



Detoxification of Reactive Carbonyl Species by Glutathione Transferase Tau Isozymes

Mano, Jun'ichi
Kanameda, Sayaka
Kuramitsu, Rika
Matsuura, Nagisa
Yamauchi, Yasuo

(Citation)

Frontiers in Plant Science, 10:487-487

(Issue Date)

2019-04-24

(Resource Type)

journal article

(Version)

Version of Record

(Rights)

© 2019 Mano, Kanameda, Kuramitsu, Matsuura and Yamauchi.

This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited a...

(URL)

<https://hdl.handle.net/20.500.14094/90005935>





Detoxification of Reactive Carbonyl Species by Glutathione Transferase Tau Isozymes

Jun'ichi Mano^{1*}, Sayaka Kanameda², Rika Kuramitsu², Nagisa Matsuura³ and Yasuo Yamauchi^{3*}

¹ Science Research Center, Organization for Research Initiatives, Yamaguchi University, Yamaguchi, Japan, ² Faculty of Agriculture, Yamaguchi University, Yamaguchi, Japan, ³ Graduate School of Agricultural Science, Kobe University, Kobe, Japan

OPEN ACCESS

Edited by:

Peter Schröder,
Helmholtz Center Munich German
Research Center for Environmental
Health, Germany

Reviewed by:

Karl-Josef Dietz,
Bielefeld University, Germany
Alessandra Francini,
Sant'Anna School of Advanced
Studies, Italy

*Correspondence:

Jun'ichi Mano
mano@yamaguchi-u.ac.jp
Yasuo Yamauchi
yamauchi@kobe-u.ac.jp

Specialty section:

This article was submitted to
Plant Physiology,
a section of the journal
Frontiers in Plant Science

Received: 09 July 2018

Accepted: 29 March 2019

Published: 24 April 2019

Citation:

Mano J, Kanameda S,
Kuramitsu R, Matsuura N and
Yamauchi Y (2019) Detoxification
of Reactive Carbonyl Species by
Glutathione Transferase Tau
Isozymes. *Front. Plant Sci.* 10:487.
doi: 10.3389/fpls.2019.00487

Oxidative stimuli to living cells results in the formation of lipid peroxides, from which various aldehydes and ketones (oxylipin carbonyls) are inevitably produced. Among the oxylipin carbonyls, those with an α,β -unsaturated bond are designated as reactive carbonyl species (RCS) because they have high electrophilicity and biological activity. Plants have arrays of dehydrogenases and reductases to metabolize a variety of RCS that occur in the cells, but these enzymes are not efficient to scavenge the most toxic RCS (i.e., acrolein) because they have only low affinity. Two glutathione transferase (GST) isozymes belonging to the plant-specific Tau class were recently observed to scavenge acrolein with K_M values at a submillimolar level. This suggests that GST could also be involved in the defense system against RCS. We tested the activities of 23 Tau isozymes of *Arabidopsis thaliana* for five types of RCS, and the results revealed that 11 isozymes recognized either acrolein or 4-hydroxy-(E)-2-nonenal or both as a substrate(s). Such RCS-scavenging activities indicate the potential contribution of GST to RCS scavenging in plants, and they may account for the stress tolerance conferred by several Tau isozymes. RCS are therefore a strong candidate for endogenous substrates of plant GSTs.

Keywords: acrolein, lipid peroxide, oxidative stress, oxylipin, reactive electrophile species, redox signal

MAIN TEXT

Reactive Carbonyl Species (RCS) Are Signaling/Damaging Agents That Act Downstream of ROS

The production of reactive oxygen species (ROS) such as superoxide radical, hydrogen peroxide (H_2O_2), and singlet oxygen (1O_2) intrinsically accompanies aerobic life. One important aspect of ROS *in vivo*, although not always noticed, is that ROS are often produced in the close vicinity of membranes, in association with chloroplastic and mitochondrial electron transport chains and the

plasma membrane-bound respiratory burst NADPH oxidase homologs (RBOHs). Membrane lipids are therefore constitutively oxidized due to the basal generation of ROS (Mène-Safrané et al., 2007). The resulting lipid peroxides are relatively unstable and decompose or are metabolized to a variety of compounds called oxylipins, in which many types of aldehydes and ketones (oxylipin carbonyls) with different carbon chain lengths and extents of unsaturation are present. Carbonyl compounds are more reactive than corresponding alcohols and carboxylic acids, and the α,β -unsaturated carbonyls [reactive carbonyl species (RCS)] in particular have high electrophilicity and play critical biological roles in a range of functions from gene regulation to cytotoxicity (Esterbauer et al., 1991; Farmer and Mueller, 2013). Typical and well-studied RCS are acrolein, 4-hydroxy-(*E*)-2-nonenal (HNE), 4-oxo-(*E*)-2-nonenal, and malondialdehyde (MDA). The participation of RCS in oxidative injury and oxidative signaling in cells has been established for animals (Schopfer et al., 2011).

