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Social wasps, crickets and cockroaches contribute to pollination of the holoparasitic plant Mitrastemon yamamotoi (Mitrastemonaceae) in southern Japan

Suetsugu, Kenji

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| 1 | Social wasps, crickets, and cockroaches contribute to the pollination of the |
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| 2 | holoparasitic plant Mitrastemon yamamotoi (Mitrastemonaceae) in southern Japan |
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| 4 | KENJI SUETSUGU ^{1,*} |
| 5 | |
| 6 | ¹ Department of Biology, Graduate School of Science, Kobe University, 1-1 Rokkodai, |
| 7 | Nada-ku, Kobe, 657-8501, Japan |
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| 9 | Running title: Social wasps, crickets, and cockroaches pollination |
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| 11 | Correspondence K. Suetsugu, Department of Biology, Graduate School of Science, |
| 12 | Kobe University, 1-1 Rokkodai, Nada-ku, Kobe, 657-8501, Japan. E-mail: |
| 13 | kenji.suetsugu@gmail.com |
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1 ABSTRACT

Mitrastemon yamamotoi is completely embedded within the tissues of its hosts, except
during the reproductive stage, when aboveground parts emerge from the host tissues. Its
highly modified appearance has attracted the attention of many botanists, but very little
is known about *M. yamamotoi* its reproductive system.

Floral visitors of *M. yamamotoi* were observed in southern Japan. Pollination
experiments were conducted to determine the plant's self-compatibility and pollen
limitation, as well as the contribution of diurnal and nocturnal visitors to fruit set and
outcrossing.

Mitrastemon yamamotoi was mainly pollinated by social wasps, but previously
 unnoticed pollinators (i.e., crickets and cockroaches) are also important, based on
 visitation frequency and pollen loads. The results of the pollination experiments
 suggested that nocturnal visitors, such as crickets and cockroaches, contribute to
 geitonogamous pollination, whereas diurnal visitors, such as social wasps, facilitate
 outcrossing.

• The unexpected pollinator assemblage of *M. yamamotoi* might be influenced by multiple factors, including the highly modified flowers that are produced close to the ground in dark understory environments, the species' winter-flowering habit, and the location of the study site (i.e., near the northern limit of the species' range). Considering that *M. yamamotoi* occurs widely in subtropical and tropical forests in Asia, additional studies are needed to assess the pollinator assemblages of *M. yamamotoi* at other locations.

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24 Keywords: Achlorophyllous plant; breeding system; dung beetle; parasitic plant;

 $\mathbf{2}$

1 heterotrophic plant; reproductive biology

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Non-photosynthetic plants have long attracted interest because of their peculiar 3 morphological features (Kuijt 1969). The genus Mitrastemon, the sole member of 4 5 Mitrastemonaceae, includes two to six holoparasitic species and has diversification 6 centers in Southeast Asia and Central America. It was formerly classified in 7 Rafflesiaceae together with the members of Apodanthaceae and Cytinaceae, whose vegetative parts are completely embedded within the host tissues except during the 8 9 reproductive period, due to their unusual lifestyles (de Vega et al. 2007). However, based on recent phylogenetic studies, the former Rafflesiaceae was split into four 10 11 families belonging to four different orders (Barkman et al. 2004; Nickrent et al. 2004; 12Barkman et al. 2007; Filipowicz & Renner 2010). Barkman et al. (2004) revealed that 13the genus *Mitrastemon* belongs to the order Ericales, while Rafflesiaceae itself belongs 14 to the order Malpighiales, based on mitochondrial markers. Mitrastemon yamamotoi 15occurs naturally in the subtropical or tropical forests of Borneo, Sumatra, Indochina, and Japan, while the debate on Mitrastemon species delimitation remains unsettled 16(Meijer & Velkamp 1993). 17

The extraordinary appearance of *Mitrastemon* species has attracted the interest of many botanists, but very little is known about its their reproductive systems. For example, some birds (e.g., *Zosterops, Melidectes*, and *Oedistoma* species) have been reported to forage for nectar among *M. yamamotoi* flowers (Matuda 1947; Beehler 1994). However, no studies have confirmed the effectiveness of birds as pollinators. Beehler (1994) also hypothesized that small nocturnal mammals could pollinate *M. yamamotoi* flowers. However, nocturnal visitors have never been formally observed.

