



# Improved Fc epsilon RI-Mediated CD203c Basophil Responsiveness Reflects Rapid Responses to Omalizumab in Chronic Spontaneous Urticaria

Oda, Yoshiko ; Fukunaga, Atsushi ; Washio, Ken ; Imamura, Shinya ; Mizuno, Mayuko ; Hatakeyama, Mayumi ; Ogura, Kanako ; Nishigori,...

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**Improved FcεRI-mediated CD203c basophil responsiveness reflects rapid-responses to omalizumab in chronic spontaneous urticaria**

Yoshiko Oda, M.D, Atsushi Fukunaga, M.D., Ph.D. \*, Ken Washio, M.D., Ph.D., Shinya

Imamura, M.D., Mayuko Mizuno, M.D., Mayumi Hatakeyama, M.D., PhD., Kanako Ogura, M.D., Ph.D., and Chikako Nishigori M.D., Ph.D.

Division of Dermatology, Department of Internal Related, Kobe University Graduate School of Medicine, 7-5-1 Kusunoki-cho, Chuo-ku, Kobe 650-0017, Japan.

**\*Correspondence:** Atsushi Fukunaga, M.D., Division of Dermatology, Department of Internal Related, Kobe University Graduate School of Medicine, 7-5-1 Kusunoki-cho, Chuo-ku, Kobe 650-0017 Japan. Tel: +81-78-382-6134; Fax: +81-78-382-6149. E-mail: atsushi@med.kobe-u.ac.jp

**E-mail addresses**

Yoshiko Oda, yoda0320@med.kobe-u.ac.jp

Atsushi Fukunaga, atsushi@med.kobe-u.ac.jp

Ken Washio, washio@med.kobe-u.ac.jp

19 Mayuko Mizuno, mayukosumi@yahoo.co.jp

20 Shinya Imamura, shinyaimamura1987@yahoo.ne.jp

21 Mayumi Hatakeyama, mayumi1201o@yahoo.co.jp

22 Kanako Ogura, kanakoogura0115@gmail.com

23 Chikako Nishigori, chikako@med.kobe-u.ac.jp

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35

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## **Abstract**

**Background:** Omalizumab is effective in chronic spontaneous urticaria (CSU) patients

although its mechanism of action is poorly understood. Several studies reported that decreased

FcεRI-mediated histamine release and/or responsiveness was characteristic of basophils in CSU

patients. However, few studies have focused on the relationship between changes in basophil

responsiveness via FcεRI after omalizumab treatment and the therapeutic effect in CSU

patients.

**Objective:** To assess basophil responsiveness via FcεRI stimulation, as well as FcεRI

expression and IgE binding on blood basophils from CSU patients before and after omalizumab

treatment and its possible association with the clinical response.

**Methods:** We analyzed 34 CSU patients treated with omalizumab who were categorized as fast

responders (FRs) (n=20) and non or slow responders (N/SRs) (n=14). CD203c expression

induced by FcεRI stimulation, and IgE and FcεRI expressions on blood basophils from CSU

patients before and after omalizumab treatment were analyzed. Basophil responsiveness via

FcεRI stimulation was observed *in vitro* using basophils pre-treated with omalizumab.

54     **Results:** FRs had increased CD203c responsiveness after treatment with omalizumab compared  
55     with N/SRs. This improvement of basophil responsiveness via FcεRI stimulation in FRs was not  
56     observed in peripheral blood basophils pre-incubated with omalizumab *in vitro*, suggesting  
57     omalizumab does not directly affect circulating pre-existing abnormal basophils.

58     **Conclusion:** Increased basophil responsiveness via FcεRI after omalizumab treatment is  
59     associated with the therapeutic effect and mechanism of action of omalizumab.

## **Highlights box**

### **What is already known about this topic?**

Basophil functional abnormalities such as the low-responsiveness of basophils via FcεRI were reported in chronic spontaneous urticaria. Moreover, several studies reported increased FcεRI-mediated histamine release after omalizumab treatment.

### **What does this article add to our knowledge?**

Improvement of attenuated basophil responsiveness via FcεRI stimulation in chronic spontaneous urticaria is associated with rapid clinical effectiveness and the mechanism of action of omalizumab might be due to improved responsiveness of newly circulating basophils.

### **How does this study impact current management guidelines?**

Changes in basophil responsiveness via FcεRI stimulation in some patients with chronic spontaneous urticaria before and after omalizumab treatment suggest the importance of basophil status as an action point of omalizumab treatment.

**Key words:** chronic spontaneous urticaria, omalizumab, CD203c, basophil activation test

### ***Abbreviations used***

ASST: autologous serum skin test

BAT: basophil activation test

- 79 BHRA: basophil histamine release assay
- 80 CSU: chronic spontaneous urticaria
- 81 DLQI: dermatology life quality index
- 82 FcεRI: high-affinity IgE receptor
- 83 FRs: fast responders
- 84 GRs: good responders
- 85 HCs: healthy controls
- 86 IgE: immunoglobulin E
- 87 MFI: mean fluorescence intensity
- 88 N/PRs: non or partial responders
- 89 N/SRs: non or slow responders
- 90 Syk: spleen tyrosine kinase
- 91 UAS: urticaria activity score
- 92 UCT: urticaria control test

## 93    **Introduction**

94    Chronic spontaneous urticaria (CSU) is characterized by the spontaneous appearance of wheals,  
95    angioedema, or both for > 6 weeks and is associated with known (autoreactivity) or unknown  
96    causes<sup>1</sup>. Although the pathophysiology of CSU is unclear, basophils have been increasingly  
97    recognized as having critical roles and some investigators have described an association  
98    between disease pathophysiology and basophil function and number in CSU.  
99    Several reports documented that circulating basophils from patients with CSU released less  
100    histamine than healthy controls (HCs) when stimulated with anti-IgE antibody or anti-high-  
101    affinity IgE receptor (FcεRI) antibody<sup>2,3,4,5</sup>. Furthermore, reduced CD63 expression on basophils  
102    after anti-FcεRI stimulation was reported in a subgroup of CSU patients compared with HC  
103    basophils<sup>6</sup>. In our previous study<sup>7</sup>, basophils from CSU patients had attenuated upregulation of  
104    the activation marker CD203c against IgE and FcεRI antibodies compared with basophils from  
105    HCs, and the attenuated basophil responsiveness in patients with CSU was associated with the  
106    severity of disease and a relatively shorter disease duration. In addition, Huang et al. reported  
107    that basophil non-responders (< 10% of total histamine release to IgE) and basopenics  
108    (histamine concentrations < 5ng/mL blood leukocytes) have more severe, but shorter disease  
109    compared to responders<sup>8</sup>.



CSU patients have unique features related to basophil number as well as basophil function and responsiveness in the circulating blood. Rorsman first described a reduction of peripheral blood basophils in CSU patients<sup>9</sup>. Subsequently, basopenia was reported to be correlated with urticaria activity<sup>10,11</sup>. Furthermore, basophils were reported to be present in skin lesions of urticaria at levels higher than in non-lesion skin<sup>12</sup>. These findings suggest basopenia may reflect the recruitment of basophils to skin tissues. Although basophil migration was investigated in previous studies<sup>13,14</sup>, the recruitment pathway remains unknown.

Omalizumab is a recombinant humanized anti-IgE monoclonal antibody that selectively binds to the C3 domain of IgE, thereby blocking the binding of IgE to high-affinity receptors on effector cells, and inhibiting IgE-mediated cellular responses<sup>15</sup>. Omalizumab has been approved for use in patients with CSU who remain symptomatic despite H1 antihistamine treatment. Although omalizumab binds to free IgE, which lowers free IgE levels and causes FcεRI receptors on basophils and mast cells to be internalized and degraded, the mechanism of action in CSU is not understood completely<sup>16,17</sup>. Several reports demonstrated changes in basophil number and function after treatment with omalizumab. Furthermore, the numbers of circulating blood basophils increased after omalizumab treatment<sup>18</sup> in parallel with clinical improvement<sup>19, 20</sup>. With respect to basophil function, Gericke et al. reported increased anti-IgE-induced histamine release from blood basophils of CSU patients treated with omalizumab

compared with baseline value<sup>18</sup>. In contrast, it was reported that CD63-based basophil “releasability” by IL-3 coincubation after stimulation with anti-IgE antibody was decreased after omalizumab treatment compared with baseline values<sup>21</sup>.

Many reports have described pre-treatment biomarkers to predict the efficacy of omalizumab in CSU. Palacios et al. reported a lack of basophil CD203c upregulation in serum, which might reflect a lack of autoantibodies to IgE and/or FcεRI but which correlated with a higher clinical response<sup>22</sup>. Similarly, Deza et al. showed that a higher rate of positive autologous serum skin test (ASST) was associated with insufficient therapeutic effect<sup>23</sup>. Gericke et al. showed that a positive basophil histamine release assay (BHRA) or ASST as an indicator of serum autoreactivity was predictive of a slow response to treatment with omalizumab<sup>24</sup>. Additionally, baseline levels of basophil FcεRI expression were significantly lower in non-responders to omalizumab<sup>23</sup> and higher in fast responders than in slow responders<sup>25</sup>. Omalizumab responsiveness was also predicted by total serum IgE levels<sup>26</sup> and changes in total serum IgE levels<sup>27</sup>. Furthermore, Altrichter et al. observed that reduced serum IL-31 was associated with omalizumab responses<sup>28</sup>. However, few studies have focused on the relationship between baseline and/or altered basophil responsiveness via FcεRI, as well as IgE and FcεRI expression on basophils and the omalizumab therapeutic effect in CSU patients.