Reactive carbonyl species, e.g., acrolein and HNE, exhibit toxicity to plant cells and organelles when they are added exogenously (Millar and Leaver, 2000; Alméras et al., 2003; Mano et al., 2005, 2009). The occurrence of RCS in plant tissues was verified by extensive carbonyl analyses as follows. Yin et al. (2010) showed that tobacco roots contain dozens of carbonyls including several RCS, and they reported that the levels of some carbonyls were increased by the toxic level of aluminum ion. In leaves also, various carbonyls have been detected in tobacco (Mano et al., 2010), *Arabidopsis thaliana* (Yamauchi et al., 2012), and cyclamen (Kai et al., 2012), and their levels were increased by a high intensity of light (Mano et al., 2010), methyl viologen (Yamauchi et al., 2012), high salinity (Mano et al., 2014), injury (Matsui et al., 2012), and heat stress (Kai et al., 2012). **Table 1** summarizes the stress-related RCS and carbonyls identified in plants. The observed increases of RCS are ascribed to the increased levels of ROS by the stressors. These endogenously produced RCS were concluded to be responsible for the tissue damage because the extent of damage and the RCS levels correlated positively in transgenic plants that overexpress or lack an RCS-detoxifying enzyme (Mano et al., 2010; Yin et al., 2010; Yamauchi et al., 2012).

Reactive carbonyl species in plants also play signaling roles. Bate and Rothstein (1998) demonstrated that the exogenous addition of (*E*)-2-hexenal to *A. thaliana* plants induced a group of genes involved in defense against pathogens. The induced gene members vary by the RCS type (Alméras et al., 2003; Weber et al., 2004). Yamauchi et al. (2015) found that 2-alkenals of carbon chain length 4–8, when added as volatiles, induced heat-shock response genes in *A. thaliana*. Endogenous RCS produced upon an oxidative stimulus act as initiators of programmed cell death (PCD) in tobacco cultured cells (Biswas and Mano, 2015) by activating caspase-3-like protease (Biswas and Mano, 2016). In the stomata closure signaling of abscisic acid (ABA), the ROS production in guard cells is followed by increases in RCS, and the genetic suppression of RCS inhibited the stomata response to ABA (Islam et al., 2016). Together these observations, compiled over the past decade, indicate

that RCS are endogenous agents that mediate ROS stimuli to downstream responses.

Enzymatic Regulation of RCS

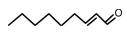
Dozens of oxylipin carbonyls in plants (**Table 1**) have a broad range of reactivity and thus different toxicity and signaling effects (Mano et al., 2009; Biswas and Mano, 2015). Plants have three types of oxidoreductases for metabolizing carbonyls: (i) aldehyde dehydrogenase (ALDH) to oxidize an aldehyde to a carboxylic acid with NAD⁺, (ii) aldehyde reductase to reduce an aldehyde or a ketone to a corresponding alcohol with NAD(P)H; two types of proteins, one belonging to aldo-keto reductase (AKR) family and the other to short-chain dehydrogenase/reductase family, can catalyze this reaction (Yamauchi et al., 2011), and (iii) 2-alkenal reductase (AER) and alkenal/one oxidoreductase (AOR) to reduce an RCS at the carbonyl-conjugated C-C double bond with NAD(P)H (Mano et al., 2002, 2005; Yamauchi et al., 2011). These enzyme classes, respectively, have multiple isozymes, and each isozyme shows distinct substrate specificity. **Table 1** summarizes the plant isozymes of these enzyme classes and reported substrates. Some of these isozymes have been shown to detoxify carbonyls *in planta*; their overexpression in transgenic plants reduced the carbonyl levels and conferred tolerance against several types of environmental stressors (reviewed by Mano, 2012).

Acrolein Is Scavenged by Glutathione Transferase

Among the RCS, acrolein (or 2-propenal), the C3 alkenal, is the most highly reactive and toxic compound (Esterbauer et al., 1991). It can inactivate photosynthetic machinery (Mano et al., 2009; Shimakawa et al., 2013) and induce PCD (Biswas and Mano, 2016). As seen in **Table 1**, many reductases recognize acrolein as a substrate, but they have higher affinity to longer-chain aldehydes and show only low affinity to acrolein, i.e., K_M values > 2 mM (Mano et al., 2017).

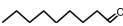

Acrolein reacts with the reduced form of glutathione (GSH) very rapidly (Esterbauer et al., 1975), and certain isozymes of glutathione transferase (GST) can catalyze the conjugation of acrolein with GSH. Human GST isozymes Alpha1, Mu1, and Pi1 recognize acrolein as a substrate (Berhane et al., 1994). Several plant GSTs have been known to react with RCS; for example, GST B1/B2 from sorghum recognizes HNE as a substrate (Gronwald and Plaisance, 1998). BI-GST and four Tau class isozymes from tomato reacted with (*E,E*)-2,4-nonadienal, and so did two isozymes from grapevine with (*E*)-2-hexenal (Kobayashi et al., 2011). We have investigated acrolein-scavenging GST activity and detected it in *A. thaliana*, spinach, rice, and Chinese cabbage. We then isolated a Tau isozyme from spinach for scavenging acrolein with the K_M value 93 μ M for acrolein. A homologous GST isozyme Tau19 from *A. thaliana* (AtGSTU19) also scavenged acrolein with the K_M value 30 μ M (Mano et al., 2017). The enzymatic scavenging of acrolein in plants had previously been attributed only to AER and AKR reactions (**Table 1**), but these GST Tau (GSTU) isozymes appear to be physiologically more

TABLE 1 | RCS and related carbonyls that are present in plants, and the plant enzymes that metabolize the carbonyls.