The pollinator assemblage of *M. yamamotoi* is likely influenced by the location of 1 $\mathbf{2}$ its flowers. These are produced close to the ground in dark understory environments, which are generally associated with pollinator species different from that found at open 3 sites (Kato 1996; Herrera 1997; Moore 1997; Rincon et al. 1999). Achlorophyllous 4 5 plants, including M. yamamotoi, can occupy low-light niches where there is little 6 competition from autotrophic plants. However, such conditions can inhibit reproduction 7 if pollinator foraging is negatively influenced by low light intensity. Most bees, for example, tend to restrict their foraging activities to areas of high light intensity. 8 9 Consequently, it is possible that plants in shaded understory habitats experience less pollination by bees (Kato 1996; Suetsugu 2015; but also see Hentrich et al. 2010). In 10 11 fact, most mycoheterotrophs studied to date appear to have abandoned bee pollinators in favor of self-pollination or alternative understory pollinators, such as fruit flies 1213(Suetsugu 2013; Martos et al. 2015; Suetsugu 2015, 2018a). In addition, unexpected 14 seed dispersal systems, such as endozoochory by camel crickets (Rhaphidophoridae), 15have been reported in non-photosynthetic plants, possibly due to the colonization of dark understory habitats, where wind is an ineffective seed dispersal agent (Suetsugu 162018b,c). Judging from the complex and intriguing reproductive systems of 17non-photosynthetic plants growing in dark understory environments, the pollination 1819 biology of *Mitrastemon* species is likely unusual.

Notably, *M. yamamoto*i blooms from late autumn to early winter in Japan. Matuda (1947) reported that *Mitrastemon* species only grow under cool, dry conditions, when its hosts are relatively inactive. Outside the tropics, winter-flowering plants may experience pollinator limitation, as insect activity is largely limited by temperature (Fang *et al.* 2012). Therefore, several studies have suggested that winter-flowering

phenologies favor bird pollination in temperate regions (Kunitake *et al.* 2004; Fang *et al.* 2012).

Here, I conducted intensive field observations and pollination experiments to 3 examine the pollination biology and reproductive system of M. yamamotoi in a 4 5 warm-temperate region, i.e., Yakushima Island, southern Japan. I hypothesized that 6 avian visitors are the primary pollinators of *M. yamamotoi* in this population because 7 previous studies reported birds foraging for nectar in *M. yamamotoi* (Matuda 1947; Beehler 1994; Hansen 1972). However, the results obtained here indicated that 8 9 relatively unnoticed floral visitors, including social wasps, crickets, and cockroaches, 10 contribute significantly to fruit set.

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12 MATERIALS AND METHODS

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14 Study species and study site

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16Mitrastemon yamamotoi is a holoparasitic plant that is completely embedded within the tissues of its hosts, except during the reproductive stage, when its flowers emerge from 1718 the host tissues. Mitrastemon species produce bisexual, protandrous flowers with 19 collar-shaped perianth tubes (Fig. 1). The stamens of the flowers are connate, forming a 20mitra-shaped androecial tube that is crowned by a fertile zone of pollen-bearing locules (Nickrent et al. 2004), and the staminal tube, which has a small hole at the top, 21circumscissily separates from the flower as it is pushed up by the growing gynoecium. 22Furthermore, the apical portion of the staminal tube is sterile, whereas the basal portion 23possesses a series of vertical rings of numerous, minute pollen sacs. The gynoecia of the 24

flowers are hypogynous and single-locular, with a thick, conical stigma (Nickrent et al. 1 $\mathbf{2}$ 2004), and the large amount of dilute nectar stored in upper scale leaves has been considered a pollinator reward (Matuda 1947; Beehler 1994). 3

The floral biology of *M. yamamotoi* was investigated in the southern part of 4 $\mathbf{5}$ Yakushima Island, Kyushu district, Japan, where M. yamamotoi parasitizes the roots of 6 Castanopsis sieboldii (Fagaceae).

