



CD47-signal regulatory protein α signaling system and its application to cancer immunotherapy

Murata, Yoji
Saito, Yasuyuki
Kotani, Takenori
Matozaki, Takashi

(Citation)

Cancer Science, 109(8):2349-2357

(Issue Date)

2018-08

(Resource Type)

journal article

(Version)

Version of Record

(Rights)

© 2018 The Authors. Cancer Science published by John Wiley & Sons Australia, Ltd on behalf of Japanese Cancer Association.

This is an open access article under the terms of the Creative Commons Attribution - NonCommercial License, which permits use, distribution and reproduction in any medium...

(URL)

<https://hdl.handle.net/20.500.14094/90005185>



REVIEW ARTICLE

CD47-signal regulatory protein α signaling system and its application to cancer immunotherapy

Yoji Murata  | Yasuyuki Saito | Takenori Kotani | Takashi Matozaki 

Division of Molecular and Cellular Signaling,
Department of Biochemistry and Molecular
Biology, Kobe University Graduate School
of Medicine, Kobe, Japan

Correspondence

Takashi Matozaki, Division of Molecular and
Cellular Signaling, Department of
Biochemistry and Molecular Biology, Kobe
University Graduate School of Medicine,
Kobe, Japan.
Email: matozaki@med.kobe-u.ac.jp

Funding information

Terumo Foundation for Life Sciences and
Arts; Japan Agency for Medical Research
and Development (P-CREATE and P-
DIRECT); Takeda Science Foundation;
Uehara Memorial Foundation; Grant-in-Aid
for Scientific Research (B) from the Japan
Society for the Promotion of Science (JSPS)
(26291022); Japanese Society of
Hematology

Tumor cells evade immune surveillance through direct or indirect interactions with various types of immune cell, with much recent attention being focused on modifying immune cell responses as the basis for the development of new cancer treatments. Signal regulatory protein α (SIRP α) and CD47 are both transmembrane proteins that interact with each other and constitute a cell-cell communication system. SIRP α is particularly abundant in myeloid cells such as macrophages and dendritic cells, whereas CD47 is expressed ubiquitously and its expression level is elevated in cancer cells. Recent studies have shown that blockade of CD47-SIRP α interaction enhances the phagocytic activity of phagocytes such as macrophages toward tumor cells in vitro as well as resulting in the efficient eradication of tumor cells in a variety of xenograft or syngeneic mouse models of cancer. Moreover, CD47 blockade has been shown to promote the stimulation of tumor-specific cytotoxic T cells by macrophages or dendritic cells. Biological agents, such as Abs and recombinant proteins, that target human CD47 or SIRP α have been developed and are being tested in preclinical models of human cancer or in clinical trials with cancer patients. Preclinical studies have also suggested that CD47 or SIRP α blockade may have a synergistic antitumor effect in combination with immune checkpoint inhibitors that target the adaptive immune system. Targeting of the CD47-SIRP α signaling system is thus a promising strategy for cancer treatment based on modulation of both innate and acquired immune responses to tumor cells.

KEYWORDS

cancer immunotherapy, CD47, macrophage, phagocytosis, signal regulatory protein α (SIRP α)

1 | INTRODUCTION

The tumor microenvironment consists of immune cells and a variety of stromal cell types, including fibroblasts and endothelial cells, as well as soluble and insoluble factors, such as cytokines, chemokines, and

extracellular matrix.^{1,2} This microenvironment plays an important role in the regulation of tumor progression by promoting tumor cell survival, invasion, and metastasis as well as angiogenesis.¹⁻³ Cross-talk between tumor and immune cells in the tumor microenvironment is also thought to contribute to the evasion of tumor cells from immune surveillance. For example, the binding of PD-1 on cytotoxic T lymphocytes to its ligand PD-L1 on tumor cells prevents killing of the latter cells by the former.⁴ Indeed, Abs to PD-1 and to PD-L1 are now in clinical use for the treatment of diverse solid tumors, including advanced melanoma, renal cell carcinoma, and non-small-cell lung cancer.^{5,6}

Abbreviations: ADCC, antibody-dependent cellular cytotoxicity; ADCP, antibody-dependent cellular phagocytosis; CTLA-4, cytotoxic T lymphocyte-associated antigen-4; DC, dendritic cell; HER2, human epidermal growth factor receptor 2; NK, natural killer; PD-1, programmed cell death-1; PD-L1, programmed cell death-ligand 1; RBC, red blood cell; SH2, Src homology 2; Shp, SH2 domain-containing phosphatase; SIRP α , signal regulatory protein α .

This is an open access article under the terms of the Creative Commons Attribution-NonCommercial License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited and is not used for commercial purposes.

© 2018 The Authors. *Cancer Science* published by John Wiley & Sons Australia, Ltd on behalf of Japanese Cancer Association.

In addition, Abs to the T-cell molecule CTLA-4, which is also thought to suppress T-cell responses on interaction with CD80 or CD86 on antigen-presenting cells, are given to treat melanoma as well as prostate and lung cancers.^{5,7} Molecules that participate in the negative regulation of the antitumor response of immune cells are thus promising targets for cancer therapy, with drugs that target such molecules being known as immune checkpoint inhibitors.⁸ We and others have recently shown that blocking Abs to SIRP α , which is highly expressed in macrophages and DCs, also has the potential to function as immune checkpoint inhibitors—that target innate immunity, in particular—for cancer treatment.^{9–11}

2 | SIGNAL REGULATORY PROTEIN α AND ITS BIOLOGICAL FUNCTIONS

Signal regulatory protein α (also known as SHPS-1, p84, BIT, or CD172a) is a transmembrane protein that was originally identified as a highly expressed glycoprotein in the brain and a binding partner or putative substrate for 2 cytoplasmic-type protein tyrosine phosphatases, SH2 domain-containing phosphatase 1 (Shp1, also known as PTPN6) and Shp2 (PTPN11).^{12–15} Indeed, SIRP α contains 3 Ig-like domains in its extracellular region and 4 tyrosine residues that are putative phosphorylation sites in its cytoplasmic region (Figure 1A).^{16,17} The extracellular region of SIRP α interacts with its ligand, CD47, which was originally identified in association with $\alpha_v\beta_3$ integrin and is also a member of the Ig superfamily of proteins, with an Ig-V-like extracellular domain, 5 putative membrane-spanning segments, and a short cytoplasmic tail (Figure 1A).¹⁸ In their phosphorylated state, the tyrosine phosphorylation sites (in particular, the 2 COOH-terminal sites) in the cytoplasmic region of SIRP α bind to the SH2 domains of Shp1 and Shp2 and thereby activate these phosphatases (Figure 1A). Tyrosine phosphorylation of the cytoplasmic region of SIRP α is triggered by various growth factors and cytokines as well as by integrin-mediated cell adhesion to extracellular matrix proteins.^{14,19} Ligation of SIRP α by CD47 also promotes tyrosine phosphorylation of its cytoplasmic region.^{20,21} SIRP α thus functions as a docking protein for the recruitment and activation of Shp1 and Shp2 at the cell membrane in response to extracellular stimuli,¹⁶ with these phosphatases then being thought to play an important role in signaling downstream of SIRP α . SIRP α is especially abundant in neurons and in hematopoietic cells of the myeloid lineage such as macrophages, neutrophils, and DCs,^{22–26} whereas CD47 is expressed in most cell types.¹⁸