Compound name	Structure	Stimulus/stressor	Compatible enzyme			
			ALDH	AKR	AER	GST
Acrolein (propenal)		H ₂ O ₂ , MV, HL, heat, NaCl, Al, ABA		AtChIAKR, AKR4C9 AKR4C10, CHR Slr0942, Slr1192	AER CsAOR AtAOR	GST-Acr AtGSTUs*
Malondialdehyde		H ₂ O ₂ , NaCl, Al		AKR4C9, MsALR		
Crotonaldehyde (E)-2-butenal		MV, NaCl, Al		AtChIAKR, CsAOR AtAOR, Slr0942 Slr1192	AER	AtGSTUs*
Methylvinylketone (3-butene-2-one)		heat			AER AtAOR CsAOR	
Methacrolein						AtGSTU19
(E)-2-Pentenal		HL, ABA		AtChIADR AtCytADR AtChIAKR, CHR	AER	
3-Penten-2-one		MV			AER CsAOR AtAOR	
(E)-2-Hexenal		HL, NaCl, heat, injury	ALDH3H1 ALDH3I1	AtChIADR AtCytADR AtChIAKR, AKR4C9 CHR	AER	GST (grapevine) AtGSTU25*
HHE		H ₂ O ₂ , NaCl, Al, ABA			AER	
4-Oxo-(E)-2-Hexenal		injury				
(E)-2-Heptenal		Al		CHR		
(E)-2-Octenal		MV		CHR		
(E)-2-Nonenal		HL		AtChIADR AtCytADR AtChIAKR, CHR	AER	
HNE		H ₂ O ₂ , Al, ABA		AKR4C9	AER	GSTB1/B2 (sorghum) AtGSTUs*
(E,E)-2,4-Nonadienal						BI-GST (tomato)
Formaldehyde	HCHO	Al, ABA	ALDH			
Acetaldehyde		H ₂ O ₂ , Al, ABA	ALDH			
Propionaldehyde		H ₂ O ₂ , ABA		AtChIADR AtCytADR AtChIAKR, CHR Slr0942, Slr1192		
Butyraldehyde			ALDH	AtChIADR AtCytADR AtChIAKR, CHR		
n-Pentanal			ALDH	CHR		
n-Hexanal		H ₂ O ₂ , Al	ALDH	CHR		
(Z)-3-Hexenal		Al, ABA, injury		AtChIADR, AtCytADR, AtChIAKR, CHR		
n-Heptanal		H ₂ O ₂ , Al	ALDH	CHR		
n-Octanal		Al		CHR		

(Continued)

TABLE 1 | Continued

Compound name	Structure	Stimulus/stressor	Compatible enzyme			
			ALDH	AKR	AER	GST
<i>n</i> -Nonanal				CHR		
Methylglyoxal		NaCl		AKR4C9, AKR4C10 AtChIADR AtCytADR AtChIAKR, Slr0315 Slr0942, Slr1192		

**This study. Compound names in boldface are an RCS. 'Stimulus/stressor' indicates the exogenous stressors and endogenous stimuli reported to increase the level of each carbonyl in plants (see the main text). Compatible enzymes represent isozymes for which a catalytic activity for the corresponding carbonyl has been reported. References for enzyme activity: AER (Mano et al., 2002, 2005), AKR4C9 (Simpson et al., 2009; Saito et al., 2013), AKR4C10 (Saito et al., 2013), ALDH (Asker and Davies, 1985), ALDH3H1 and ALDH3I1 (Stiti et al., 2011), AtAOR, AtChIAKR, AtChIADR, and AtCytADR (Yamauchi et al., 2011), AtGSTU19 (Mano et al., 2017), Bt-GST (Kobayashi et al., 2011), CHR (Tanaka et al., 2018), MsALR (Oberschall et al., 2000), CsAOR (Yamauchi et al., 2011), GST-Acr (Mano et al., 2017), GST B1/B2 (Gronwald and Plaisance, 1998), GST from grapevine (Kobayashi et al., 2011), and Slr0545, Slr0942, Slr1192 and Slr1503 (Shimakawa et al., 2013). Slashed boxes, No data is available.*