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8 Pollinator observation

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10 Pollination observations were conducted from late October to late November from 2008 11 to 2011. Direct observations were made for ca. 100 h in total, in 4- to 6-h bouts, which 12were scheduled to cover all hours within a day. The behavior of potential visitors was observed by walking around the study site, sitting next to flower patches, or hiding in 1314 the vegetation near (1-2 m) flower clusters. Nocturnal observations involved the use of 15red lamps, which minimized the effect of light on potential floral visitors. The 16frequency, duration, and visitation pattern (single or sequential) were recorded for each floral visitor, i.e., for individuals landing on, passing the floral patches, or foraging for 17floral nectar or pollen grains. 18

A subset of the observed floral visitors (at least one species from each order) 1920was captured using a sweep net or aspirator. Pollen grains carried on the bodies of visitors were counted under a dissecting microscope. Those forming clumps were first 21removed from visitor bodies using basic fuchsine jelly (Kearns & Inouye 1993), and 22then spread on glass slides. 23

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From November 7 to 21 in 2011, a remote camera with built-in infrared motion

sensors (Sensor Camera Fieldnote; Marif Co., Ltd., Japan) was also used at all hours of the day and night to record any additional potential floral visitors that might have been deterred by our direct observation (e.g., birds and mammals). This camera was set ca. 2 m from the target flowering plants. The observation by the remote camera was used only for detecting visitations by mammals and birds. In contrast, I used the data based on direct observations for visitations by invertebrates.

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8 Breeding system

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Pollination experiments were conducted in early November 2008 to determine the 10 11 breeding system (i.e., the capacity for self-fertilization) of M. yamamotoi. Flower buds 12were bagged using nylon netting and subject to one of four treatments: (1) bagged only 13to exclude floral visitors (40 flowers; autonomous selfing); (2) self-pollinated by hand 14 as soon as each flower's androecial tube became circumscissile and dehiscent (40 15flowers; artificial self-pollination treatment); (3) cross-pollinated by hand as soon as each flower's androecial tube became circumscissile and dehiscent. To avoid crossing 16genetically identical plants, all plants used for the cross-pollination experiments were 17located at least 5 m from their nearest neighbor (40 flowers; artificial cross-pollination 1819 treatment); or (4) the nylon netting had small openings around the stems to allow 20visitation by ants (40 flowers; ants-only treatment). In addition, 100 flowers were 21marked to analyze fruit set under natural conditions (open treatment).

When all stigmas became blackish and unreceptive, the plants in each treatment group were bagged using nylon-mesh cages to prevent seed consumption by potential seed dispersers and facilitate a precise determination of fruit set. Fruit number was

counted at ca. 2–3 months after the flowering season, and the effect of treatment was determined using Fisher's exact test. In addition, 100 randomly selected seeds from each fruit capsule were examined under a dissecting microscope, to assess the proportion of seeds with embryos, and the effect of treatment was determined using analysis of variance (ANOVA).

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7 Effectiveness of diurnal and nocturnal visitors

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9 The effectiveness of diurnal and nocturnal visitors was evaluated in November 2011. Based on the observations from 2008 to 2010, most nocturnal visitors were flightless. 10 11 The visitors likely contribute to pollination within the same patches. Furthermore, 12because the endophytic system of *M. yamamotoi* grows intercellularly within host roots, it is highly possible that *M. yamamotoi* flowers on the same host plant are genetically 1314 identical. Therefore, the effectiveness of outcrossing pollinators in both diurnal and 15nocturnal visitors was examined by emasculating all M. yamamotoi flowers that parasitized a single tree, as these were considered genetically identical. 16