Signal regulatory protein α mutant mice, which express a mutant form of SIRP α that lacks most of the cytoplasmic region and thus fail to bind Shp1 and Shp2,²⁴ manifested mild anemia associated with a short lifetime of RBCs as a result of increased phagocytotic activity of splenic macrophages against RBCs,^{21,24} suggesting the importance of SIRP α for both the lifespan of RBCs and their number in the circulation. In addition, DC-specific SIRP α knockout mice showed a reduced number of DCs, as well as of fibroblastic reticular cells, a subset of stromal cells, in the spleen.²⁷ Moreover, SIRP α mutant mice, as well as CD47-deficient mice, were resistant to the development of autoimmune animal models, such

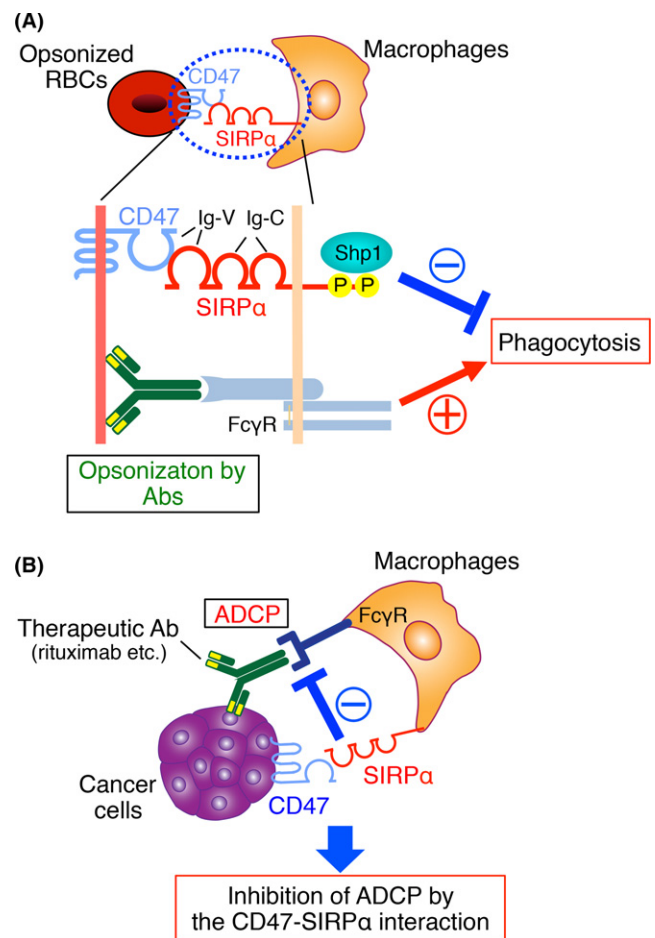


FIGURE 1 The CD47-signal regulatory protein α (SIRP α) signaling system and its role in the regulation of phagocytosis by macrophages. A, SIRP α is a transmembrane protein that contains 3 Ig-like domains (1 V-like and 2 C1-like Ig domains) in its NH₂-terminal extracellular region and 2 key tyrosine phosphorylation sites in its COOH-terminal cytoplasmic region. The tyrosine-phosphorylated sites of SIRP α bind and thereby activate the protein tyrosine phosphatases Shp1 and Shp2. The SIRP α ligand CD47 is also a member of the Ig superfamily, with an Ig-V-like extracellular domain, 5 membrane-spanning segments, and a short cytoplasmic tail. The Ig-V-like domain of CD47 interacts in trans with the NH₂-terminal Ig-V-like domain of SIRP α and thereby triggers the tyrosine phosphorylation of the latter protein. Ligation of SIRP α on macrophages by CD47 on opsonized red blood cells (RBCs) thus promotes tyrosine phosphorylation of SIRP α and its subsequent association with Shp1, resulting in inhibition of RBCs phagocytosis by the macrophages elicited by the interaction of the Fc region of RBC-bound Abs with the macrophage Fc γ receptor (Fc γ R). B, Interaction of CD47 on tumor cells with SIRP α on macrophages attenuates phagocytosis by the macrophages of the tumor cells triggered by opsonization with tumor antigen-specific therapeutic Abs such as rituximab. ADCP, antibody-dependent cellular phagocytosis

as experimental autoimmune encephalomyelitis,^{28,29} suggesting that the interaction of SIRP α with CD47 is involved in the development of autoimmune diseases. SIRP α and CD47 are also thought to play a role in the regulation of central nervous system functions. Both SIRP α mutant mice and CD47-deficient mice, indeed, showed prolonged immobility

(depression-like behavior) in the forced swim test.³⁰ The CD47-SIRP α signaling system is thus likely to act as a signaling platform for the brain response to stress and the regulation of depression-like behavior in the forced swim test.

By contrast, mutations associated with human diseases, including hematological disorders, autoimmune diseases and neurological disorders, have not been identified within *CD47* and *SIRP α* genes. Although the N-terminal IgV-like domain of human SIRP α , which is responsible for the interaction with CD47, is known to be highly polymorphic,³¹ any relationship between such polymorphisms of SIRP α and the incidence of diseases has not been reported so far.