145           Therefore, the current study analyzed the expression of CD203c via FcεRI stimulation,  
146   as well as IgE and FcεRI expression on basophils from CSU patients before and after  
147   omalizumab treatment and its possible association with the clinical response. Moreover, we  
148   examined these parameters as a potential predictor of responses to omalizumab therapy. In  
149   addition, we performed an *in vitro* study using FR basophils pre-treated with omalizumab  
150   whether omalizumab could affect basophil reactivity in short term incubation.

## Material and Methods

### Study population

Thirty-four patients with CSU who remained symptomatic despite H1-antihistamines (even up to two times the recommended dose in Japan) and who were treated with omalizumab were enrolled at the Dermatological Institute of Kobe University Hospital. CSU was defined as recurrent wheals occurring for more than 6 weeks without an identifiable cause. Omalizumab 300 mg was injected subcutaneously at least three times at 4 weeks intervals. Clinical variables were evaluated using the Urticaria Control Test (UCT), which is an outcome instrument to retrospectively assess urticaria control with a recommended cutoff value of 12 for controlled disease<sup>29</sup>. It was reported that UCT has a strong correlation with Dermatology Life Quality Index (DLQI)<sup>30</sup> and 7-day Urticaria Activity Score (UAS7)<sup>31</sup>. UCT scores were measured at day 0, and on weeks 4, 8, and 12 of treatment (i.e., before the 1st, 2nd, 3rd and 4th injections). Patients were categorized into FRs (n = 20) and N/SRs (n = 14) (Fig. E1) (Table I) based on the following criteria: FRs with UCT scores  $\geq 12$  up on week 4 (Fig. E2, A, see in this article's Online Repository); and N/SRs with UCT scores  $< 12$  up on week 4 (Fig. E2, B, see in this article's Online Repository). UAS7 could be measured in some patients and the data was presented in Fig. E2, C, D, see in this article's Online Repository. In addition, we classified patients into good responders (GRs) (n = 26) and N/PRs (n = 7) based on UCT scores on week

12 after treatment of omalizumab (Fig. E1, Table E1, see in this article's Online Repository):  
GRs with UCT scores  $\geq 12$  up on week 12 and N/PRs with UCT scores  $< 12$  on week 12. One  
patient was followed-up a clinical course up to 4 weeks but could not follow up until 12 weeks.  
Therefore, this patient was excluded from the GRs vs. N/PRs dataset. All study participants  
provided oral consent for this study after verbal and written explanations.

#### **Basophil activation test**

Whole blood was taken just before the first and second omalizumab administration using blood  
collection tubes containing ethylenediaminetetraacetic acid (EDTA) and assays were performed  
within 24 hours of blood sampling. The Allergenicity Kit (Beckman Coulter, Fullerton, CA,  
USA) was modified and used for the quantification of basophil CD203c expression as  
previously described<sup>7</sup>. Basophils were stimulated with anti-IgE antibody (clone: E124-2-8D)  
(0.1  $\mu\text{g/ml}$ ) (Beckman Coulter, Fullerton, CA, USA), VioBlue conjugated anti-IgE antibody  
(clone: MB10-5C4) (1.1  $\mu\text{g/ml}$ ) (Miltenyi Biotec, Bergisch Gladbach, Germany) or biotinylated  
anti-Fc $\epsilon$ RI antibody (clone: CRA1) (16  $\mu\text{g/ml}$ ) (BioAcademia, Osaka, Japan). Basophils  
incubated with phosphate-buffered saline (PBS) were used as a negative control.

Briefly, blood was stained with reagents consisting of CRTH2-FITC, CD203c-PE, and  
CD3-PC7 to identify basophils, and were then mixed with the respective stimulant or PBS at  
37°C for 15 minutes. Biotinylated antibodies against CRA1 were then coupled with APC

streptavidin (BD, Franklin Lakes, NJ, USA) at 4°C for 30 minutes. Erythrocytes were lysed and after washing twice with PBS, cells were resuspended in 0.3 mL PBS 0.1% formaldehyde and evaluated 500 basophils by flow cytometry (FACS verse, BD Biosciences, San Jose, CA, USA).

Flow cytometry results were analyzed by FlowJo software (FlowJo, LLC, Ashland, OR, USA). Results of antibody stimulation were expressed as a proportion of CD203c<sup>high</sup> basophils. The proportion of CD203c<sup>high</sup> basophils was determined using a threshold defined as the expression level above which only 5% of basophils in the negative control sample fluoresced<sup>32</sup>. Changes in basophil responsiveness via FcεRI stimulation after omalizumab treatment compared with before treatment were calculated as followed. After/before treatment ratio of basophil CD203c responsiveness was defined as CD203c<sup>high</sup> basophil (%) stimulated with each antibody after treatment /CD203c<sup>high</sup> basophil (%) stimulated with each antibody before treatment.

#### **Measurement of IgE and FcεRI levels on basophils**

Basophils were incubated with VioBlue conjugated anti-IgE antibody (clone: MB10-5C4; (Miltenyi Biotec) and biotinylated anti-FcεRI antibody (clone: CRA1; BioAcademia) and analyzed by flow cytometry using the same method as for IgE and FcεRI levels on basophils and FlowJo analysis using the same method as for basophil activation after anti-IgE or CRA1 antibody stimulation. IgE and FcεRI levels were evaluated as the mean fluorescent intensity

(MFI). Changes in IgE and FcεRI levels on basophils after omalizumab treatment compared with before treatment were calculated as follows:  $\Delta$ IgE expression ( $\Delta$ FcεRI expression): IgE levels (FcεRI levels) on basophils before treatment — after treatment.

#### ***In vitro* study using basophils pre-treated with omalizumab**

Whole blood obtained from two CSU patients before omalizumab administration using blood collection tubes containing EDTA was incubated for 1, 12 or 24 hours at room temperature with 30 µg/mL omalizumab (Novartis Pharma, Tokyo, Japan)<sup>33</sup>. Samples with/without omalizumab preincubation were measured for CD203c expression after stimulation with anti-IgE antibody (clone: E124-2-8D, MB10-5C4). Results of antibody stimulation were expressed as the proportion of CD203c<sup>high</sup> basophils and IgE levels on basophils were expressed as the MFI, and sequential changes were investigated.

#### **Serum total IgE levels and basophil counts**

Serum total IgE levels were measured by immunoglobulin E-radioimmunosorbent test. Numbers of blood basophils (/µl) were assessed by an automated analyzer in the laboratory of the Kobe University Graduate School of Medicine.

Changes in total serum IgE levels and basophil counts on basophils after omalizumab treatment compared with before treatment were calculated as follows:  $\Delta$ total serum IgE ( $\Delta$ basophil counts): total serum levels (basophil counts) after treatment — before treatment.

223     **Statistical analysis**

224     The non-parametric Mann–Whitney *U*-test, unpaired *t*-test and Fischer’s exact test were used to  
225     assess differences between GRs and N/PRs. Kruskal-Wallis result with Dunn test was used for  
226     comparing three groups of nonparametric variables. To assess correlations between two factors,  
227     adjusted  $r_s$  (Spearman’s rank correlation coefficient) values were calculated. All statistical  
228     analyses were carried out using GraphPad Prism 7 (GraphPad Software, San Diego, CA, USA).  
229     Two-sided *p*-values  $< 0.05$  were considered statistically significant.



## **Results**

### **Study population**

CSU patients were categorized into two subgroups based on UCT scores 4 weeks after omalizumab treatment. Twenty out of 34 patients (58.8%) who achieved significant clinical improvement (UCT scores  $\geq 12$ ) were classified as FRs and the remaining fourteen (41.1%) who did not achieve significant clinical improvement (UCT scores  $< 12$ ) were classified as N/SRs (Fig. E1, see in this article's Online Repository). The baseline clinical characteristics of these subgroups are described in Table I. They exhibited no significant differences regarding sex, age, disease duration, serum IgE levels, and basophil counts. Additionally, there were no significant differences regarding the ASST positive rate in contrast to a previous study<sup>24</sup>. There were no differences among the two groups with respect to baseline UCT, UAS7, or DLQI score. In addition, although we divided CSU patients into GRs and N/PRs based on UCT scores on week 12 after treatment of omalizumab. There were no differences in serum IgE and ASST when comparing GRs and N/PRs (Table EI, see in this article's Online Repository). However, it was noted that one patient among N/PRs did not have data of total serum IgE before treatment and the patient was excluded from the GRs vs. N/PRs dataset (Table EI).