Patches with more than 20 individuals parasitizing the same tree with unopened 17flower buds were selected before anthesis, and, within each patch, flowers were 1819 assigned to one of four treatment groups. Five patches were used per treatment: (1) 20nocturnal visitors were excluded from sunset to sunrise using a nylon-mesh cage (110 flowers; diurnal natural pollination); (2) diurnal visitors were excluded from sunrise to 21sunset using a nylon-mesh cage (110 flowers; nocturnal natural pollination); (3) 22nocturnal pollinators were excluded from emasculated flowers from sunset to sunrise 23using a nylon-mesh cage (112 flowers; diurnal cross-pollination); or (4) diurnal 24

pollinators were excluded from emasculated flowers from sunrise to sunset using a
nylon-mesh cage (112 flowers; nocturnal cross-pollination).

After all the stigmas had become blackish and unreceptive, the plants in each 3 treatment group were bagged using nylon-mesh cages, in order to prevent seed 4 $\mathbf{5}$ consumption by potential seed dispersers and facilitate a precise determination of fruit 6 set. Fruit number was counted at ca. 2–3 months after the flowering season, and the 7 effect of treatment was determined using Fisher's exact test. In addition, 10 fruit capsules were randomly collected from each treatment group. After that, 100 randomly 8 9 selected seeds from each of these fruit capsules were examined under a dissecting microscope, to assess the proportion of seeds with embryos, and the effect of treatment 10 11 was determined using ANOVA.

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13 RESULTS

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15 Observations of floral visitors

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Recordings by the motion-sensor-equipped camera showed that *Zosterops japonicus*(Zosteropidae) rarely landed on *M. yamamotoi* patches and never foraged for nectar. In
contrast, direct observation showed that *M. yamamotoi* flowers were frequently visited
by social wasps, crickets, cockroaches, flies, dung beetles, stag beetles, and ants (Fig. 2;
Table 1).

Social wasps (e.g., *Vespa mandarinia*, *Vespa analis*, *Vespa simillima xanthoptera*, *Vespula flaviceps*, and *Vespula shidai*) were the main diurnal visitors and were observed
to sequentially visit multiple plants during foraging. Multiple wasps visited the same *M*.

yamamotoi patches simultaneously and they were often observed to aggressively defend
their feeding territories from one another. Furthermore, even though social wasps did
not actively collect pollen grains from the *M. yamamotoi* flowers, these were always
attached to the wasps' legs, heads, abdomens, and mouthparts (Table 2).

5 Other diurnal visitors included flies, such as *Drosophila* species and members of 6 the Calliphoridae, Sarcophagidae, Muscidae, and Tachinidae, and ants, such as 7 Crematogaster species. Flies sometimes visited multiple plants within the patches, at a single visitation, while foraging for nectar. However, despite touching the anthers and 8 9 stigmas of the flowers with their legs and mouthparts, pollen grains were rarely found 10 attached to either (Table 2). Ants also visited *M. yamamotoi* flowers to harvest nectar, 11 but usually only visited single flowers, where they stayed for a long time, and rarely 12made successive visits to multiple flowers.

Nocturnal visitors included various crane fly, orthopteran, cockroach, beetle, 1314 centipede, and moth species. Crane fly Limonia sp. occasionally visited M. yamamotoi 15flowers to lay eggs but only oviposited on the plants' scale-leaves and rarely touched 16either the male or female flowers. However, orthopterans, such as camel crickets (Diestrammena yakumontana) and field crickets (Duolandrevus ivani), which were 17some of the most frequent nocturnal floral visitors, usually visited multiple plants 18sequentially, touched anthers and stigmas during feeding, and, consequently, carried 1920pollen grains on their legs, heads, and mouthparts (Table 2). Similarly, cockroaches, such as Opisthoplatia orientalis and Onychostylus pallidiolus, were also observed to 21visit multiple plants sequentially, touch anthers and stigmas, and carry pollen grains 22(Table 2). 23