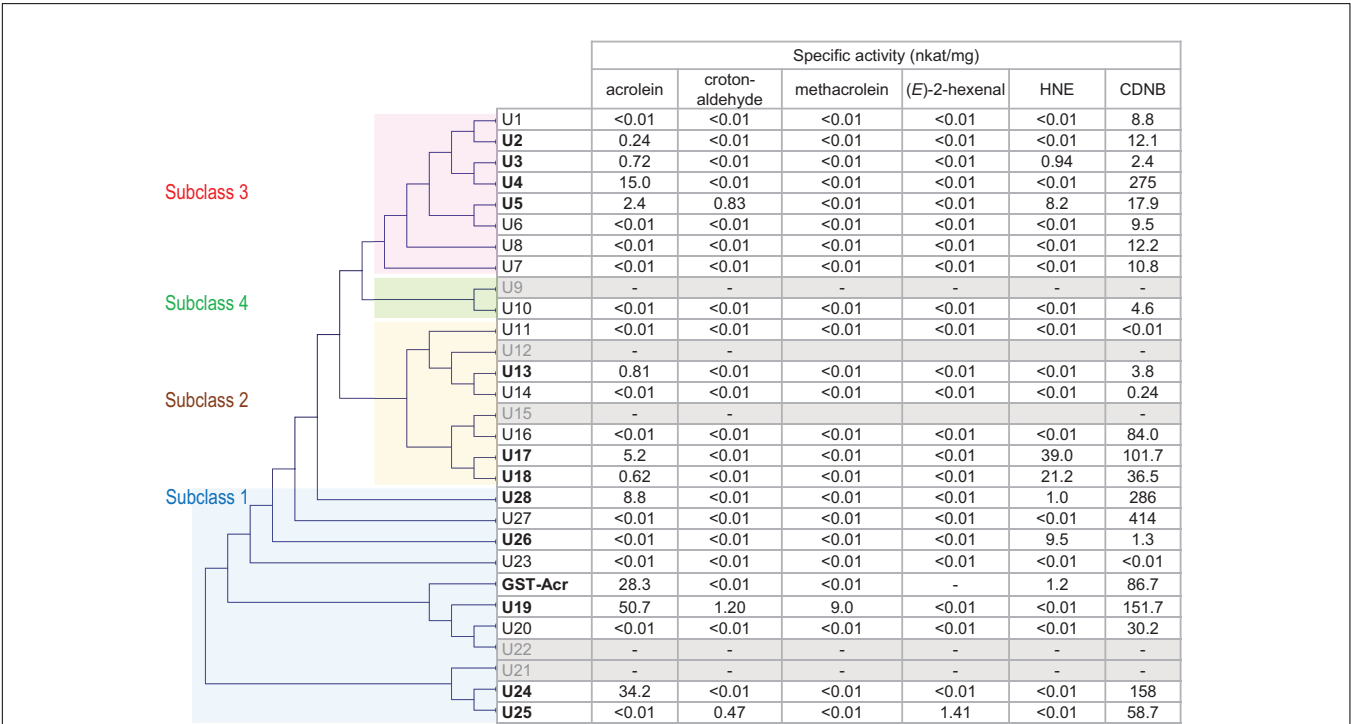


FIGURE 1 | Substrate specificity of AtGSTU isozymes and spinach GST-Acr. Gray rows represent the isozymes that were not recovered as soluble protein. The assay conditions are described in the “Materials and Methods” section. The GST-Acr and AtGSTU19 data are from our earlier study (Mano et al., 2017). The isozymes are arranged in the order of the phylogenetic tree, which was constructed on the amino acid sequence similarity by the neighbor-joining method using the multiple sequence alignment software Clustal W 2.0 (Larkin et al., 2007). The amino acid sequence of the spinach GST-Acr was deduced from the assembled RNA sequence (Mano et al., 2017).

relevant because their K_M values are close to the physiological range of acrolein.

Eleven AtGSTU Isozymes Recognize RCS as Substrates

GST Tau is a plant-specific class and is the largest group of GST isozymes in angiosperms, gymnosperms, and ferns (Flova, 2003; Monticcolo et al., 2017). GSTU isozymes have important roles in plants’ defense against environmental stress.

The overexpression of a GSTU gene from the extreme halophyte *Salicornia brachiata* (Jha et al., 2011) in tobacco conferred salt tolerance. *A. thaliana* plants overexpressing rice *OsGSTU4* gene (Sharma et al., 2014), *AtGSTU17* gene (Chen et al., 2012), and *AtGSTU19* gene (Xu et al., 2016) showed more tolerance to salinity and oxidative stress than the wild type. GSTU isozymes therefore constitute part of the anti-oxidative defense, but the underlying biochemical mechanism remains unclear because the physiologically relevant

substrates of GSTU have not been elucidated. The efficient acrolein-scavenging activity observed in the two GSTU isoforms described above suggested to us the possibility that GSTU isoforms can be counted as RCS-scavenging enzymes. To test this possibility, we determined the RCS-scavenging activity of AtGSTU isoforms (**Figure 1**).

Complementary DNA of 28 AtGSTU isoforms was cloned and expressed in *Escherichia coli*, and pure recombinant proteins were obtained (see Materials and Methods). Recombinant AtGSTU9, AtGSTU12, AtGSTU15, AtGSTU21, and AtGSTU22 proteins were not recovered as the soluble form. The isoforms obtained as soluble proteins (23 in total) were first tested for activity for a universal GST substrate 1-chloro-2,4-dinitrobenzene (CDNB). Recombinant AtGSTU11 and AtGSTU23 proteins were incompetent, and the other 21 showed the CDBN-conjugating activity.

We then examined the 23 isoforms for the activity to scavenge five types of RCS, i.e., acrolein, crotonaldehyde, methacrolein, (*E*)-2-hexenal, and HNE (**Figure 1**). Acrolein-scavenging activity was detected (higher than 0.01 nkat mg⁻¹) in the following ten isoforms: AtGSTU2, AtGSTU3, AtGSTU4, AtGSTU5, AtGSTU13, AtGSTU17, AtGSTU18, AtGSTU19, AtGSTU24, and AtGSTU28. The specific activity of these isoforms ranged from 0.24 nkat mg⁻¹ (AtGSTU2) to 50.7 nkat mg⁻¹ (AtGSTU19). These results show that acrolein is a common endogenous substrate of GSTU.