### **Baseline basophil responsiveness via Fc $\epsilon$ RI stimulation**

Although several studies described biomarkers for therapeutic responses to omalizumab<sup>22,23,26,27,28</sup>, few studies have focused on basophil responsiveness via FcεRI as a predictor of the therapeutic response to omalizumab. Therefore, we analyzed the baseline expression of the activation marker CD203c with or without anti-IgE and/or FcεRI stimulation in CSU patients. The expression of CD203c on peripheral blood basophils from CSU patients in the steady state without anti-IgE and/or FcεRI stimulation was similar between FRs and N/SRs (Fig. E3, see in this article's Online Repository). There were no differences in the percentage of CD203c<sup>high</sup> basophils stimulated with anti-IgE (E124-2-8D, MB-105C4) and FcεRI (CRA1) antibody between groups (Fig. 1A-C). Thus, basophil responsiveness via FcεRI stimuli did not predict the treatment response with omalizumab.

Furthermore, we classified patients with CSU into two groups based on the baseline proportion CD203c<sup>high</sup> basophils after anti-IgE stimulation as in the previous report<sup>7</sup> (14 non-responders [ $<10\%$  CD203c<sup>high</sup> basophil] and 20 responders [ $>10\%$  CD203c<sup>high</sup> basophil]) (Fig. E4, see in this article's Online Repository). However, there were no differences in fast or good effectiveness of omalizumab between non-responders and responders.

**Improvement in basophil responsiveness via FcεRI stimulation after omalizumab treatment**

It was previously demonstrated that blood basophil histamine release tended to increase with anti-IgE stimulation after omalizumab treatment compared with baseline<sup>18</sup>. In this study, we compared CD203c responsiveness of basophils via FcεRI stimulation with anti-IgE and FcεRI antibody before and after omalizumab treatment. The percentage of CD203c<sup>high</sup> basophils stimulated with any antibody increased in many FRs but not in most N/SRs (Fig. E5, Table EII, see in this article's Online Repository). Therefore, we compared the after/before treatment ratio of basophil CD203c responsiveness via FcεRI between FRs and N/SRs. When peripheral blood basophils were stimulated with three types of antibody, the after/before treatment ratio was significantly higher in FRs than in N/SRs (Fig. 2, Fig. E5, see in this article's Online Repository). Furthermore, the after/before ratio was around 1.5-2.0 for the anti-IgE antibody in FRs, but around 4 for the anti-FcεRI antibody (Fig. 2). These data suggest that omalizumab improved basophil responsiveness via FcεRI in FRs but not in N/SRs and that the improvement effect was associated with the rapid therapeutic response of omalizumab. However, it was needed to note that FRs included a group with improved basophil responsiveness and a group with no improvement, and N/SRs included some patients with improved basophil responsiveness (Fig. E5, see in this article's Online Repository).

#### **Baseline IgE and FcεRI levels of basophils**

Previous reports showed that the treatment effect of omalizumab may be predicted by serum IgE levels<sup>26,27</sup>. We previously reported that serum IgE levels were strongly correlated with IgE or FcεRI expression on basophils from CSU patients<sup>7</sup> and the correlations were also observed in overall participants of this study (Fig. E6, see in this article's Online Repository). Therefore, we investigated whether baseline basophil IgE expression was associated with the therapeutic response. No significant difference in IgE and FcεRI (CRA1) expression between FRs and N/SRs was observed (Fig. 3A, B). However, when CSU patients were categorized into GRs and N/PRs using the UCT score 12 weeks after omalizumab treatment, IgE expression on basophils from GRs was higher than those from N/PRs, whereas there was no difference in serum IgE between GRs and N/PRs (Fig. 3C, Table EI). However, it was noted that one patient among N/PRs did not have data of total serum IgE before treatment and the patient was excluded from the GRs vs. N/PRs dataset (Table EI). Additionally, there was no difference in FcεRI (CRA1) expression between GRs and N/PRs (Fig. 3D).

#### **Changes in IgE and FcεRI levels on basophils after omalizumab treatment**

It is widely accepted that omalizumab neutralizes free IgE and subsequently suppresses FcεRI and surface IgE expression on circulating basophils<sup>17</sup>. Deza et al. reported that patients exhibiting significant clinical improvement had a marked reduction in the levels of basophil FcεRI after 4 weeks<sup>23</sup>. Based on these findings, we evaluated the association between changes

in basophil FcεRI and IgE levels after omalizumab treatment and the therapeutic effect.  $\Delta$ IgE expression (IgE levels on basophils before treatment — after treatment) was not different between FRs and N/SRs (Fig. 4A). In contrast to a previous report<sup>23</sup>, there was no difference in  $\Delta$ FcεRI (CRA1) expression between the two groups (Fig. 4B). However,  $\Delta$ IgE expression was higher in GRs than N/PRs (Fig. 4C), whereas there was no difference in  $\Delta$ FcεRI (CRA1) expression between the two groups (Fig. 4D). These data indicate a more efficient inhibition of IgE binding on basophils in GRs than in N/PRs following omalizumab administration.

#### **Changes in serum total IgE levels and basophil counts**

Low serum IgE levels that increased after the start of omalizumab treatment were associated with insufficient clinical responses<sup>27</sup>. Additionally, Saini et al. showed that improved basopenia was associated with reduced clinical symptoms<sup>19</sup>. Thus, we evaluated the potential association between changes in serum IgE levels and circulating basophil counts in the blood after treatment with omalizumab and the therapeutic effect. When CSU patients were categorized into FRs and N/SRs, no differences in  $\Delta$ serum total IgE and  $\Delta$ basophil counts were observed between the groups (Fig. 5A, B). When CSU patients were categorized into GRs and N/PRs,  $\Delta$ serum total IgE and  $\Delta$ basophil counts just tended to be higher with no significant difference (Fig. 5C, D). However, three patients among N/PRs did not have data of total serum IgE and basophil count either before or after treatment and these patients were excluded from the GRs

vs. N/PRs dataset (Fig. 5C, D). Therefore, the number of samples in Fig. 5C and D was not be sufficient for statistical analyses, and if more samples were added, significant differences could be detected as previously reported<sup>19,20,27</sup>.

**Different effects of omalizumab on basophil responsiveness via FcεRI stimulation *in vivo* and *in vitro***

Finally, we investigated whether the responsiveness of basophils via FcεRI improved before omalizumab administration by incubating basophils from CSU patients with omalizumab *in vitro* to clarify the mechanism of action of omalizumab. Increased basophil responsiveness via FcεRI after *in vivo* omalizumab treatment was observed for cells isolated from the two FRs used in this study (Fig. 6A, B). However, improved basophil responsiveness via FcεRI stimulation was not observed at any time despite pre-treatment with omalizumab *in vitro* (Fig. 6C, D). These data suggest that pre-existing circulating basophils that exhibited low responsiveness via FcεRI before omalizumab administration did not have improved function after *in vitro* omalizumab treatment.

## 331 Discussion

332 Basophils have unique features and play a critical role in CSU. It was reported that basophils  
 333 from CSU patients released less histamine and/or exhibited low responsiveness when stimulated  
 334 via FcεRI compared with basophils from HCs<sup>2,3,4,5,6,7</sup>. We recently reported that the low  
 335 responsiveness of basophils via FcεRI reflected severe disease activity in CSU<sup>7</sup>. In addition to  
 336 the functional abnormalities of basophils, basopenia was correlated with disease activity in  
 337 CSU<sup>10,11</sup> and the cutaneous lesion recruitment of basophils in CSU. Although omalizumab was  
 338 effective in most patients with CSU who remained symptomatic despite H1 antihistamine  
 339 treatment, its mechanism of action in CSU is poorly understood. In the current study, we  
 340 focused on basophils as a predictive marker of the clinical effectiveness of omalizumab and as a  
 341 component of the mechanism of action of omalizumab. We classified and compared CSU  
 342 patients into two groups, FRs and N/SRs or GRs and N/PRs, based on the clinical efficacy of  
 343 omalizumab 4 or 12 weeks after treatment with omalizumab.

344 First, we focused on baseline parameters before omalizumab treatment as a pre-  
 345 treatment predictive marker of the clinical effectiveness of omalizumab. Baseline CD203c  
 346 responsiveness after stimulation with anti-IgE and FcεRI antibodies was not a useful pre-  
 347 treatment predictive marker, which is similar for the CD63-based report from Aghdam et al<sup>34</sup>  
 348 (Fig. 1). However, baseline IgE expression on basophils from GRs before omalizumab

treatment was higher than that in basophils from N/PRs (Fig. 3C). This difference was not observed between FRs and N/SRs (Fig. 3A). These observations suggested that higher IgE expression on basophils might predict a significant therapeutic effect 12 weeks after omalizumab treatment, even if the therapeutic effect was insufficient 4 weeks after treatment. In contrast to previous reports<sup>25</sup>, there were no differences in baseline FcεRI expression on basophils between FRs and N/SRs before omalizumab treatment (Fig. 3B). This difference may be related to differences in the definition of treatment response. In addition, a good predictor of N/PRs was found on surface IgE (Fig. 3C), but not serum IgE (Table EI). However, serum IgE strongly correlated with surface IgE on basophils (Fig. E6, see in this article's Online Repository), and the difference of results between the previous studies<sup>26,27</sup> that serum IgE was a good baseline predictor of omalizumab treatment efficacy and this study may be dependent on the small number of samples in this study.