The dung beetle Onthophagus yakuinsulanus often visited multiple flowers to

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forage for nectar and pollen grains, and specimens sometimes exhibited pollen grains 1 $\mathbf{2}$ attached to their legs and abdomens. The stag beetle *Aegus laevicollis* visited multiple plants to forage for nectar, and pollen grains were also found attached to their legs and 3 abdomens (Table 2). The centipedes Thereuopoda clunifera and Scolopendra 4 $\mathbf{5}$ subspinipes were observed to visit M. yamamotoi flowers, possibly to feed on nectar or 6 prey on other animals attracted by the nectar, and pollen grains were sometimes found 7 attached to their legs. Finally, an unidentified pyralid moth was observed to land on flowers and to oviposit on the surface of scale-leaves. 8

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10 Breeding system

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Flowers in the pollinator-excluded treatment showed low levels of fruit set (Table 3). 1213The fruit set observed in bagged flowers was unexpected; the stamen tube was pushed 14 off by the growth of the pistil after the pollen had been shed and, therefore, its floral 15structure should have prevented self-fertilization. Low levels of fruit set in bagged flowers were possibly due to unintended ant intrusion or apomixis. There were more 16developed fruits in the ants-only treatment than in the bagged-only treatment, although 17the difference was insignificant (P = 0.59). High fruit set and seed viability were 18 19 observed in both the artificial self-pollination and cross-pollination groups, whereas the 20fruit set and seed viability of the open-pollinated group were significantly lower thereby indicating some pollinator limitation (Table 3). 21

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23 Effectiveness of diurnal and nocturnal visitors

In both the diurnal and nocturnal treatment groups, approximately half of the flowers 1 $\mathbf{2}$ developed fruit capsules, and there were no significant differences between the fruit set and seed viability of the two groups (P = 0.50). This indicated that both diurnal and 3 nocturnal floral visitors contributed to pollination (Table 4). Although seed viability was 4 $\mathbf{5}$ similar for nocturnal and diurnal cross-pollination groups, (P = 0.41), fruit set in the 6 nocturnal cross-pollination group was significantly lower than that in the diurnal cross-pollination group (P < 0.01). This indicated that the relative contribution of 7 diurnal cross-pollinators to fruit set was higher than that of nocturnal cross-pollinators. 8

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10 DISCUSSION

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The findings of the present study indicate that unexpected floral visitors, such as social 1213wasps, crickets, and cockroaches, contribute significantly to M. yamamotoi fruit set. 14 This unexpected pollinator assemblage seems to be influenced by multiple factors, 15including the highly modified flowers produced close to the ground in dark understory environments and the winter-flowering habit of M. yamamotoi, as well as the location of 16the study site (i.e., near the northern limit of the species' range). In addition, pollination 17experiments suggested diurnal and nocturnal pollinators have a differential contribution 18to reproductive success. While there were no significant differences in fruit set and seed 1920viability between the diurnal- and nocturnal-pollinators treatment groups, fruit set in the nocturnal cross-pollination group was significantly lower than that in the diurnal 21cross-pollination group. These results indicate that nocturnal visitors, such as crickets, 22cockroaches, and dung beetles, mainly contributed to the geitonogamous pollination. 23

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The potential factor contributing to the observed difference is that most nocturnal

visitors were flightless arthropods. It is intriguing that endemic flightless insects such as *Diestrammena yakumontana* and *Onthophagus yakuinsulanus* appear to contribute to
fruit set of *M. yamamotoi* in Yakushima Island. Flightless insects might outcompete
other species on island environments because mainland predators are often absent
(Carlquist 1974). Thus, pollination by flightless insects might represent unique
pollination biology in island ecosystems, since Yakushima Island is known to form a
unique ecosystem harboring many endemic taxa (Yahara *et al.* 1987).