3 | BINDING OF CD47 TO SIRP α PREVENTS PHAGOCYTOSIS IN MACROPHAGES

Oldenborg and colleagues^{20,32} first showed that the rate of clearance of transfused CD47-deficient RBCs from the bloodstream of WT mice was markedly increased compared with that of transfused WT cells.^{20,32} In addition, the phagocytosis of CD47-deficient RBCs by isolated splenic macrophages from WT mice in vitro was greatly enhanced compared with that of WT RBCs.²⁰ We also subsequently showed that the rate of clearance of transfused WT RBCs from the bloodstream was markedly increased in SIRP α mutant mice. Phagocytosis of antibody-opsonized WT RBCs by isolated macrophages from the SIRP α mutant mice was also enhanced compared with that seen with macrophages from WT mice.²¹ Collectively, these observations indicated that the binding of CD47 on RBCs to SIRP α on macrophages prevents the Fc γ receptor (Fc γ R)-dependent phagocytosis of the former cells by the latter (Figure 1A). The activity of Shp1, which binds to the tyrosine-phosphorylated cytoplasmic region of SIRP α , is thought to be important for this prevention of phagocytosis (Figure 1A).³²

4 | BLOCKADE OF THE CD47-SIRP α SIGNALING SYSTEM HAS ANTITUMOR EFFECTS

Two major groups of molecularly targeted drugs are currently in clinical use for cancer therapy. One group includes potent inhibitors of a variety of signaling molecules that are essential for the proliferation of tumor cells, such as inhibitors of various tyrosine kinases as well as of the Raf-MEK and PI3K-mTOR (mammalian target of rapamycin) signaling pathways.³³⁻³⁷ The other group consists of mAbs to surface molecules that are highly expressed in particular cancers, such as rituximab (to CD20 in lymphoma), trastuzumab (to HER2 in breast cancer), and cetuximab (to epidermal growth factor receptor [EGFR] in colon cancer). These mAbs are thought to bind to their targets on tumor cells and thereby induce the killing of these cells by complement-dependent cytotoxicity as well as via ADCP mediated by macrophages and ADCC mediated by NK cells, both of which require the interaction of Fc receptors on the effector cells with the Fc domain of the bound

mAbs.³⁸ Many studies have shown that the interaction of CD47 on cancer cells with SIRP α on macrophages serves to inhibit phagocytosis of the former cells by the latter cells (Figure 1B).^{9,39-41} Blockade of the CD47-SIRP α signaling system was thus shown to enhance the ADCP-mediated killing of Ab-opsonized tumor cells by macrophages (Figure 2A) as well as to suppress tumor growth and metastasis in preclinical mouse models of cancer.

4.1 | Targeting of CD47

CD47 was shown to be identical to the tumor antigen OV-3, which interacts with integrins and is markedly upregulated in ovarian carcinoma cells.⁴² Expression of CD47 was also found to be increased in AML stem cells, non-Hodgkin lymphoma, and various solid tumors compared with their normal counterparts,^{40,43,44} and such increased expression was associated with poor prognosis in patients with these malignancies.^{40,43-46} Incubation with a blocking Ab to CD47 either alone or in combination with mAbs to tumor antigens in vitro promoted the phagocytosis by macrophages of cancer cells such as AML stem cells, non-Hodgkin lymphoma cells, colorectal cancer stem cells, and breast cancer cells (Figure 2A).^{40,43,44} Such Ab treatment also attenuated tumor growth and metastasis in a variety of xenograft or syngeneic mouse models of AML, non-Hodgkin lymphoma, pediatric brain tumors, as well as ovarian, colon, breast, bladder, and small-cell lung cancers.^{40,43,44,47-49} Moreover, an engineered recombinant SIRP α protein, which has a higher affinity for CD47 than does the WT protein and prevents endogenous CD47-SIRP α interaction, was found to significantly enhance the efficacy of therapeutic Abs such as rituximab, trastuzumab, and alemtuzumab (an Ab to CD52) in mouse xenograft models of human cancer.⁵⁰

In addition to its effect on ADCP of tumor cells by macrophages, CD47 blockade promotes the activation of tumor-specific cytotoxic T cells by DCs or macrophages.^{40,44,51-53} Treatment of tumor-bearing mice with blocking Abs to CD47 thus promoted the recognition of tumor-derived DNA through the stimulator of interferon genes (STING) pathway.⁵² Such recognition increased type I interferon production by DCs and enhanced the cross-priming of tumor-specific cytotoxic T cells.⁵³ Tumor-infiltrating macrophages were also found to participate in the cross-priming of tumor-specific cytotoxic T cells.⁵¹ Blockade of CD47-SIRP α interaction is thus thought to promote the phagocytosis of tumor cells by macrophages and DCs, which, in turn, activate tumor-specific cytotoxic T cells at the site of tumor rejection. In contrast, given that CD47 is expressed on most cell types, off-target effects of CD47 blockade on normal cells—in particular, erythrocytes—is a potential concern with regard to treatment with drugs that target CD47. Indeed, treatment with Abs to CD47 induced in a dose-dependent way the development of transient anemia associated with reticulocytosis in cynomolgus monkeys.⁴⁷

4.2 | Targeting of SIRP α

Blockade of SIRP α in combination with Abs to tumor antigens is also a promising strategy for cancer therapy. Indeed, Abs to mouse SIRP α enhanced the rituximab-induced elimination of human Burkitt's

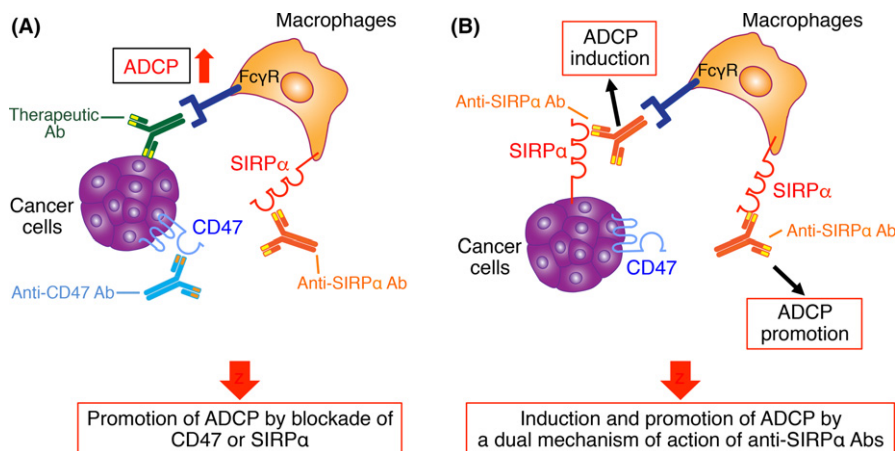


FIGURE 2 Suppression of tumor growth and metastasis by blockade of CD47-signal regulatory protein α (SIRP α) interaction and consequent promotion of macrophage-mediated antibody-dependent cellular phagocytosis (ADCP). A, Inhibition of the interaction of CD47 on tumor cells with SIRP α on macrophages by biological agents such as Abs to these proteins promotes macrophage-mediated ADCP of the tumor cells triggered by opsonization with therapeutic Abs to tumor antigens and thereby leads to suppression of tumor growth and metastasis. B, Blocking Abs to SIRP α bind to this protein on both macrophages and certain tumor cells such as melanoma and renal cell carcinoma cells. Such binding results in both direct induction of macrophage-mediated ADCP of the tumor cells as well as blockade of CD47-SIRP α signaling that negatively regulates such phagocytosis