Next, we investigated changes in basophil parameters and IgE after omalizumab treatment to determine their contribution to the mechanism of action of omalizumab in CSU. Regarding changes in basophil parameters, the after/before treatment ratios of basophil responsiveness (CD203c response) were significantly higher in FRs compared with N/SRs (Fig. 2), suggesting that the improvement of low responsiveness in circulating basophils via FcεRI was related to the rapid therapeutic effect of omalizumab. In addition, the after/before ratio was



around 1.5-2.0 for the anti-IgE antibody in FRs, but around 4 for the anti-FcεRI antibody. It has been reported that during treatment with omalizumab, spleen tyrosine kinase (Syk) expression increases in peripheral blood basophils, offsetting the functional effects mediated by the drug-induced reduction in cell surface density of FcεRI and its bound IgE<sup>35</sup>. Basophils which recovered functionally after omalizumab treatment might promote Syk signal more strongly when stimulated with anti- FcεRI than when anti-IgE.

CD203c upregulation after anti-IgE stimulation was demonstrated to be earlier than CD63 upregulation<sup>36</sup>. In addition, Ebo et al mentioned that up-regulation of CD203c does not per se indicate histamine release<sup>37</sup> and CD203c is not a degranulation marker like CD63. Thus, increased CD203c response to the IgE concentration after treatment of omalizumab observed in current study does not mean an increase in histamine release. Aghdam et al reported that when CD63 expression on basophils was used as a biomarker in the omalizumab responder, no significant increase after stimulation of the anti-IgE antibody was observed in non-responders after omalizumab treatment, contrary no increase in responders<sup>34</sup>. This difference between our study might be due to differences in the markers between CD203c and CD63 or to the definition of responder and non-responder. Indeed, Agdham et al. evaluates disease activity in six months.<sup>34</sup> Contrary, we evaluate the effectiveness in one month and three months.

384           The reduction of IgE expression on basophils 4 weeks after omalizumab treatment  
385    was larger in GRs compared with N/PRs (Fig. 4C). Regarding the changes in serum IgE,  
386    increased serum IgE levels 4 weeks after omalizumab treatment tended to be higher in GRs than  
387    in N/PRs (Fig. 5C). These differences were not observed between FRs and N/SRs (Fig. 4A,  
388    5A). Based on these results related to IgE, the larger reduction of IgE expression on basophils  
389    and the higher increase of serum IgE levels 4 weeks after omalizumab treatment suggested that  
390    good clinical responses were achieved 12 weeks after omalizumab treatment, even if they were  
391    not achieved 4 weeks after treatment. Moreover, these data imply that omalizumab efficiently  
392    inhibited the binding of IgE to basophils by binding to free serum IgE in GRs compared with  
393    N/PRs, which was the cause of the increase in apparent serum IgE levels in GRs.

394           Whether this improvement of basophil responsiveness by omalizumab treatment acts  
395    directly on low responsive circulating basophils before omalizumab treatment is an important  
396    issue related to the mechanism of action of omalizumab in CSU. Finally, we investigated  
397    whether omalizumab acted directly on circulating basophils in FRs before treatment *in vitro*. An  
398    improvement of basophil responsiveness via FcεRI stimulation was not observed at any time  
399    despite *in vitro* omalizumab treatment (Fig. 6C, D). These data indicate that in FRs, the  
400    functions of pre-existing basophils that exhibited low responsiveness via FcεRI stimulation  
401    before omalizumab treatment could not be improved. In contrast, *in vivo* treatment with

omalizumab in these cases improved basophil responsiveness via FcεRI stimulation and induced a rapid and good clinical response (Fig. 6A, B). These differences between *in vitro* and *in vivo* omalizumab treatment suggest that newly circulating basophils, possibly from skin tissue or bone marrow, maintained normal function in the blood under the state of low IgE binding to their surface by the efficient formation of an immune complex with omalizumab and serum IgE. Indeed, it was reported that the life span of mature basophils is approximately 60–70 hours<sup>38</sup>. A recent review proposed that basophils in CSU patients are mildly activated, persistently release a small amount of histamine, and are involved in the earliest stages of the cascade of CSU pathogenesis<sup>39</sup>.

This study includes limitations about grouping. The main focus of this study was to observe differences in basophil responsiveness between FRs and other groups (N/SRs) at 4 weeks after treatment. However, because we considered that the comparison of FRs and N/SRs alone was not enough to evaluate scientific fairly, we added the comparison data of GRs and N/PRs using another classification method. It would be ideal if the NRs could be independent for comparison but the number of NRs is overwhelmingly small as in the existing reports and a larger-scale research is needed for this comparison. Additionally, we used only one concentration antibody to stimulate the basophils and might have missed a curve shift of basophil reactivity.

420           In summary, improvement of attenuated basophil responsiveness via FcεRI stimulation  
421   in patients with CSU was associated with the rapid clinical effectiveness of omalizumab.  
422   However, further research is needed to elucidate the role of basophils in the mechanism of  
423   action of omalizumab in CSU. Although this study had some limitations, including a small  
424   number of cases, it highlights the importance of basophil status as an action point of  
425   omalizumab in CSU.

426

#### 427   **Acknowledgments**

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## References

1. Zuberbier T, Aberer W, Asero R, Abdul Latiff AH, Baker D, Ballmer-Weber B, et al. The EAACI/GA<sup>2</sup>LEN/EDF/WAO guideline for the definition, classification, diagnosis and management of urticaria. *Allergy* 2018;73:1393-414.
2. Greaves MW, Plummer VM, McLaughlan P, Stanworth DR. Serum and cell bound IgE in chronic urticaria. *Clin Allergy* 1974;4:265.
3. Kern F, Lichtenstein LM. Defective histamine release in chronic urticaria. *J Clin Invest* 1976;57:1369-77.
4. Eckman JA, Hamilton RG, Gober LM, Sterba PM, Saini SS. Basophil phenotypes in chronic idiopathic urticaria in relation to disease activity and autoantibodies. *J Invest Dermatol* 2008;128:1956-63.
5. Vonakis BM, Vasagar K, Gibbons SP Jr, Gober L, Sterba PM, Chang H, et al. Basophil FcεpsilonRI histamine release parallels expression of Src-homology 2-containing inositol phosphatases in chronic idiopathic urticaria. *J Allergy Clin Immunol*. 2007;119:441-8.
6. Rauber MM, Pickert J, Holiangu L, Möbs C, Pfützner W. Functional and phenotypic analysis of basophils allows determining distinct subtypes in patients with chronic urticaria. *Allergy* 2017;72:1904-11.
7. Oda Y, Fukunaga A, Washio K, Imamura S, Hatakeyama M, Ogura K, et al. Low responsiveness of basophils via FcεpsilonRI reflects disease activity in chronic spontaneous urticaria. *J Allergy Clin Immunol Pract* 2019 doi: 10.1016/j.jaip.2019.05.020.
8. Huang AH, Chichester KL, Saini SS. Association of basophil parameters with disease severity and duration in chronic spontaneous urticaria (CSU). *J Allergy Clin Immunol Pract* 2019 doi: 10.1016/j.jaip. 2019.08.004.
9. Rorsman H. Basophilic leucopenia in different forms of urticaria. *Acta Allergol* 1962;17:168-84.
10. Grattan CE, Dawn G, Gibbs S, Francis DM. Blood basophil numbers in chronic ordinary urticaria and healthy controls: diurnal variation, influence of loratadine and prednisolone and relationship to disease activity. *Clin Exp Allergy* 2003;33:337-41.
11. Oliver ET, Sterba PM, Saini SS. Interval shifts in basophil measures correlate with disease activity in chronic spontaneous urticaria. *Allergy* 2015;70:601-3.
12. Kay AB, Ying S, Ardelean E, Mlynek A, Kita H, Clark P, et al. Elevations in vascular markers and eosinophils in chronic spontaneous urticarial weals with low-level persistence in uninvolved skin. *Br J Dermatol* 2014;171:505-11.