In the present study, social wasps were the main diurnal visitors of *M. yamamotoi*, 8 9 and both visitor observation and pollination experiments suggested that social wasps are 10 the most effective pollinators of *M. yamamotoi*, as they transfer pollen grains between 11 patches. Because Vespa species do not harvest pollen to feed their brood (Richter 2000), 12wasps are not considered typical floral visitors (Brodmann et al. 2008, 2009). However, 13social wasps often feed on nectar to fuel their own activity (Richter 2000). In addition, 14 Vislobokov & Galinskaya (2018) reported that wasps are the main pollinators of the 15holoparasitic plant Balanophora harlandii, which occupies an ecological niche that is 16similar to that of *Mitrastemon* species.

Although both ants and flies were main diurnal visitors, and occasionally visited multiple plants within patches during their nectar foraging, the pollen loads on their bodies were much lower than those found for social wasps, crickets, and cockroaches. Therefore, their effectiveness as pollinators was lower than that of other frequent visitors. This result was also supported by the pollination experiment results, as there were no significant differences on fruit set between the ants-only treatment and the bagged-only treatment.

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Some orthopteran species were the most frequent nocturnal visitors. Orthopterans

are generally considered herbivores, rather than pollinators (Suetsugu & Tanaka 2014; 1 $\mathbf{2}$ Tan et al. 2017). In the present study, D. yakumontana and D. ivani consumed pollen grains and nectar of M. yamamotoi, but did not damage the other floral parts such as 3 stigmas. In addition, they often visited both male and female flowers sequentially to 4 5 forage for nectar. Consequently, they had many pollen grains attached to their bodies, 6 which clearly suggests that orthopterans can pollinate *Mitrastemon* species, even though 7 the relationship between *Mitrastemon* and crickets is unspecialized. These results, along those of previous studies (Micheneau et al. 2010; Tan & Tan 2018), suggest that 8 9 orthopteran pollination might be more common than previously recognized.

10 Cockroaches were also among the most common nocturnal visitors. Cockroaches 11 are generally omnivorous scavengers and detritus feeders (Schal et al. 1984). However, 12cockroach pollination has been reported in at least three plant species, including Uvaria 13Nagamitsu & Inoue 1997), Balanophora tobiracola elmeri (Annonaceae; 14 (Balanophoraceae; Kawakita & Kato 2002), and Clusia sellowiana (Clusiaceae; 15Vlasáková et al. 2008). Here, cockroaches visited multiple flowers and had numerous 16pollen grains attached to their bodies, thereby confirming their pollinator status.

The dung beetle Onthophagus yakuinsulanus often visited multiple flowers in 17succession and carried pollen grains (Table 2). Indeed, Onthophagus species have been 18reported to pollinate other plant species, such as Orchidantha inouei (Lowiaceae) in 1920Sarawak, Malaysia (Sakai & Inoue 1999). Because some dung beetles are excellent dung searchers and fly long distances to locate specific types of dung, Sakai & Inoue 21(1999) considered that they could function as long-distance pollinators. However, the 22elytra of O. yakuinsulanus are fused, thereby forming a closed carapace and rendering 23the species flightless. Nevertheless, based on its visitation behavior and ability to carry 24

1 pollen, *O. yakuinsulanus* can at least work as a geitonogamous pollinator.

 $\mathbf{2}$ It is intriguing that most *M. yamamotoi* visitors included social wasps, crickets, and cockroaches, which are known to forage for foods such as fermented sap, using 3 olfactory cues (Yoshimoto et al. 2005; Brodmann et al. 2008; Dormont et al. 2010; 4 5 Micheneau et al. 2010; Stökl et al. 2010). Because the scale-leaves of M. yamamotoi 6 store nectar that sometimes smells fermented, the unique assembly of M. yamamotoi 7 pollinators might be attracted by volatiles produced by nectarivorous yeasts. As these yeasts degrade floral nectar by metabolizing nectar sugar, they are often regarded as 8 9 exploitative antagonists of plant-pollinator mutualistic relationships (Herrera et al. 10 2008). However, nectarivorous yeasts may enhance pollination by altering volatile 11 profiles emitted from flowers (Rering et al. 2018). Thus, how nectar-dwelling yeasts 12affect *M. yamamotoi* pollination success should be addressed in future studies.