For crotonaldehyde, three isoforms (AtGSTU5, AtGSTU19, and AtGSTU25) showed significant activity. For methacrolein, only AtGSTU19 and for (*E*)-2-hexenal only AtGSTU25 showed the activity. For HNE, six isoforms (AtGSTU3, AtGSTU5, AtGSTU17, AtGSTU18, AtGSTU26, and AtGSTU28) exhibited significant activity. In total, at least 11 of the 28 isoforms showed RCS-scavenging activity.

GST Tau isoforms are grouped into four subclasses based on their amino acid sequence similarity (Monticcolo et al., 2017) as indicated in **Figure 1**. RCS-scavenging activity was identified in the isoforms in subclasses 1, 2, and 3, and these three subclasses also have RCS-incompatible isoforms (**Figure 1**). This suggests that the acquisition of the RCS-recognizing ability during the molecular evolution of AtGSTUs occurred multiple times independently. An alternative possibility is that the common ancestor of AtGSTU had the RCS-scavenging activity and it was lost during the molecular evolution.

The Physiological Relevance of the RCS-Scavenging Activity of GST

Among these 11 RCS-compatible isoforms, AtGSTU13 and AtGSTU19 are expressed in almost all tissues except the male organ (**Supplementary Table S2**, data extracted from Dixon et al., 2010). In shoot tissues, AtGSTU1, AtGSTU17, and AtGSTU18 are strongly expressed, and in root tissues, AtGSTU2, AtGSTU4, AtGSTU24, and AtGSTU28 are expressed. AtGSTU18 and AtGSTU19 are constitutively expressed. In particular, AtGSTU19 is the most abundant GSTU isoform in *A. thaliana* (Dixon and Edwards, 2009) and appears to be a key isoform to protect the whole plant body from the toxicity of RCS, especially acrolein.

Other isoforms, in contrast, are induced by various stressors such as salt, high osmolarity, and UV-B [**Supplementary Table S2**, from AtGenExpress database (Kilian et al., 2007)], which commonly increase the intracellular levels of ROS. Notably, two isoforms (AtGSTU17 and AtGSTU19) show relatively high RCS-scavenging activities, and they conferred stress tolerance to transgenic plants (Chen et al., 2012; Xu et al., 2016). The physiological function of GSTUs as RCS scavengers can be verified by analyses of the RCS levels in these samples.

We reported the acrolein-scavenging GST activity in leaf extracts of *A. thaliana*, *Brassica rapa* var. *pekinensis* (both are Brassicales), *Oryza sativa* (Poales), and *Spinacia oleracea* (Caryophyllales), ranging from 120 to 255 nmol/min/mg protein, as determined by the HPLC analysis of the acrolein decrease rate (Mano et al., 2017). We here detected the activity in four more species (values indicate the activity in nmol/min/mg protein): extracts from leaves of *Allium cepa* (Liliales), 185; *Apium graveolens* var. *dulce* (Apiales), 148; *Glebionis coronaria* (Asterales), 208, and green bell fruits of *Capsicum annuum* var. *grossum* (Solanales), 155. The occurrence of the activity in all tested species (seven orders of angiosperms) supports the importance of acrolein-scavenging GST activity in plants.

Conclusion

It was revealed that at least 11 of the 28 GSTU isoforms in *A. thaliana* can recognize RCS as substrates, indicating that RCS are important endogenous substrates of GSTU. Some members of the RCS-compatible GSTU isoforms are expressed in various tissues constitutively, and others are induced by a variety of environmental stressors. Acrolein-scavenging GST activity was observed in a broad range of angiosperms at similar levels of specific activity. These findings demonstrate that RCS-scavenging GST activity is a significant element constituting the anti-oxidative defense in plants.

MATERIALS AND METHODS

The cDNA Cloning of AtGSTUs, Expression, and Purification of Recombinant Proteins

Recombinant AtGSTU19 with an N-terminal poly(His) tag was obtained as described (Mano et al., 2017). For other isoforms, the cDNA of the corresponding open reading frame (ORF) was obtained by polymerase chain reaction (PCR)-based cloning. Briefly, total RNA was prepared from 3-week-old *A. thaliana* using an RNeasy Plant Mini Kit (Qiagen, Hilden, Germany), and then cDNA was synthesized by a ReverTra Ace Kit (Toyobo, Osaka, Japan). The ORF of the GSTU was amplified by PCR using proof-reading KOD DNA polymerase (Toyobo) and the primers listed in **Supplementary Table S1**. Amplified PCR products were subcloned into pMD19 (Takara Bio, Shiga, Japan), and the identity of subcloned cDNA was verified by DNA sequencing. The confirmed ORF of

the GSTU was ligated into the multicloning site in pASK15-plus expression vector (IBA, Göttingen, Germany), which produces the recombinant GSTU comprising an N-terminal streptavidin tag.

Escherichia coli BL21 cells transformed with the expression plasmid were grown at 37°C in LB broth containing 100 µg ml⁻¹ ampicillin. Expression of the transgene was induced with anhydrotetracycline, and the recombinant protein was purified via affinity chromatography using Strep-Tactin Sepharose (IBA) according to the manufacturer's instruction. The purity of the GSTU was verified by sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE). The purified GSTU fraction was dialyzed against 10 mM Tris-HCl, pH 7.5, and finally mixed with an equal volume of 80%(v/v) glycerol and stored at -20°C until use.