- 467 13. Oliver ET, Sterba PM, Devine K, Vonakis BM, Saini SS. Altered expression of  
468 chemoattractant receptor-homologous molecule expressed on T(H)2 cells on blood  
469 basophils and eosinophils in patients with chronic spontaneous urticaria. *J Allergy Clin*  
470 *Immunol* 2016;137:304-6 e1.
- 471 14. Nakashima C, Otsuka A, Kabashima K. Recent advancement in the mechanism of  
472 basophil activation. *J Dermatol Sci* 2018;91:3-8.
- 473 15. Easthope S, Jarvis B. Omalizumab. *Drugs* 2001;61:253-60.
- 474 16. MacGlashan D Jr, Xia HZ, Schwartz LB, Gong J. IgE-regulated loss, not IgE-regulated  
475 synthesis, controls expression of FcεRI in human basophils. *J Leukoc Biol*  
476 2001;70:207-18.
- 477 17. Kaplan AP, Giménez-Arnau AM, Saini SS. Mechanisms of action that contribute to  
478 efficacy of omalizumab in chronic spontaneous urticaria. *Allergy* 2017;72:519-33.
- 479 18. Gericke J, Ohanyan T, Church MK, Maurer M, Metz M. Omalizumab may not inhibit  
480 mast cell and basophil activation in vitro. *J Eur Acad Dermatol Venereol* 2015;29:1832-  
481 6.
- 482 19. Saini SS, Omachi TA, Trzaskoma B, Hulter HN, Rosén K, Sterba PM, et al. Effect of  
483 omalizumab on blood basophil counts in patients with chronic idiopathic/spontaneous  
484 urticaria. *J Invest Dermatol* 2017;137:958-61.
- 485 20. Kishimoto I, Kambe N, Ly NTM, Nguyen CTH, Okamoto H. Basophil count is a  
486 sensitive marker for clinical progression in a chronic spontaneous urticaria patient  
487 treated with omalizumab. *Allergol Int* 2019;68:388-90.
- 488 21. Jörg L, Pecaric-Petkovic T, Reichenbach S, Coslovsky M, Stalder O, Pichler W, et al.  
489 Double-blind placebo-controlled trial of the effect of omalizumab on basophils in  
490 chronic urticaria patients. *Clin Exp Allergy* 2018;48:196-204.
- 491 22. Palacios T, Stillman L, Borish L, Lawrence M. Lack of basophil CD203c-upregulating  
492 activity as an immunological marker to predict response to treatment with omalizumab  
493 in patients with symptomatic chronic urticaria. *J Allergy Clin Immunol Pract*  
494 2016;4:529-30.
- 495 23. Deza G, Bertolín-Colilla M, Pujol RM, Curto-Barredo L, Soto D, García M, et al.  
496 Basophil FcεRI Expression in Chronic Spontaneous Urticaria: A Potential  
497 Immunological Predictor of Response to Omalizumab Therapy. *Acta Dermato*  
498 *Venereologica* 2017;97:698-704.
- 499 24. Gericke J, Metz M, Ohanyan T, Weller K, Altrichter S, Skov PS, et al. Serum  
500 autoreactivity predicts time to response to omalizumab therapy in chronic spontaneous  
501 urticaria. *J Allergy Clin Immunol* 2017;139:1059-61.e1.

25. Deza G, Bertolin-Colilla M, Sanchez S, Soto D, Pujol RM, Gimeno R, et al. Basophil FcεRI expression is linked to time to omalizumab response in chronic spontaneous urticaria. *J Allergy Clin Immunol* 2018;141:2313-6 e1.
26. Straesser MD, Oliver E, Palacios T, Kyin T, Patrie J, Borish L, et al. Serum IgE as an immunological marker to predict response to omalizumab treatment in symptomatic chronic urticaria. *J Allergy Clin Immunol Pract* 2018;6:1386-8.e1.
27. Ertas R, Ozyurt K, Atasoy M, Hawro T, Maurer M. The clinical response to omalizumab in chronic spontaneous urticaria patients is linked to and predicted by IgE levels and their change. *Allergy* 2018;73:705-12.
28. Altrichter S, Hawro T, Hänel K, Czaja K, Lüscher B, Maurer M, et al. Successful omalizumab treatment in chronic spontaneous urticaria is associated with lowering of serum IL-31 levels. *J Eur Acad Dermatol Venereol*. 2016;30:454-5.
29. Weller K, Groffik A, Church MK, Hawro T, Krause K, Metz M, et al. Development and validation of the Urticaria Control Test: A patient-reported outcome instrument for assessing urticaria control. *J Allergy Clin Immunol* 2014;133:1365-72.e6.
30. Itakura A, Tani Y, Kaneko N, Hide M. Impact of chronic urticaria on quality of life and work in Japan: Results of a real-world study. *J Dermatol* 2018;45:963-70.
31. Nakatani S, Oda Y, Washio K, Fukunaga A, Nishigori C. The Urticaria Control Test and Urticaria Activity Score correlate with quality of life in adult Japanese patients with chronic spontaneous urticaria. *Allergology International* 2018;68:279-81.
32. De Week AL, Sanz ML, Gamboa PM, Aberer W, Bienvenu J, Blanca M, et al. Diagnostic tests based on human basophils: more potentials and perspectives than pitfalls. II. Technical issues. *J Investig Allergol Clin Immunol* 2008;18:143-55.
33. FDA, Omalizumab Clinical Pharmacologic review, 2013. Available at: <http://www.fda.gov/downloads/Drugs/DevelopmentApprovalProcess/DevelopmentResources/UCM393855.pdf>
34. Alizadeh Aghdam M, Knol EF, van den Elzen M, den Hartog Jager C, van Os-Medendorp H, Knulst AC, et al. Response of FcεRI-bearing leucocytes to omalizumab in chronic spontaneous urticaria. *Clin Exp Allergy*. 2020;50:364-71.
35. MacGlashan DW Jr, Saini SS. Syk expression and IgE-mediated histamine release in basophils as biomarkers for predicting the clinical efficacy of omalizumab. *J Allergy Clin Immunol* 2017;139:1680-2.
36. Hennersdorf F, Florian S, Jakob A, Baumgärtner K, Sonneck K, Nordheim A, et al. Identification of CD13, CD107a, and CD164 as novel basophil activation markers and dissection of two response patterns in time kinetics of IgE-dependent upregulation. *Cell Res* 2005; 15: 325-35.

- 538 37. Ebo DG, Bridts CH, Mertens CH, Hagendorens MM, Stevens WJ, De Clerck LS.  
539 Analyzing histamine release by flow cytometry (HistaFlow):  
540 a novel instrument to study the degranulation patterns of basophils. J Immunol Methods  
541 2012 ; 375: 30-8.
- 542 38. Siracusa MC, Comeau MR, Artis D. New insights into basophil biology: initiators,  
543 regulators, and effectors of type 2 inflammation. Ann NY Acad Sci 2011;1217:166-77.
- 544 39. Yanase Y, Takahagi S, Hide M. Chronic spontaneous urticaria and the extrinsic  
545 coagulation system. Allergol Int 2018;67:191-4.  
546



Figure legends

Fig. 1 Comparison of the baseline proportion of CD203c<sup>high</sup> basophils between fast responders and non or slow responders.

A. Anti IgE antibody (E124-2-8D) stimulation.

B. Anti IgE antibody (MB10-5C4) stimulation.

C. Anti-FcεR1 antibody (CRA1) stimulation.

Statistical analyses were performed using the Mann-Whitney *U*-test.

Fig. 2 Comparison of the after/before treatment ratio of basophil CD203c responsiveness between fast responders and non or slow responders.

A. Anti IgE antibody (E124-2-8D) stimulation.

B. Anti IgE antibody (MB10-5C4) stimulation.

C. Anti-FcεR1 antibody (CRA1) stimulation.

The dotted line indicates no change (value = 1).

Statistical analyses were performed using the Mann-Whitney *U*-test.

Fig. 3 Comparison of baseline IgE and FcεRI on basophils (MFI) between fast responders and non or slow responders (A, B) or good responders and non or partial responders (C, D).

A, C. IgE expression.

B, D. FcεRI expression (CRA1).

Statistical analyses were performed using the unpaired *t*-test (A, C) or Mann-Whitney *U*-test (B, D).

MFI, mean fluorescence intensity.

Fig. 4 Comparison of changes in IgE or FcεRI levels on basophils (MFI) between fast responders and non or slow responders (A, B) or good responders and non or partial responders (C, D).

A, C. ΔIgE expression.

B, D. ΔFcεRI expression.

Statistical analyses were performed using the unpaired *t*-test (A, C) or Mann-Whitney *U*-test (B, D).

MFI, mean fluorescence intensity.

Fig. 5 Comparison of changes in total serum IgE levels and circulating basophil counts between fast responders and non or partial responders or good responders and non or partial responders (C, D).

583 A, C.  $\Delta$ total serum IgE; increase of total serum IgE.  
584 B, D.  $\Delta$ basophil counts; increase of circulating basophil counts.  
585 The dotted line indicates no change.  
586 Statistical analyses were performed using the Mann-Whitney *U*-test (A) or unpaired *t*-test (B).  
587 Three patients among N/PRs did not have data of total serum IgE and basophil count either  
588 before or after treatment and these patients were excluded from the GRs vs. N/PRs dataset (Fig.  
589 5C, D).  
590  
591 Fig. 6 Sequential changes in the parameters of *in vivo* omalizumab treatment (on 4 weeks of  
592 treatment) (A, B) and *in vitro* pre-treatment basophils with omalizumab in fast responders (C,  
593 D).  
594 A. Urticaria control test.  
595 B, C. CD203c<sup>high</sup> basophils (%) stimulated with anti-IgE antibody (E124-2-8D).  
596 D. CD203c<sup>high</sup> basophils (%) stimulated with anti-IgE antibody (MB10-5C4).  
597 MFI, mean fluorescence intensity.