It is puzzling that no birds visited *M. yamamotoi* flowers for nectar, especially 1314 because Zosterops, Melidectes, and Oedistoma species have been previously reported to 15forage for nectar in *M. yamamotoi* patches (Matuda 1947, Beehler 1994) and because *M*. 16yamamotoi produces large amounts of dilute nectar that is well suited for avian pollination (Cronk & Ojeda 2008). However, preliminary observations suggested that Z. 17japonicus, Turdus pallidus, and Erithacus akahige only visit M. yamamotoi during the 18 fruiting season (Suetsugu, unpublished data). Therefore, records of bird visitation might 1920have been incorrectly attributed to floral visitation, rather than visitation during the fruiting season, at least in Japanese populations. Yet, Beehler (1994) reported that 21Melidectes and Oedistoma species aggressively and frequently forage for nectar among 22M. yamamotoi flowers in New Guinea. 23

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It should be noted that the investigated population is near the northern limit of *M*.

yamamotoi distribution. Because pollinator assemblages can vary among locations (e.g., 1 Aguiar et al. 2012), other M. yamamotoi populations might be ecologically distinct and $\mathbf{2}$ utilize different pollinators, including birds. Therefore, further research is needed to 3 elucidate the pollinator assemblage of *M. yamamotoi* in other areas. In addition, the 4 $\mathbf{5}$ discovery of pollination by camel crickets and cockroaches suggests that pollination 6 systems involving unusual and unexpected taxa might be more widespread than 7 previously thought, especially in non-photosynthetic plants with highly modified floral morphology. 8

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| | | Number | 0 |
|-----------------------------------|-----------------------------|-------------|----|
| Order | Species | individuals | or |
| | | flowers | |
| Blattodea | Opisthoplatia orientalis | 67 | |
| | Onychostylus pallidiolus | 35 | |
| Coleoptera | Onthophagus yakuinsulanus | 21 | |
| | Allecula fuliginosa | 2 | |
| | Aegus laevicollis | 1 | |
| Diptera | Drosophila spp. | >200 | |
| | Calliphoridae spp. | >200 | |
| | Sarcophagidae spp. | >100 | |
| | Muscidae spp. | >100 | |
| | Tachinidae spp. | >200 | |
| | Limonia sp. | 13 | |
| | Mycetophilidae spp. | 9 | |
| Hymenoptera | Vespa analis | 183 | |
| | Vespa mandarinia | 56 | |
| | Vespa simillima xanthoptera | 38 | |
| | Vespula flaviceps | 24 | |
| | Vespula shidai | 11 | |
| | Paratrechina flavipes | >100 | |
| | Crematogaster spp. | >100 | |
| | Camponotus sp. | 15 | |
| | Odontomachus monticola | 3 | |
| Lepidoptera | Pyralidae sp. | 4 | |
| Orthoptera | Diestrammena yakumontana | u 89 | |
| | Duolandrevus ivani | 41 | |
| | Aphonoides rufescens | 8 | |
| | Ornebius kanetataki | 5 | |
| Scutigeromorpha | Thereuopoda clunifera | 10 | |
| Chilopoda Scolopendra subspinipes | | 14 | |

Table 1. Floral visitors of Mitrastemon yamamotoi.