lymphoma Raji cells transplanted into immunodeficient non-obese diabetic (NOD)/SCID mice,¹¹ whose endogenous SIRP α has a high affinity for human CD47.³¹ In addition, whereas tumor growth or metastasis did not differ between SIRP α mutant and WT mice injected with syngeneic melanoma cells, treatment with therapeutic Abs specific for a melanoma antigen eliminated tumor cells to a markedly greater extent in the mutant mice than in the WT animals.⁴⁵ Moreover, an Ab to human SIRP α that inhibits CD47-SIRP α interaction enhanced killing by human phagocytes of HER2-positive breast cancer cells opsonized with the HER2-specific mAb trastuzumab *in vitro*.⁴⁵ Abs to mouse SIRP α also enhanced the phagocytic activity of bone marrow-derived macrophages from NOD mice toward Raji cells opsonized with rituximab.¹¹ Collectively, these findings suggest that blocking Abs to SIRP α has the potential to promote tumor elimination *in vivo* by enhancement of macrophage-mediated ADCP of cancer cells opsonized by Abs to tumor antigens (Figure 2A). In addition to Abs to SIRP α , recombinant CD47 proteins that contain the NH₂-terminal Ig-V-like domain and block CD47-SIRP α interaction may also contribute to the killing of tumor cells. Indeed, an NH₂-terminal variant of CD47 with a higher affinity for SIRP α than WT CD47 was found to act synergistically with tumor-specific mAbs to promote macrophage-mediated phagocytosis of tumor cells *in vitro*,⁵⁴ although its potential antitumor effects *in vivo* have yet to be evaluated.

Blocking Abs to SIRP α might also have therapeutic effects as single agents in the case of SIRP α -expressing tumors, with the antitumor effects of such Abs being mediated by a dual mechanism of action: direct induction of ADCP of tumor cells by macrophages and blockade of CD47-SIRP α signaling that negatively regulates such phagocytosis (Figure 2B). Similar to the increased expression of CD47 apparent in many types of cancer, SIRP α is also more prominently expressed in tumor tissue from patients with renal cell carcinoma or melanoma compared with the surrounding normal tissue.¹¹ Of note, treatment with a blocking Ab to mouse SIRP α resulted in a marked reduction in

the tumor burden of immunocompetent mice injected with syngeneic renal cell carcinoma or melanoma cells, both of which highly expressed endogenous SIRP α .¹¹ This antitumor action of the Ab was significantly attenuated by selective depletion of macrophages. Moreover, the expression of SIRP α on tumor cells and the Fc region of the Ab to SIRP α were required for promotion of the phagocytosis of mouse renal cell carcinoma cells by macrophages *in vitro*, suggesting that opsonization of the tumor cells by the Ab induces ADCP through activation of the Fc receptor on macrophages.

In addition to macrophages, both cytotoxic T cells and NK cells may contribute to the antitumor effects of blocking Abs to SIRP α .¹¹ Such Abs might thus initiate cross-priming of tumor-specific cytotoxic T cells by macrophages and DCs. Moreover, the Abs might induce NK cell-mediated ADCC toward tumor cells directly and contribute to the activation of NK cells by tumor-infiltrating macrophages or DCs.⁵⁵⁻⁵⁷ Of interest, treatment with blocking Abs to SIRP α did not induce obvious adverse effects including anemia in mice.¹¹ Treatment of mice with Abs to SIRP α also did not cause any obvious neurotoxicity regardless of the high expression of SIRP α in the neuron.¹¹ Such Abs might thus be worth pursuing for cancer therapy, especially for the treatment of melanoma and renal cell carcinoma.

4.3 | Therapeutic drugs targeting CD47 or SIRP α

Several biological agents, including Abs and recombinant proteins, that are able to bind to human CD47 specifically and block the human CD47-SIRP α interaction have been developed for cancer treatment (Table 1). Hu5F9-G4 was the first drug to be developed as a humanized mAb to human CD47.⁴⁷ This agent enhances the phagocytosis of tumor cells by macrophages *in vitro* and eradicates human hematological malignancies and solid tumors in xenograft mouse models.⁴⁷⁻⁴⁹ Phase I or I/II clinical trials of Hu5F9-G4 and

TABLE 1 Therapeutic agents in preclinical or clinical development that target CD47 or SIRP α

Company	Country	Drug	Description	Phase	Disease	Strategy	Combination agent	ID
Forty Seven Inc.	USA	Hu5F9-G4	Anti-CD47 Ab (IgG4)	I	Solid tumors, NHL	Mono		NCT02216409
				I	AML, MDS	Mono		NCT02678338
				I/II	CRC, solid tumors	Combi	Cetuximab	NCT02953782
				I/II	NHL	Combi	Rituximab	NCT02953509
				I	AML, MDS	Mono/Combi	Azacitidine	NCT03248479
Celgene	USA	CC-90002	Anti-CD47 Ab (IgG4)	I	Solid tumors, MM, NHL	Mono/Combi	Rituximab	NCT02367196
				I	AML, MDS	Mono		NCT02641002
Trillium Therapeutics Inc.	Canada	TTI-621	SIRP α -Fc fusion protein (IgG1)	I	Hematological malignancies, solid tumors	Mono/Combi	Rituximab, nivolumab	NCT02663518
				I	Solid tumors	Mono/Combi	PD-1/PD-L1 inhibitor, PEG-IFN- α 2a, T-Vec, radiation	NCT02890368
Alexo Therapeutics	USA	ALX148	SIRP α V1-Fc fusion protein	I	Solid tumors, NHL	Mono/Combi	Pembrolizumab, trastuzumab, rituximab	NCT03013218
Novimmune SA	Switzerland	NI-1701	Anti-CD47/CD19 bispecific Ab	Preclinical				
		NI-1801	Anti-CD47/mesothelin bispecific Ab	Preclinical				
Arch Oncology	USA	AO-104, -108, -176	Anti-CD47 Ab	Preclinical				
Surface Oncology Inc.	USA	SRF231	Anti-CD47 Ab	Preclinical				
Hummingbird Bioscience	Singapore	HMBD004	Anti-CD47/CD33 bispecific Ab	Preclinical				
OSE Immunotherapeutics	France	OSE-172	Anti-SIRP α Ab	Preclinical				

Combi, combination therapy; CRC, colorectal carcinoma; ID, clinicaltrials.gov identifier; MDS, myelodysplastic syndrome; MM, multiple myeloma; Mono, monotherapy; NHL, non-Hodgkin lymphoma; PD-1, programmed cell death-1; PD-L1, programmed cell death-ligand 1; PEG-IFN- α 2a, pegylated interferon- α 2a; SIRP α , signal regulatory protein α ; T-Vec, talimogene laherparepvec.