598 Table I. Demographic characteristics based on the rapid therapeutic effect of omalizumab

	Fast responders (n = 20)	Non or slow responders (n = 14)	<i>P</i> value
Age, years	44.9 ± 4.4	52.7 ± 5.0	.29
Female, n (%)	12 (63.1%)	9 (64.2%)	>.99
Disease duration, years	2 (0.2–33)	3 (0.2–25)	.83
Total IgE (IU/ml)	126 (22.9–2535)	185 (3–4393)	.89
Basophil counts (/μl)	18.3 (0–118)	23 (0–98)	.41
ASST positive rate, n (%)	3/9 (33.3%)	4/6 (66.6%)	.31
UCT	6.1 ± 2.8	5.5 ± 2.7	.54
UAS7	22.9 ± 2.1	21.7 ± 3.4	.76
DLQI	7.0 (2–24)	7.0 (2–17)	.80

599 Data are given as the mean ± SD for age, UCT and UAS7; n (%) for sex, ASST positive rate;  
600 median (range) for disease duration, serum total IgE, basophil counts, and DLQI.  
601 Statistical differences between two groups were analyzed by unpaired *t*-test for age, UCT, and  
602 UAS7, Fisher's exact test for female and ASST positive rate, and Mann-Whitney *U*-test for  
603 disease duration, serum total IgE, basophil counts, and DLQI.  
604 ASST, autologous serum skin test; UCT, urticaria control test; UAS7, 7-day urticaria activity  
605 score; DLQI, dermatology life quality index.  
606

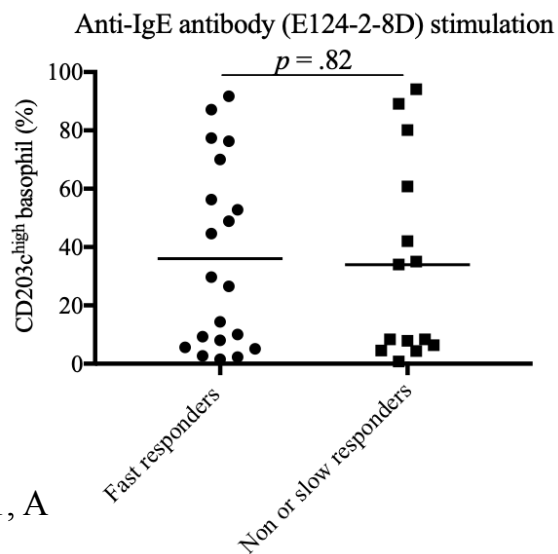


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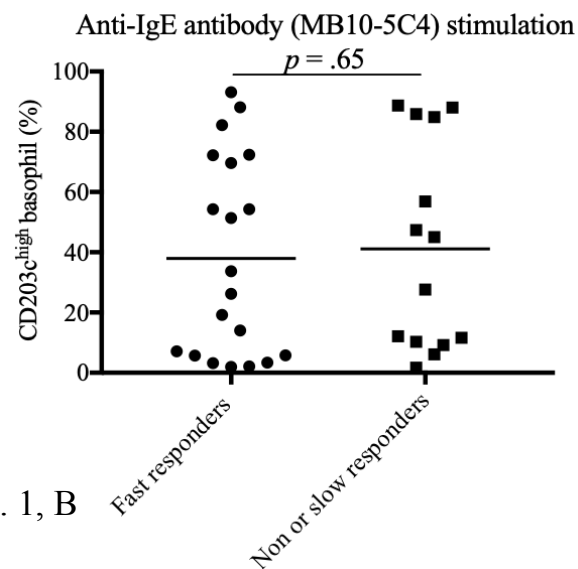


Fig. 1, B

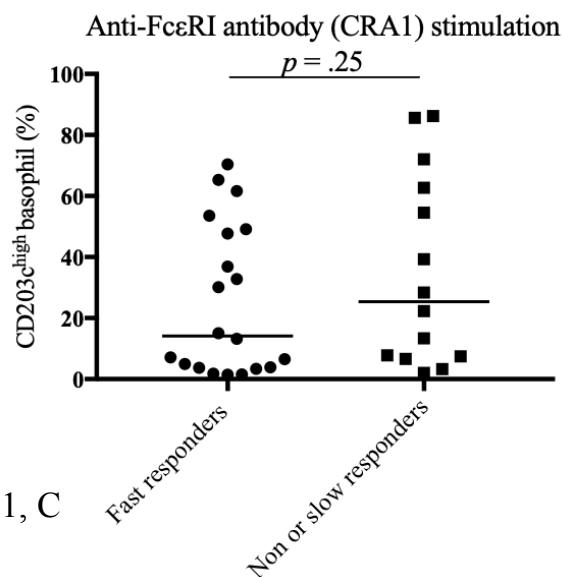


Fig. 1, C

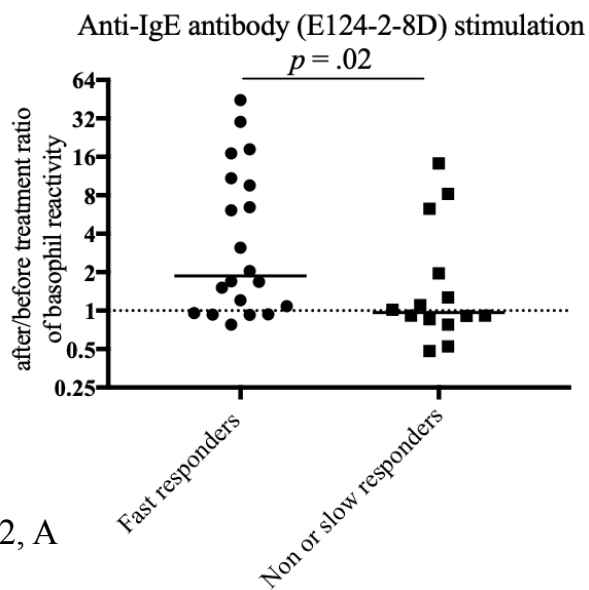


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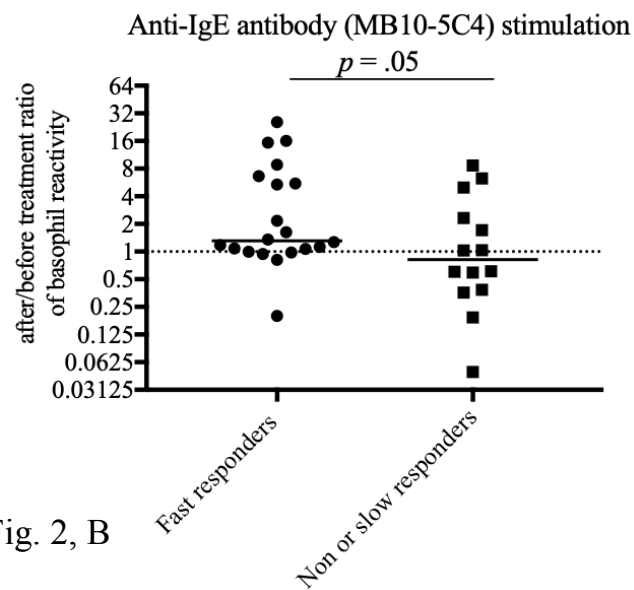


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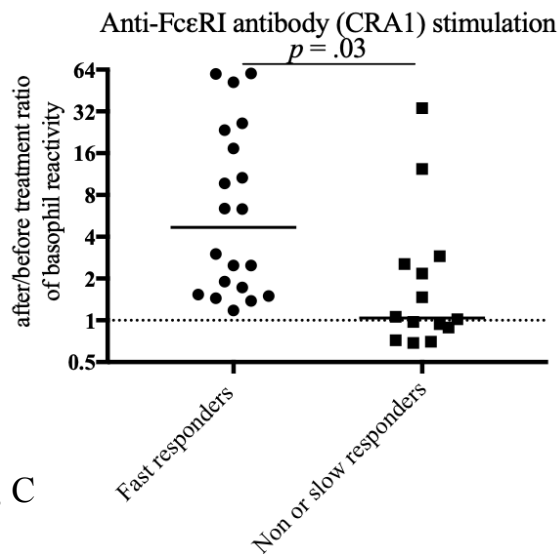


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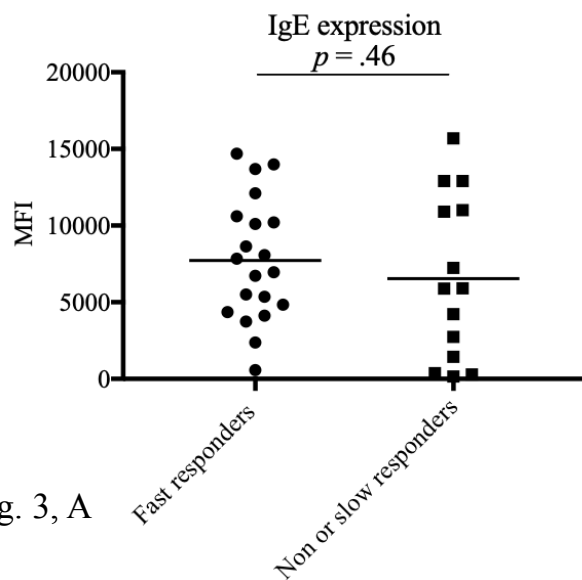


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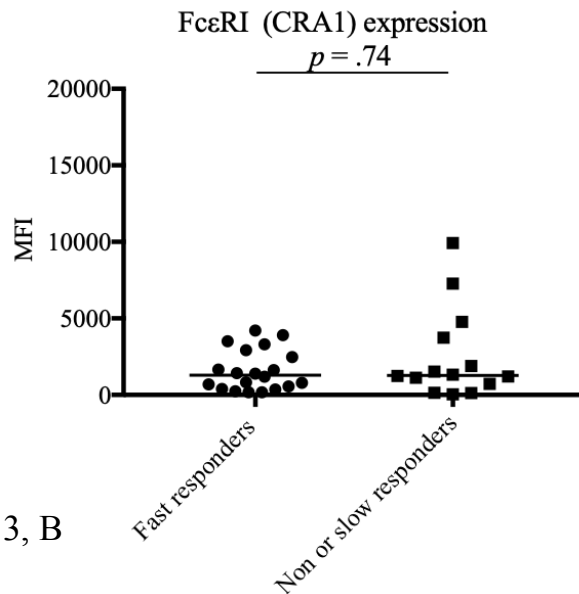