The Assay Conditions for Each Substrate

CDNB: 100 mM potassium phosphate buffer, pH 6.5, 1.0 mM GSH, and 1.0 mM CDNB. The activity was monitored as the rate of absorbance increase at 340 nm (extinction coefficient 9.6 mM⁻¹ cm⁻¹). Acrolein, methacrolein, and (*E*)-2-hexenal: 10 mM MES-NaOH, pH 6.0, 0.1 mM GSH and 0.1 mM aldehyde [for (*E*)-2-hexenal, 0.05 mM]. Absorbance decreases at 215 nm (15.0 mM⁻¹ cm⁻¹) for acrolein, 220 nm (10.96 mM⁻¹ cm⁻¹) for methacrolein, and 224 nm (19.5 mM⁻¹ cm⁻¹) for (*E*)-2-hexenal. Crotonaldehyde and HNE: 100 mM Na-phosphate

buffer, pH 7.5, 0.1 mM GSH, and 0.1 mM aldehyde. Absorbance decreases at 240 nm (10.7 mM⁻¹ s⁻¹) for crotonaldehyde and 221 nm (13.1 mM⁻¹ cm⁻¹) for HNE. The rate of non-enzymatic conjugation was subtracted as a background.

AUTHOR CONTRIBUTIONS

JM and YY conceived the project and wrote the manuscript with contributions from all of the authors. NM performed the overexpression and purification of proteins. SK and RK analyzed the GST activity.

FUNDING

This work was supported by the Japan Society for the Promotion of Science (JSPS) KAKENHI, grant no. 17H03700.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fpls.2019.00487/full#supplementary-material>

REFERENCES

- Alm ras, E., Stolz, S., Vollenweider, S., Reymond, P., M ne-Saffran , L., and Farmer, E. E. (2003). Reactive electrophile species activate defense gene expression in *Arabidopsis*. *Plant J.* 34, 205–216. doi: 10.1046/j.1365-313x.2003.01718.x
- Asker, H., and Davies, D. D. (1985). Mitochondrial aldehyde dehydrogenase from higher plants. *Phytochemistry* 24, 689–693. doi: 10.1016/s0031-9422(00)84877-3
- Bate, N. J., and Rothstein, S. J. (1998). C6-volatiles derived from the lipoxygenase pathway induce a subset of defence-related genes. *Plant J.* 16, 561–569. doi: 10.1046/j.1365-313x.1998.00324.x
- Berhane, K., Widersten, M., Engstr m,  , Kozarich, J. W., and Mannervik, B. (1994). Detoxication of base propenals and other alpha,beta-unsaturated aldehyde products of radical reactions and lipid peroxidation by human glutathione transferases. *Proc. Natl. Acad. Sci. U.S.A.* 91, 1480–1484. doi: 10.1073/pnas.91.4.1480
- Biswas, M. S., and Mano, J. (2015). Lipid peroxide-derived short-chain carbonyls mediate hydrogen peroxide-induced and salt-induced programmed cell death in plants. *Plant Physiol.* 168, 885–898. doi: 10.1104/pp.115.256834
- Biswas, M. S., and Mano, J. (2016). Reactive carbonyl species activate caspase-3-like protease to initiate programmed cell death in plants. *Plant Cell Physiol.* 57, 1432–1442.
- Chen, J.-H., Jiang, H.-W., Hsieh, E.-J., Chen, H.-Y., Chien, C.-T., Hsieh, H.-L., et al. (2012). Drought and salt stress tolerance of an *Arabidopsis* glutathione S-transferase U17 knockout mutant are attributed to the combine effect of glutathione and abscisic acid. *Plant Physiol.* 158, 340–351. doi: 10.1104/pp.111.181875
- Dixon, D. P., and Edwards, R. (2009). Selective binding of glutathione conjugates of fatty acid derivatives by plant glutathione transferases. *J. Biol. Chem.* 284, 21249–21256. doi: 10.1074/jbc.M109.020107
- Dixon, D. P., Skipsey, M., and Edwards, R. (2010). Roles for glutathione transferases in plant secondary metabolism. *Phytochemistry* 71, 338–350. doi: 10.1016/j.phytochem.2009.12.012
- Esterbauer, H., Schaur, R., and Zollner, J. H. (1991). Chemistry and biochemistry of 4-hydroxynonenal, malonaldehyde and related aldehydes. *Free Radic. Biol. Med.* 11, 81–128. doi: 10.1016/0891-5849(91)90192-6
- Esterbauer, H., Zollner, J., and Scholz, N. (1975). Reaction of glutathione with conjugated carbonyls. *Z. Naturforsch.* 30, 466–473. doi: 10.1515/znc-1975-7-808
- Farmer, E. E., and Mueller, M. J. (2013). ROS-mediated lipid peroxidation and RES-activated signaling. *Annu. Rev. Plant Biol.* 64, 429–450. doi: 10.1146/annurev-arplant-050312-120132
- Flova, C. (2003). The plant glutathione transferase gene family: genomic structure, functions, expression and evolution. *Physiol. Plant* 119, 469–479. doi: 10.1046/j.1399-3054.2003.00183.x
- Gronwald, J. W., and Plaisance, K. L. (1998). Isolation and characterization of glutathione S-transferase isozymes from sorghum. *Plant Physiol.* 117, 877–892. doi: 10.1104/pp.117.3.877
- Islam, M. M., Ye, W., Matsushima, D., Munemasa, S., Okuma, E., Nakamura, Y., et al. (2016). Reactive carbonyl species mediate abscisic acid signaling in guard cells. *Plant Cell Physiol.* 57, 2552–2563. doi: 10.1093/pcp/pcw166
- Jha, B., Sharma, A., and Misra, A. (2011). Expression of SbGSTU (tau class glutathione S-transferase) gene isolated from *Salicornia brachiata* in tobacco for salt tolerance. *Mol. Biol. Rep.* 38, 4823–4832. doi: 10.1007/s11033-010-0625-x
- Kai, H., Hirashima, K., Matsuda, O., Ikegami, H., Winkelmann, T., Nakahara, T., et al. (2012). Thermotolerant cyclamen with reduced acrolein and methyl vinyl ketone. *J. Exp. Bot.* 63, 4143–4150. doi: 10.1093/jxb/ers110
- Kilian, J., Whitehead, D., Horak, J., Wanke, D., Weinl, S., Batistic, O., et al. (2007). The AtGenExpress global stress expression data set: protocols, evaluation and model data analysis of UV-B light, drought and cold stress responses. *Plant J.* 50, 347–363. doi: 10.1111/j.1365-313x.2007.03052.x
- Kobayashi, H., Takase, H., Suzuki, Y., Tanzawa, F., Tanaka, R., Fujita, K., et al. (2011). Environment stress enhances biosynthesis of flavor precursors, S-3-(hexan-1-ol)-glutathione and S-3-(hexan-1-ol) L-cysteine, in grapevine through glutathione S-transferase activation. *J. Exp. Bot.* 62, 1325–1336. doi: 10.1093/jxb/erq376