another humanized mAb to human CD47, CC-90002, are now being conducted for both solid tumors and hematological malignancies. Recombinant SIRP α proteins, such as TTI-621 and ALX148, are also in phase I clinical trials for patients with hematological malignancies or solid tumors (Table 1). TTI-621 consists of the Ig-V-like domain of human SIRP α linked to the Fc region of human IgG1, and it was shown to enhance the phagocytosis of tumor cells by macrophages in vitro and to effectively control tumor growth in xenograft models of aggressive AML or B lymphoma as well as in a syngeneic mouse model of B lymphoma.^{58,59} Like TTI-621, ALX148 has a higher affinity for human CD47 than does WT SIRP α , but it comprises a variant of the Ig-V-like domain of human SIRP α fused to an inactive Fc domain. Both of these drugs are currently being tested in clinical trials as monotherapy or in combination either with molecularly

targeted agents including Abs to tumor antigens and immune checkpoint inhibitors or with radiotherapy. Various other CD47-targeting drugs including bispecific Abs that bind to both tumor-specific antigens and CD47 are also in preclinical development (Table 1).

5 | COMBINATION OF BLOCKING Abs TO CD47 OR TO SIRP α WITH IMMUNE CHECKPOINT INHIBITORS

Treatment with immune checkpoint inhibitors that target PD-1, PD-L1, or CTLA-4 provides substantial clinical benefit in patients with a wide range of malignancies including metastatic melanoma, renal cell carcinoma, and non-small-cell lung cancer.⁵⁻⁷ However, many

patients remain unresponsive to these therapies. Combinations of immune checkpoint inhibitors with standard chemotherapeutic drugs, small-molecule compounds, cancer vaccines, and immune-stimulatory agents are currently under evaluation in preclinical models and in clinical trials in attempts to increase the efficacy of immune checkpoint inhibition in such unresponsive patients.

Immune checkpoint inhibitors that target PD-1 or PD-L1 prevent the interaction of PD-1 on cytotoxic T cells with PD-L1 on cancer cells, which generates an inhibitory signal within the T cells. Blocking of this interaction is therefore thought to enhance the killing of tumor cells by cytotoxic T cells.^{4,6} Inhibition of both the CD47-SIRP α and PD-1-PD-L1 axes might therefore be expected have a synergistic antitumor action, given that blockade of CD47 is thought to exert antitumor effects by promoting the phagocytosis of tumor cells and tumor cell-releasing substances by macrophages or DCs as well as by enhancing the cross-priming of tumor-specific cytotoxic T cells by these antigen-presenting cells (Figure 3).^{51,52} Indeed, a nanobody (an antigen-binding fragment of an Ab heavy chain) that reacts with CD47 and thereby inhibits the CD47-SIRP α interaction was found to synergize with PD-L1 antagonism to attenuate the growth of tumors formed in immunocompetent mice by s.c. injected syngeneic melanoma cells.^{60,61} The efficacy of combined therapy with Abs to PD-1 and to CTLA-4 in a mouse model of esophageal squamous cell cancer was also enhanced by blocking Abs to CD47.⁶² Moreover, combined blockade of SIRP α and PD-1 had a synergistic antitumor effect in a syngeneic mouse model of colon cancer.¹¹ The combination of

inhibitors of the CD47-SIRP α interaction with immune checkpoint inhibitors that target the PD-1-PD-L1 axis is thus a potential new approach to immunotherapy for a broad range of cancers.

6 | XENOGRAFT TUMOR MODELS FOR PRECLINICAL VALIDATION OF THE ANTITUMOR EFFECTS OF Abs TO HUMAN SIRP α

Animal models that mimic human diseases are important tools with which to investigate the potential therapeutic efficacy and adverse effects of biological agents in patients. However, the antitumor effects of Abs to human SIRP α in preclinical cancer models have remained unclear because such Abs have failed to bind to endogenous SIRP α expressed on macrophages of immunodeficient mice, likely because of differences in the amino acid sequence of the NH₂-terminal Ig-V-like domain between human and mouse SIRP α . In this regard, a blocking Ab to human SIRP α was recently shown to promote the antitumor effects of rituximab and of vorsetuzumab (an Ab to CD70) in human tumor-bearing *Rag2*^{-/-}*Il2rg*^{-/-} immunodeficient mice in which the DNA sequence encoding the extracellular domain of mouse SIRP α was replaced with the corresponding human sequence (hSIRP α ^{KI} mice).^{63,64} With the use of *Rag2*^{-/-}*Il2rg*^{-/-} mice expressing human SIRP α under the control of human regulatory elements (hSIRP α ^{Tg} mice),⁶⁵ we also showed that a blocking Ab to human SIRP α enhanced the inhibitory effect of rituximab on the growth of tumors formed by s.c. injected Raji cells.⁶⁶ This Ab to human SIRP α also increased the phagocytic activity of macrophages from hSIRP α ^{Tg} mice, but not that of those from *Rag2*^{-/-}*Il2rg*^{-/-} mice, as measured in vitro with human cancer cells opsonized with Abs to tumor antigens, suggesting that the interaction of human CD47 on human cancer cells with human SIRP α on mouse macrophages generates an inhibitory signal for macrophage ADPC. Moreover, in both of these genetically modified mouse models transplanted with human tumor cells, the antitumor effects of the Abs to human SIRP α were found to be dependent, at least in part, on macrophages.^{64,66} These mice may thus serve as models for preclinical validation of Abs to human SIRP α in cancer immunotherapy (Figure 4).