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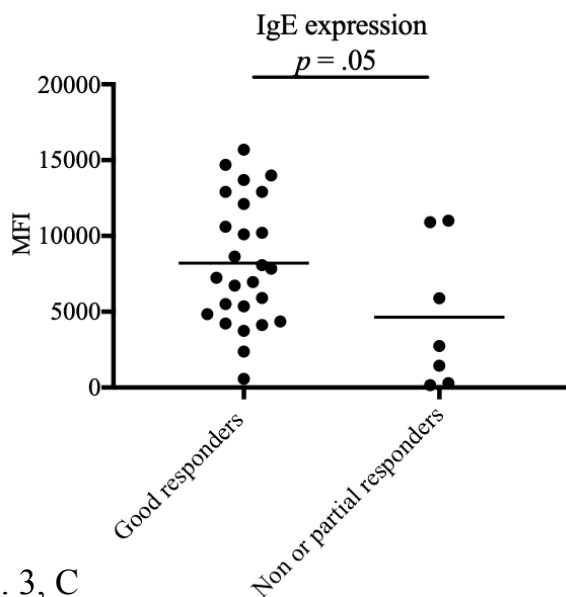


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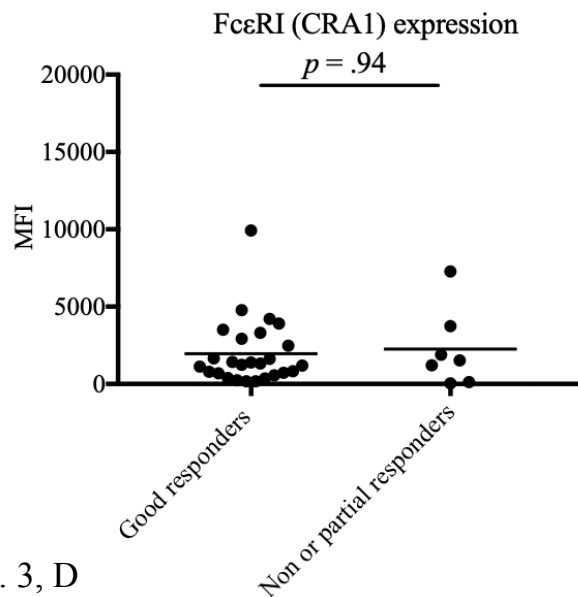


Fig. 3, D

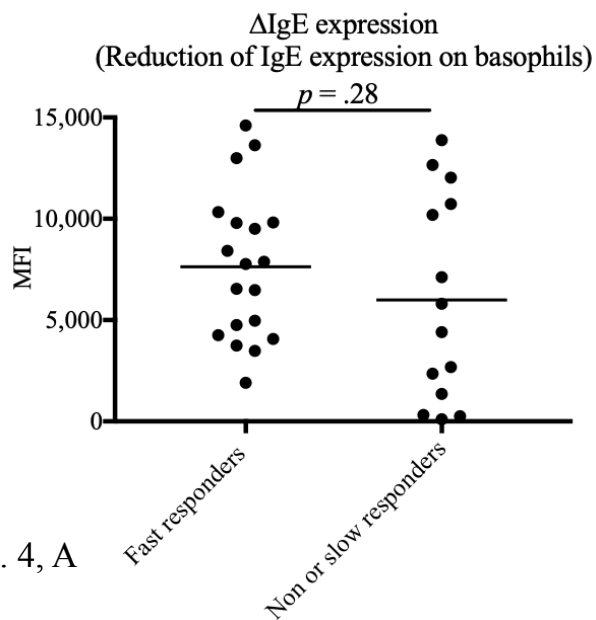


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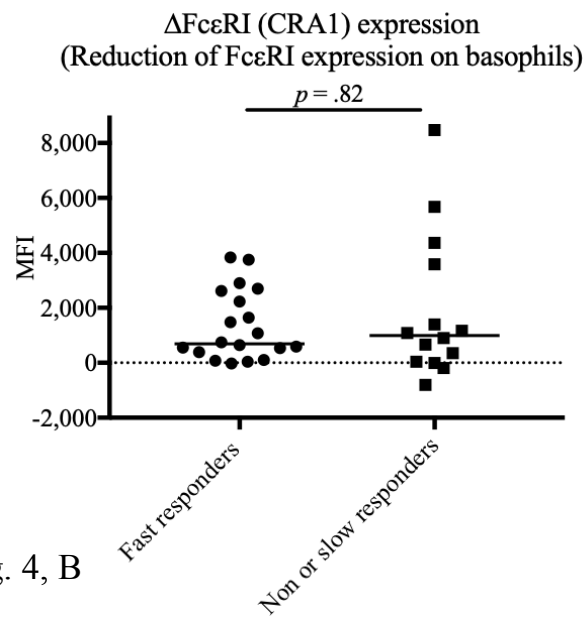


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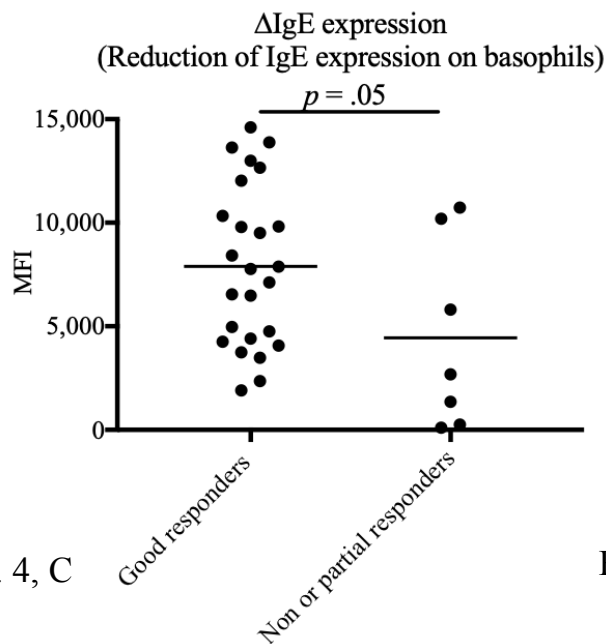


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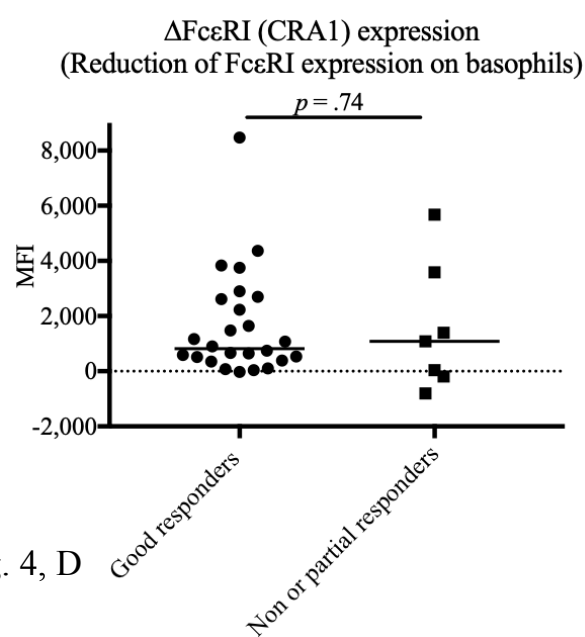