- Larkin, M. A., Blackshields, G., Brown, N. P., Chenna, R., McGettigan, P. A., McWilliam, H., et al. (2007). Clustal W and Clustal X version 2.0. *Bioinformatics* 23, 2947–2948. doi: 10.1093/bioinformatics/btm404
- Mano, J. (2012). Reactive carbonyl species: Their production from lipid peroxides, action in environmental stress, and the detoxification mechanism. *Plant Physiol. Biochem.* 59, 90–97. doi: 10.1016/j.plaphy.2012.03.010
- Mano, J., Belles-Boix, E., Babiychuk, E., Inzé, D., Hiraoka, E., Takimoto, K., et al. (2005). Protection against photooxidative injury of tobacco leaves by 2-alkenal reductase. detoxication of lipid peroxide-derived reactive carbonyls. *Plant Physiol.* 139, 1773–1783. doi: 10.1104/pp.105.070391
- Mano, J., Ishibashi, A., Muneuchi, H., Morita, C., Sakai, H., Biswas, S., et al. (2017). Acrolein-detoxifying isozymes of glutathione transferase in plants. *Planta* 245, 255–264. doi: 10.1007/s00425-016-2604-5
- Mano, J., Khorobrykh, S., Matsui, K., Iijima, Y., Sakurai, N., Suzuki, H., et al. (2014). Acrolein is formed from trienoic fatty acids in chloroplasts: A targeted metabolomics approach. *Plant Biotechnol.* 31, 535–544.
- Mano, J., Miyatake, F., Hiraoka, E., and Tamoi, M. (2009). Evaluation of the toxicity of stress-related aldehydes to photosynthesis in chloroplasts. *Planta* 230, 639–648. doi: 10.1007/s00425-009-0964-9
- Mano, J., Tokushige, K., Mizoguchi, H., and Khorobrykh, S. A. (2010). Accumulation of lipid peroxide-derived, toxic α,β -unsaturated aldehydes (E)-2-pentenal, acrolein and (E)-2-hexenal in leaves under photoinhibitory illumination. *Plant Biotechnol.* 27, 193–197. doi: 10.5511/plantbiotechnology.27.193
- Mano, J., Torii, Y., Hayashi, S., Takimoto, K., Matsui, K., Nakamura, K., et al. (2002). The NADPH:quinone oxidoreductase P1-(-crystallin in *Arabidopsis* catalyzes the α,β -hydrogenation of 2-alkenals: detoxication of the lipid peroxide-derived reactive aldehydes. *Plant Cell Physiol.* 43, 1445–1455. doi: 10.1093/pcp/pcf187
- Matsui, K., Sugimoto, K., Mano, J., Ozawa, R., and Takabayashi, J. (2012). Differential metabolism of green leaf volatiles in injured and intact parts of a wounded leaf meet distinct ecophysiological requirements. *PLoS One* 7:e36433. doi: 10.1371/journal.pone.0036433
- Mène-Saffrané, L., Davoine, C., Stolz, S., Majcherczyk, P., and Farmer, E. E. (2007). Genetic removal of tri-unsaturated fatty acids suppresses developmental and molecular phenotypes of an *Arabidopsis* tocopherol-deficient mutant. *J. Biol. Chem.* 282, 35749–35756. doi: 10.1074/jbc.m706838200
- Millar, A. H., and Leaver, C. J. (2000). The cytotoxic lipid peroxidation product, 4-hydroxy-2-nonenal, specifically inhibits decarboxylating dehydrogenases in the matrix of plant mitochondria. *FEBS Lett.* 481, 117–121. doi: 10.1016/s0014-5793(00)01976-1
- Monticello, F., Colantuono, C., and Chusano, M. L. (2017). Shaping the evolutionary tree of green plants: Evidence from the GST family. *Sci. Rep.* 7:14363. doi: 10.1038/s41598-017-14316-w
- Oberschall, A., Deák, M., Török, K., Sass, L., Vass, I., Kovács, I., et al. (2000). A novel aldose/aldehyde reductase protects transgenic plants against lipid peroxidation under chemical and drought stresses. *Plant J.* 24, 437–446. doi: 10.1111/j.1365-313x.2000.00885.x