| | | Number of pollen | Number of |
|-----------------|----------------------------------|-------------------|-----------|
| Order | Species | grains on visitor | |
| order | Species | bodies | examined |
| Blattodea | Opisthoplatia orientalis | 78.8 ± 32.9 | n = 5 |
| | Onychostylus pallidiolus | 19.4 ± 8.6 | n = 5 |
| Onthophagus | | 12.8 ± 7.7 | n = 5 |
| Diptera | Drosophila spp. | 2.6 ± 1.1 | n = 20 |
| | Calliphoridae spp. | 3.6 ± 1.5 | n = 20 |
| | <i>Limonia</i> sp. | 0 | n = 5 |
| Hymenoptera | Vespa analis | 118.2 ± 27.2 | n = 5 |
| | Vespa mandarinia | 156.4 ± 49.2 | n = 5 |
| | Vespa simillima xanthoptera | 71.0 ± 17.0 | n = 5 |
| | Paratrechina flavipes | 3.0 ± 1.2 | n =20 |
| Lepidoptera | Pyralidae sp. | 0 | n = 2 |
| Orthoptera | Diestrammena yakumontana | 66.6 ± 31.0 | n = 5 |
| | Duolandrevus ivani | 78.8 ± 26.8 | n = 5 |
| Scutigeromorpha | Thereuopoda clunifera | 9.7 ± 5.2 | n = 3 |
| Chilopoda | hilopoda Scolopendra subspinipes | | n = 3 |

Table 2. Number of Mitrastemon yamamotoi pollen grains on the bodies of floral visitors

The number of pollen grains on the bodies is indicated as mean \pm standard error.

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 $\mathbf{7}$

Table 3. Effect of pollination treatments on the proportions (%) of fruit set and seeds with embryo in *Mitrastemon yamamotoi*.

| | Pollinator-excluded | Manual | Manual | Ants-only | Open | |
|------------|-------------------------|-------------------------|-------------------------|-------------------|--------------------|--|
| | Formator-excluded | autogamy | allogamy | Ants-only | | |
| Fruit set | 17.5 ^a | 67.5 ^b | 65.0 ^b | 25.0 ^a | 43.0 ^{ab} | |
| Seeds with | 18.0 ± 12.0^{a} | 60.5 ± 20.8^{b} | 67.2 ± 21.5^{b} | 27.7 ± | 48.3 ± | |
| embryo | $16.0 \pm 12.0^{\circ}$ | $00.3 \pm 20.8^{\circ}$ | $07.2 \pm 21.3^{\circ}$ | 15.7 ^a | 21.3 ^c | |

Pollination treatments producing significant differences (P < 0.05) are indicated by different superscript letters.

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Table 4. Effect of diurnal and nocturnal visitors on the proportions (%) of fruit

set and seeds with embryo in Mitrastemon yamamotoi.

| | Diurnal | Nocturnal | Diurnal | Nocturnal |
|------------|---------------------|---------------------|-------------------|---------------------|
| | natural | natural | cross | cross |
| Fruit set | 49.1 ^a | 48.2 ^a | 42.0 ^a | 26.8 ^b |
| Seeds with | 48.2 ± 20.6^{a} | 42.7 ± 17.8^{a} | 41.7 ± | 34.3 ± 20.0^{a} |
| embryo | | | 20.3 ^a | |

Pollination treatments producing significant differences (P < 0.05) are indicated by different superscript letters.

- $\mathbf{5}$
- 6
- $\overline{7}$

1 Figures



3 Fig. 1. Flowering of *Mitrastemon yamamotoi*. (A) Male stage. (B) Transitional stage

- 4 (the stamen tube begun to fall off). (C) Female stage.
- $\mathbf{5}$
- 6



2 Fig. 2. Floral visitors of *Mitrastemon yamamotoi*. (A) Vespa simillima xanthoptera. (B)

- 3 Vespa analis. (C) Vespula flaviceps. (D) Vespa mandarinia. (E) Duolandrevus ivani. (F)
- 4 Diestrammena yakumontana. (G) Ornebius kanetataki. (H) Aphonoides rufescens. (I)
- 5 Aegus laevicollis. (J) Onthophagus yakuinsulanus. (K) Opisthoplatia orientalis. (L)
- 6 Onychostylus pallidiolus. (M) Tachinidae sp. (N) Sarcophagidae sp. (O) Limonia sp. (P)
- 7 Drosophila sp. (Q) Paratrechina flavipes. (R) Thereuopoda clunifera.
- 8