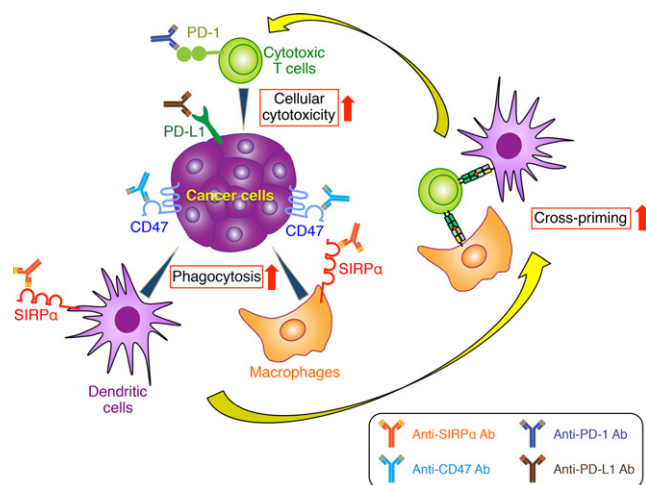


FIGURE 3 Synergistic antitumor effects of blockade of CD47-signal regulatory protein α (SIRP α) interaction combined with immune checkpoint inhibitors. Blockade of CD47 or SIRP α with corresponding specific Abs promotes the phagocytosis of tumor cells by macrophages or dendritic cells as well as the consequent cross-priming of tumor-specific cytotoxic T cells. Activity of cytotoxic T cells is also enhanced by immune checkpoint inhibitors, such as Abs to programmed cell death-1 (PD-1) or programmed cell death-ligand 1 (PD-L1) that prevent the interaction of these proteins. Blockade of CD47 or SIRP α together with giving immune checkpoint inhibitors may therefore have synergistic antitumor effects

7 | CONCLUSION

Attention has recently focused on modifying immune responses as a basis for new cancer treatments. Immunotherapy with immune checkpoint inhibitors that target PD-1, PD-L1, or CTLA-4, which enhance the antitumor activity of cytotoxic T cells, has shown clinical activity in a variety of cancer types. The CD47-SIRP α signaling system serves as an innate immune checkpoint that is thought to help tumor cells evade immune surveillance by preventing their phagocytosis by macrophages and other phagocytes. Numerous studies with preclinical mouse models of cancer have suggested

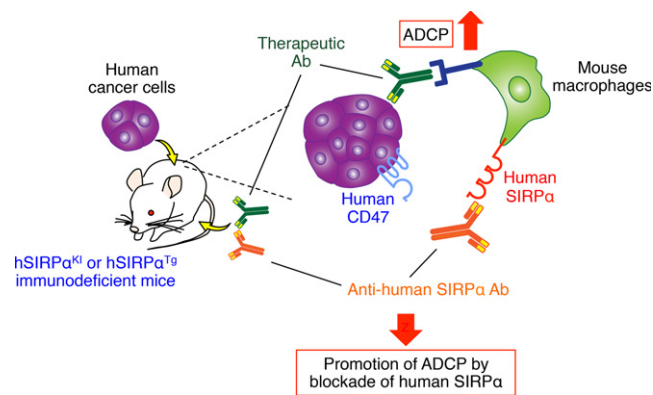


FIGURE 4 Xenograft tumor models for preclinical validation of the antitumor effects of Abs to human signal regulatory protein α (SIRP α). In xenograft tumor models in which human cancer cells are transplanted into human SIRP α knockin (hSIRP α^{KI}) or human SIRP α transgenic (hSIRP α^{Tg}) immunodeficient mice, interaction of human CD47 on human cancer cells with human SIRP α on mouse macrophages inhibits phagocytosis of the former cells by the latter. Prevention of this interaction with Abs to human SIRP α thus enhances antibody-dependent cellular phagocytosis (ADCP) by macrophages of the cancer cells opsonized with therapeutic Abs to tumor-specific antigens

that blockade of CD47 or SIRP α with Abs or recombinant proteins, either alone or in combination with other agents such as Abs to tumor-specific antigens or immune checkpoint inhibitors, holds promise for the treatment of various types of cancer. However, the mechanisms underlying the initiation and consolidation of immune responses to tumor cells by CD47 or SIRP α blockade, as well as the possible adverse effects of such blockade in vivo, remain to be fully understood. In addition, xenograft models based on immunodeficient mice that lack NK, B, and T cells may not be sufficient for validation of the efficacy of agents that target human CD47 or SIRP α , given that the effects of such agents on immune cells in these mice are limited to myeloid cells such as macrophages and DCs. Immunodeficient mice engrafted with a human immune system by transplantation of human hematopoietic stem cells have been developed as an important new tool for cancer research.^{67–69} Future preclinical studies with such humanized and immunodeficient mice, as well as clinical trials, should show the potential of targeting the CD47-SIRP α axis as a new strategy for immunotherapy of human cancer.

ACKNOWLEDGMENTS

The work in the authors' laboratory was supported by a Grant-in-Aid for Scientific Research (B) from the Japan Society for the Promotion of Science (JSPS), by the Project for Development of Innovative Research on Cancer Therapeutics (P-DIRECT) and the Project for Cancer Research and Therapeutic Evolution (P-CREATE) of the Japan Agency for Medical Research and Development, by research funds from Daiichi Sankyo Co. Ltd, and by Terumo Foundation for Life Sciences and Arts, Uehara Memorial Foundation, Takeda Science Foundation, and Japanese Society of Hematology.

CONFLICTS OF INTEREST

T.M. has received research funding from Daiichi Sankyo Co. Ltd. The other authors declare no conflicts of interest.

ORCID

Yoji Murata  <http://orcid.org/0000-0002-9576-7030>

Takashi Matozaki  <http://orcid.org/0000-0002-4393-8416>

REFERENCES

- Hanahan D, Coussens LM. Accessories to the crime: functions of cells recruited to the tumor microenvironment. *Cancer Cell*. 2012;21:309–322.
- Quail DF, Joyce JA. Microenvironmental regulation of tumor progression and metastasis. *Nat Med*. 2013;19:1423–1437.
- Munn DH, Bronte V. Immune suppressive mechanisms in the tumor microenvironment. *Curr Opin Immunol*. 2015;39:1–6.
- Okazaki T, Chikuma S, Iwai Y, et al. A rheostat for immune responses: the unique properties of PD-1 and their advantages for clinical application. *Nat Immunol*. 2013;14:1212–1218.
- Callahan MK, Postow MA, Wolchok JD. Targeting T cell co-receptors for cancer therapy. *Immunity*. 2016;44:1069–1078.
- Alsaab HO, Sau S, Alzhrani R, et al. PD-1 and PD-L1 checkpoint signaling inhibition for cancer immunotherapy: mechanism, combinations, and clinical outcome. *Front Pharmacol*. 2017;8:561.
- Callahan MK, Wolchok JD. Clinical activity, toxicity, biomarkers, and future development of CTLA-4 checkpoint antagonists. *Semin Oncol*. 2015;42:573–586.
- Perez-Gracia JL, Labiano S, Rodriguez-Ruiz ME, et al. Orchestrating immune check-point blockade for cancer immunotherapy in combinations. *Curr Opin Immunol*. 2014;27:89–97.
- Matlung HL, Szilagyi K, Barclay NA, et al. The CD47-SIRP α signaling axis as an innate immune checkpoint in cancer. *Immunol Rev*. 2017;276:145–164.
- Weiskopf K. Cancer immunotherapy targeting the CD47/SIRP α axis. *Eur J Cancer*. 2017;76:100–109.
- Yanagita T, Murata Y, Tanaka D, et al. Anti-SIRP α antibodies as a potential new tool for cancer immunotherapy. *JCI Insight*. 2017;2:e89140.
- Noguchi T, Matozaki T, Fujioka Y, et al. Characterization of a 115-kDa protein that binds to SH-PTP2, a protein-tyrosine phosphatase with Src homology 2 domains, in chinese hamster ovary cells. *J Biol Chem*. 1996;271:27652–27658.
- Ohnishi H, Kubota M, Ohtake A, et al. Activation of protein-tyrosine phosphatase SH-PTP2 by a tyrosine-based activation motif of a novel brain molecule. *J Biol Chem*. 1996;271:25569–25574.
- Fujioka Y, Matozaki T, Noguchi T, et al. A novel membrane glycoprotein, SHPS-1, that binds the SH2-domain-containing protein tyrosine phosphatase SHP-2 in response to mitogens and cell adhesion. *Mol Cell Biol*. 1996;16:6887–6899.
- Kharitonov A, Chen Z, Sures I, et al. A family of proteins that inhibit signalling through tyrosine kinase receptors. *Nature*. 1997;386:181–186.
- Matozaki T, Murata Y, Okazawa H, et al. Functions and molecular mechanisms of the CD47-SIRP α signalling pathway. *Trends Cell Biol*. 2009;19:72–80.
- Barclay AN, Van den Berg TK. The interaction between signal regulatory protein alpha (SIRP α) and CD47: structure, function, and therapeutic target. *Annu Rev Immunol*. 2014;32:25–50.

