSI
 118 after rSP treatment in the absence and presence of MV. After rSP treatment, the reaction
 119 mixture was kept in the dark for 30 min and the Fv/Fm and Pm were measured. Data
 120 were normalized to the Fv/Fm and Pm before rSP treatment, and the data represents the
 121 residual activity of PSII and PSI after rSP treatment. Data are expressed as mean \pm SEM
 122 of three independent experiments.

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Supplemental Figure S6

The kinetics of oxidized P700 induced by a short-pulse in the absence of and in the presence of PMC (5 μM). The reaction mixture contained 30 $\mu\text{g ml}^{-1}$ isolated chloroplasts, and the reaction mixture was maintained at 25°C. The data were obtained after rSP treatment was applied for 5 min. Black line shows the condition in the absence of PMC. Red line shows the condition in the presence of PMC. Experiments were at least three times. Short-pulse was illuminated every 10 s, and averaged data during three short-pulse illuminations were shown.

1 **Supplementary Tables**

2

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Supplemental Table S1

Oxygen evolution rate and Y(II) in the absence and in the presence of PMC

	control	+PMC (5 μ M)
O₂ absorption rate [μ mol O ₂ (mg Chl) ⁻¹ h ⁻¹]	2.87 \pm 0.69	2.65 \pm 0.51
Y(II)	0.035 \pm 0.022	0.035 \pm 0.013

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41 Isolated chloroplasts were illuminated by red light (500 μ E m⁻² s⁻¹) and steady state O₂
42 evolution rate and Y(II) were measured in the absence of and in the presence of PMC (5
43 μ M) in the reaction mixture. The reaction mixture contained 30 μ g ml⁻¹ isolated
44 chloroplasts, and the reaction mixture was maintained at 25°C. Data are expressed as
45 mean \pm SEM of three independent experiments.

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