- Saito, R., Shimakawa, G., Nishi, A., Iwamoto, T., Sakamoto, K., Yamamoto, H., et al. (2013). Functional analysis of the AKR4C subfamily of *Arabidopsis thaliana*: model structures, substrate specificity, acrolein toxicity, and responses to light and [CO₂]. *Biosci. Biotechnol. Biochem.* 77, 2038–2045. doi: 10.1271/bbb.130353
- Schopfer, F. J., Cipollina, C., and Freeman, B. A. (2011). Formation and signaling actions of electrophilic lipids. *Chem. Rev.* 111, 5997–6021. doi: 10.1021/cr200131e
- Sharma, R., Sahoo, A., Devendran, R., and Jain, M. (2014). Over-expression of a rice Tau class glutathione S-transferase gene improves tolerance to salinity and oxidative stresses in *Arabidopsis*. *PLoS One* 9:e92900. doi: 10.1371/journal.pone.0092900
- Shimakawa, G., Iwamoto, T., Mabuchi, T., Saito, R., Yamamoto, H., Amako, K., et al. (2013). Acrolein, an α,β -unsaturated carbonyl, inhibits both growth and PSII activity in the cyanobacterium *Synechocystis* sp. PCC 6803. *Biosci. Biotechnol. Biochem.* 77, 1655–1660. doi: 10.1271/bbb.130186
- Simpson, P. J., Tantiadapitak, C., Reed, A. M., Mather, O. C., Bunce, C. M., White, S. A., et al. (2009). Characterization of two novel aldo-keto reductases from *Arabidopsis*: Expression patterns, broad substrate specificity, and an open active-site structure suggest a role in toxicant metabolism following stress. *J. Mol. Biol.* 392, 465–480. doi: 10.1016/j.jmb.2009.07.023
- Stiti, N., Adewale, I. O., Petersen, J., Bartels, D., and Kirch, H.-H. (2011). Engineering the nucleotide coenzyme specificity and sulfhydryl redox sensitivity of two stress-responsive aldehyde dehydrogenase isoenzymes of *Arabidopsis thaliana*. *Biochem. J.* 434, 459–471. doi: 10.1042/BJ20101337
- Tanaka, T., Ikeda, A., Shiojiri, K., Ozawa, R., Shiki, K., Nagai-Kunihiro, N., et al. (2018). Identification of a hexenal reductase that modulates the composition of green leaf volatiles. *Plant Physiol.* 178, 552–564. doi: 10.1104/pp.18.00632
- Weber, H., Chételat, A., Reymond, P., and Farmer, E. E. (2004). Selective and powerful stress gene expression in *Arabidopsis* in response to malondialdehyde. *Plant J.* 37, 877–888. doi: 10.1111/j.1365-313x.2003.02013.x
- Xu, J., Tian, Y.-S., Xing, X.-J., Peng, R.-H., Zhu, B., Gao, J.-J., et al. (2016). Over-expression of AtGSTU19 provides tolerance to salt, drought and methyl viologen stresses in *Arabidopsis*. *Physiol. Plant.* 156, 164–175. doi: 10.1111/ppl.12347
- Yamauchi, Y., Hasegawa, A., Mizutani, M., and Sugimoto, Y. (2012). Chloroplastic NADPH dependent alkenal/one oxidoreductase contributes to the detoxification of reactive carbonyls produced under oxidative stress. *FEBS Lett.* 586, 1208–1213. doi: 10.1016/j.febslet.2012.03.013
- Yamauchi, Y., Hasegawa, A., Taninaka, A., Mizutani, M., and Sugimoto, Y. (2011). NADPH-dependent reductases involved in the detoxification of reactive carbonyls in plants. *J. Biol. Chem.* 286, 6999–7009. doi: 10.1074/jbc.M110.202226
- Yamauchi, Y., Kunishima, M., Mizutani, M., and Sugimoto, Y. (2015). Reactive short-chain leaf volatiles act as powerful inducers of abiotic stress-related gene expression. *Sci. Rep.* 5:8030. doi: 10.1038/srep08030
- Yin, L., Mano, J., Wang, S., Tsuji, W., and Tanaka, K. (2010). The involvement of lipid peroxide-derived aldehydes in aluminum toxicity of tobacco roots. *Plant Physiol.* 152, 1406–1417. doi: 10.1104/pp.109.151449

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2019 Mano, Kanameda, Kuramitsu, Matsuura and Yamauchi. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.