18. Oldenborg PA. CD47: a cell surface glycoprotein which regulates multiple functions of hematopoietic cells in health and disease. *ISRN Hematol.* 2013;2013:614619.
19. Tsuda M, Matozaki T, Fukunaga K, et al. Integrin-mediated tyrosine phosphorylation of SHPS-1 and its association with SHP-2. Roles of Fak and Src family kinases. *J Biol Chem.* 1998;273:13223-13229.
20. Oldenborg PA, Zheleznyak A, Fang YF, et al. Role of CD47 as a marker of self on red blood cells. *Science.* 2000;288:2051-2054.
21. Okazawa H, Motegi S, Ohyama N, et al. Negative regulation of phagocytosis in macrophages by the CD47-SHPS-1 system. *J Immunol.* 2005;174:2004-2011.
22. Seiffert M, Cant C, Chen Z, et al. Human signal-regulatory protein is expressed on normal, but not on subsets of leukemic myeloid cells and mediates cellular adhesion involving its counterreceptor CD47. *Blood.* 1999;94:3633-3643.
23. Ohnishi H, Kaneko Y, Okazawa H, et al. Differential localization of Src homology 2 domain-containing protein tyrosine phosphatase substrate-1 and CD47 and its molecular mechanisms in cultured hippocampal neurons. *J Neurosci.* 2005;25:2702-2711.
24. Ishikawa-Sekigami T, Kaneko Y, Okazawa H, et al. SHPS-1 promotes the survival of circulating erythrocytes through inhibition of phagocytosis by splenic macrophages. *Blood.* 2006;107:341-348.
25. Okajo J, Kaneko Y, Murata Y, et al. Regulation by Src homology 2 domain-containing protein tyrosine phosphatase substrate-1 of α -galactosylceramide-induced antimetastatic activity and Th1 and Th2 responses of NKT cells. *J Immunol.* 2007;178:6164-6172.
26. Saito Y, Iwamura H, Kaneko T, et al. Regulation by SIRP α of dendritic cell homeostasis in lymphoid tissues. *Blood.* 2010;116:3517-3525.
27. Saito Y, Respatika D, Komori S, et al. SIRP α ⁺ dendritic cells regulate homeostasis of fibroblastic reticular cells via TNF receptor ligands in the adult spleen. *Proc Natl Acad Sci USA.* 2017;114:E10151-E10160.
28. Murata Y, Kotani T, Ohnishi H, et al. The CD47-SIRP α signalling system: its physiological roles and therapeutic application. *J Biochem.* 2014;155:335-344.
29. Murata Y, Saito Y, Kaneko T, et al. Autoimmune animal models in the analysis of the CD47-SIRP α signaling pathway. *Methods.* 2014;65:254-259.
30. Ohnishi H, Murata T, Kusakari S, et al. Stress-evoked tyrosine phosphorylation of signal regulatory protein α regulates behavioral immobility in the forced swim test. *J Neurosci.* 2010;30:10472-10483.
31. Takenaka K, Prasolava TK, Wang JC, et al. Polymorphism in Sirpa modulates engraftment of human hematopoietic stem cells. *Nat Immunol.* 2007;8:1313-1323.
32. Oldenborg PA, Gresham HD, Lindberg FP. CD47-signal regulatory protein α (SIRP α) regulates Fc γ and complement receptor-mediated phagocytosis. *J Exp Med.* 2001;193:855-862.
33. Eckstein N, Roper L, Haas B, et al. Clinical pharmacology of tyrosine kinase inhibitors becoming generic drugs: The regulatory perspective. *J Exp Clin Cancer Res.* 2014;33:15.
34. Holderfield M, Deuker MM, McCormick F, et al. Targeting RAF kinases for cancer therapy: BRAF-mutated melanoma and beyond. *Nat Rev Cancer.* 2014;14:455-467.
35. Wu P, Nielsen TE, Clausen MH. FDA-approved small-molecule kinase inhibitors. *Trends Pharmacol Sci.* 2015;36:422-439.
36. Caunt CJ, Sale MJ, Smith PD, et al. MEK1 and MEK2 inhibitors and cancer therapy: the long and winding road. *Nat Rev Cancer.* 2015;15:577-592.
37. O'Donnell JS, Massi D, Teng MWL, et al. PI3K-AKT-mTOR inhibition in cancer immunotherapy, redux. *Semin Cancer Biol.* 2018;48:91-103.
38. Braster R, O'Toole T, van Egmond M. Myeloid cells as effector cells for monoclonal antibody therapy of cancer. *Methods.* 2014;65:28-37.
39. Jaiswal S, Jamieson CH, Pang WW, et al. CD47 is upregulated on circulating hematopoietic stem cells and leukemia cells to avoid phagocytosis. *Cell.* 2009;138:271-285.
40. Majeti R, Chao MP, Alizadeh AA, et al. CD47 is an adverse prognostic factor and therapeutic antibody target on human acute myeloid leukemia stem cells. *Cell.* 2009;138:286-299.
41. Chao MP, Weissman IL, Majeti R. The CD47-SIRP α pathway in cancer immune evasion and potential therapeutic implications. *Curr Opin Immunol.* 2012;24:225-232.
42. Brown EJ, Frazier WA. Integrin-associated protein (CD47) and its ligands. *Trends Cell Biol.* 2001;11:130-135.
43. Chao MP, Alizadeh AA, Tang C, et al. Anti-CD47 antibody synergizes with rituximab to promote phagocytosis and eradicate non-Hodgkin lymphoma. *Cell.* 2010;142:699-713.
44. Willingham SB, Volkmer JP, Gentles AJ, et al. The CD47-signal regulatory protein α (SIRP α) interaction is a therapeutic target for human solid tumors. *Proc Natl Acad Sci USA.* 2012;109:6662-6667.
45. Zhao XW, van Beek EM, Schornagel K, et al. CD47-signal regulatory protein- α (SIRP α) interactions form a barrier for antibody-mediated tumor cell destruction. *Proc Natl Acad Sci USA.* 2011;108:18342-18347.
46. Chao MP, Alizadeh AA, Tang C, et al. Therapeutic antibody targeting of CD47 eliminates human acute lymphoblastic leukemia. *Cancer Res.* 2011;71:1374-1384.
47. Liu J, Wang L, Zhao F, et al. Pre-clinical development of a humanized anti-CD47 antibody with anti-cancer therapeutic potential. *PLoS ONE.* 2015;10:e0137345.
48. Weiskopf K, Jahchan NS, Schnorr PJ, et al. CD47-blocking immunotherapies stimulate macrophage-mediated destruction of small-cell lung cancer. *J Clin Invest.* 2016;126:2610-2620.
49. Gholamin S, Mitra SS, Feroze AH, et al. Disrupting the CD47-SIRP α anti-phagocytic axis by a humanized anti-CD47 antibody is an efficacious treatment for malignant pediatric brain tumors. *Sci Transl Med.* 2017;9:eaaf2968.
50. Weiskopf K, Ring AM, Ho CC, et al. Engineered SIRP α variants as immunotherapeutic adjuvants to anticancer antibodies. *Science.* 2013;341:88-91.
51. Tseng D, Volkmer JP, Willingham SB, et al. Anti-CD47 antibody-mediated phagocytosis of cancer by macrophages primes an effective antitumor T-cell response. *Proc Natl Acad Sci USA.* 2013;110:11103-11108.
52. Liu X, Pu Y, Cron K, et al. CD47 blockade triggers T cell-mediated destruction of immunogenic tumors. *Nat Med.* 2015;21:1209-1215.
53. Xu MM, Pu Y, Han D, et al. Dendritic cells but not macrophages sense tumor mitochondrial DNA for cross-priming through signal regulatory protein α signaling. *Immunity.* 2017;47:363-373.e5.
54. Ho CC, Guo N, Sockolosky JT, et al. "Velcro" engineering of high affinity CD47 ectodomain as signal regulatory protein α (SIRP α) antagonists that enhance antibody-dependent cellular phagocytosis. *J Biol Chem.* 2015;290:12650-12663.
55. Chiba S, Ikushima H, Ueki H, et al. Recognition of tumor cells by Dectin-1 orchestrates innate immune cells for anti-tumor responses. *Elife.* 2014;3:e04177.
56. Mattioli I, Pesant M, Tentorio PF, et al. Priming of human resting NK cells by autologous M1 macrophages via the engagement of IL-1 β , IFN- β , and IL-15 pathways. *J Immunol.* 2015;195:2818-2828.
57. Bodduluru LN, Kasala ER, Madhana RM, et al. Natural killer cells: the journey from puzzles in biology to treatment of cancer. *Cancer Lett.* 2015;357:454-467.
58. Lin GHY, Chai V, Lee V, et al. TTI-621 (SIRP α Fc), a CD47-blocking cancer immunotherapeutic, triggers phagocytosis of lymphoma cells by multiple polarized macrophage subsets. *PLoS ONE.* 2017;12:e0187262.
59. Petrova PS, Viller NN, Wong M, et al. TTI-621 (SIRP α Fc): a CD47-blocking innate immune checkpoint inhibitor with broad antitumor

- activity and minimal erythrocyte binding. *Clin Cancer Res.* 2017;23:1068-1079.
60. Sockolosky JT, Dougan M, Ingram JR, et al. Durable antitumor responses to CD47 blockade require adaptive immune stimulation. *Proc Natl Acad Sci USA.* 2016;113:E2646-E2654.
61. Ingram JR, Blomberg OS, Sockolosky JT, et al. Localized CD47 blockade enhances immunotherapy for murine melanoma. *Proc Natl Acad Sci USA.* 2017;114:10184-10189.
62. Tao H, Qian P, Wang F, et al. Targeting CD47 enhances the efficacy of anti-PD-1 and CTLA-4 in an esophageal squamous cell cancer pre-clinical model. *Oncol Res.* 2017;25:1579-1587.
63. Herndler-Brandstetter D, Shan L, Yao Y, et al. Humanized mouse model supports development, function, and tissue residency of human natural killer cells. *Proc Natl Acad Sci USA.* 2017;114:E9626-E9634.
64. Ring NG, Herndler-Brandstetter D, Weiskopf K, et al. Anti-SIRP α antibody immunotherapy enhances neutrophil and macrophage antitumor activity. *Proc Natl Acad Sci USA.* 2017;114:E10578-E10585.
65. Strowig T, Rongvaux A, Rathinam C, et al. Transgenic expression of human signal regulatory protein alpha in Rag2^{-/-} γ c^{-/-} mice improves engraftment of human hematopoietic cells in humanized mice. *Proc Natl Acad Sci USA.* 2011;108:13218-13223.
66. Murata Y, Tanaka D, Hazama D, et al. Anti-human SIRP α antibody is a new tool for cancer immunotherapy. *Cancer Sci.* 2018;109:1300-1308.
67. Rongvaux A, Takizawa H, Strowig T, et al. Human hemato-lymphoid system mice: current use and future potential for medicine. *Annu Rev Immunol.* 2013;31:635-674.
68. Theocharides AP, Rongvaux A, Fritsch K, et al. Humanized hemato-lymphoid system mice. *Haematologica.* 2016;101:5-19.
69. Morton JJ, Bird G, Refaeli Y, et al. Humanized mouse xenograft models: narrowing the tumor-microenvironment gap. *Cancer Res.* 2016;76:6153-6158.

How to cite this article: Murata Y, Saito Y, Kotani T, Matozaki T. CD47-signal regulatory protein α signaling system and its application to cancer immunotherapy. *Cancer Sci.* 2018;109:2349-2357. <https://doi.org/10.1111/cas.13663>