Fig. 4, D

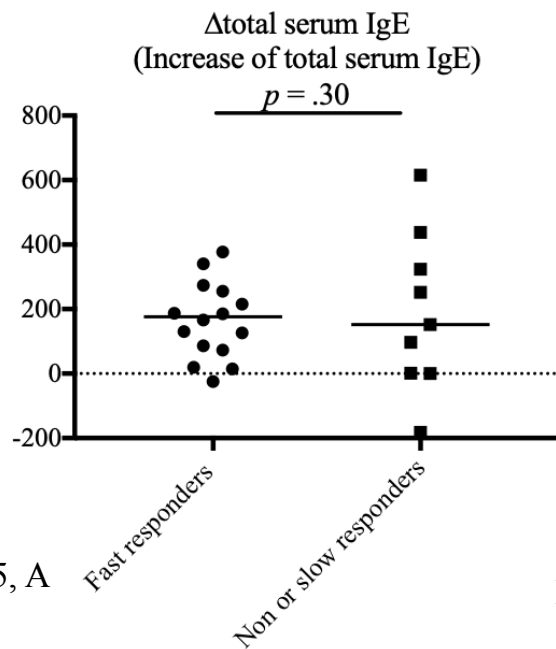


Fig. 5, A

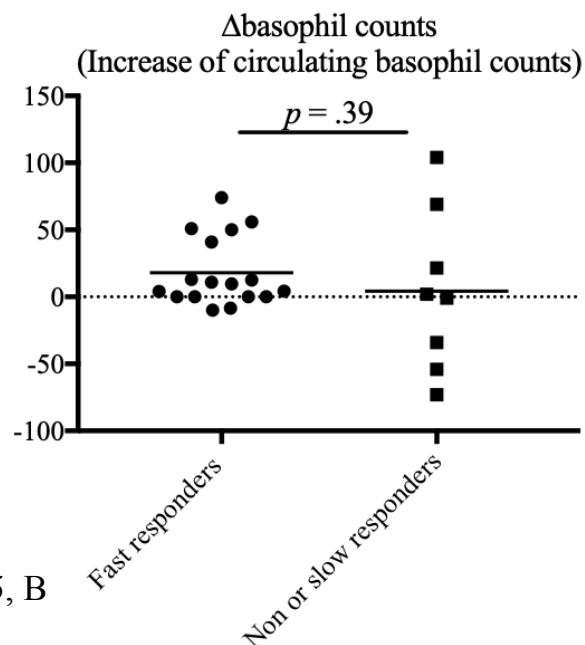


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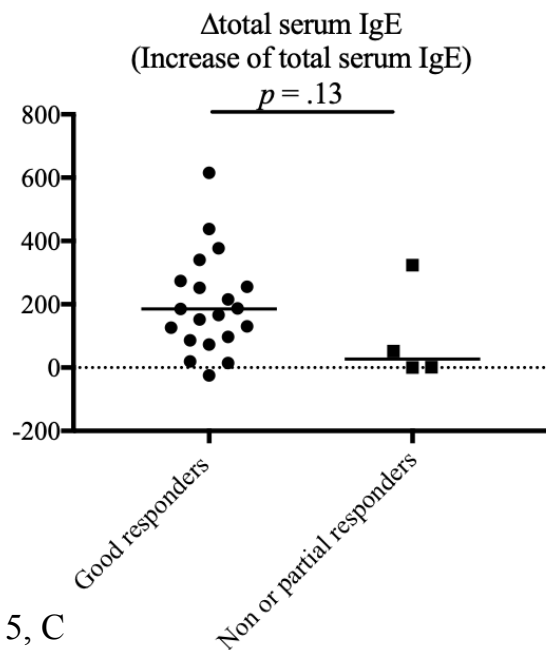


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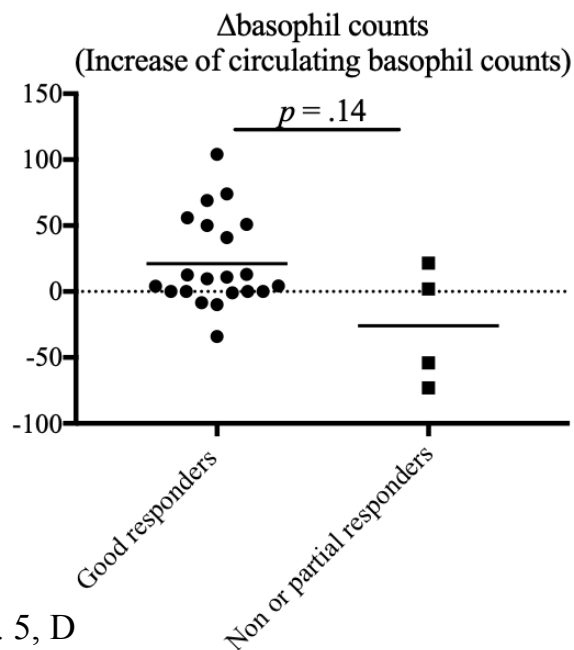


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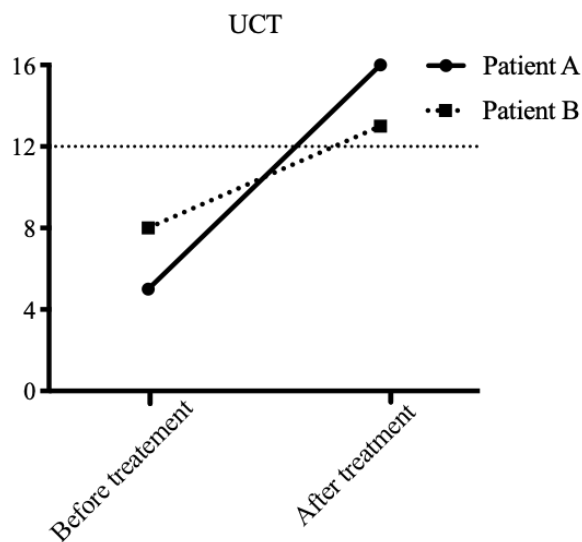


Fig. 6, A

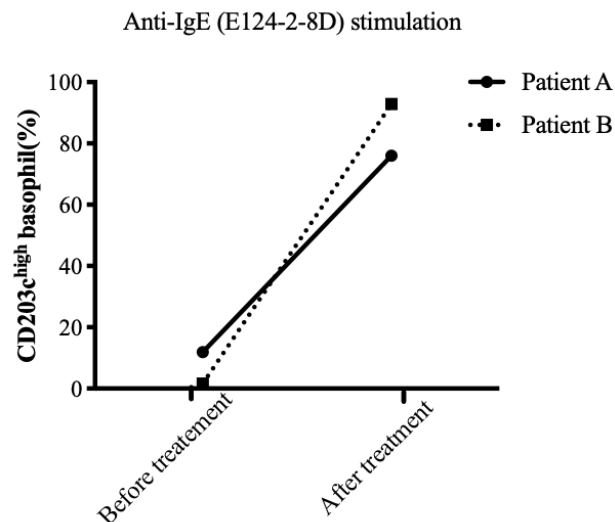


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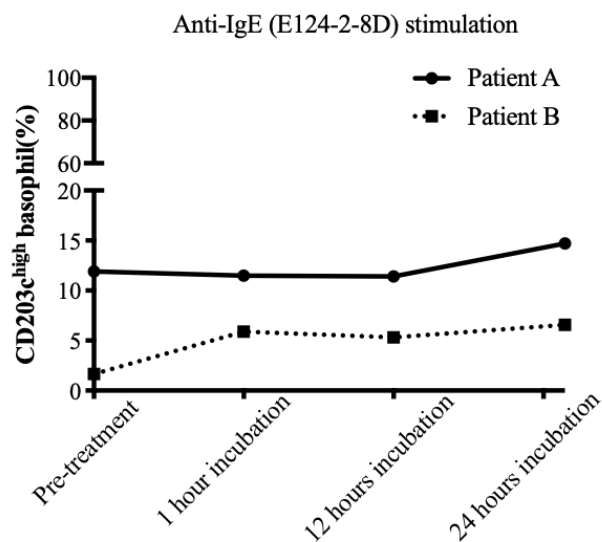


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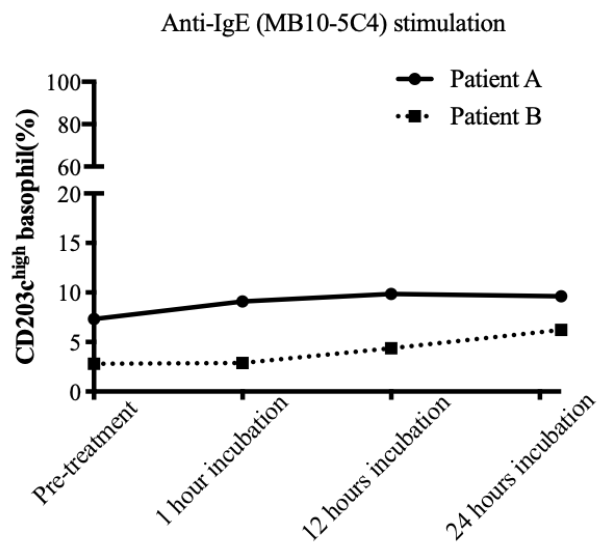


Fig. 6, D

Fig. E1 Definition of each responder to treatment.

Fast responders: UCT scores  $\geq 12$  increased at week 4.

Non or slow responders: UCT scores  $< 12$  increased at week 4.

Good responders: UCT scores  $\geq 12$  increased at week 12.

Non or partial responders: UCT scores  $< 12$  increased at week 12.

Fig. E2 Changes in urticaria control test scores (UCT) and urticaria activity score 7 (UAS7).

A, C. Fast responders.

B, D. Non or slow responders.

The dotted line indicates the cutoff value of 12.

Fig. E3 Comparison of basophil CD203c expression in the steady state without anti-IgE and/or Fc $\epsilon$ RI stimulation (MFI) between fast responders and non or slow responders.

Statistical analysis was performed using the unpaired *t*-test.

MFI, mean fluorescence intensity.

Fig. E4 The response to treatment by classifying responders and non-responders on baseline basophil reactivity

Non-responders:  $< 10\%$  CD203c<sup>high</sup> basophil to anti-IgE stimulation

Responders:  $> 10\%$  CD203c<sup>high</sup> basophil to anti-IgE stimulation

Fig. E5 Changes in CD203c<sup>high</sup> basophils before and after treatment with omalizumab in fast responders (A, C, E) and non or slow responders (B, D, F).

A, B. Anti IgE antibody (E124-2-8D) stimulation.

C, D. Anti IgE antibody (MB10-5C4) stimulation.

E, F. Anti-Fc $\epsilon$ R1 antibody (CRA1) stimulation.

Fig. E6 Correlation of serum total IgE (IU/ml) and basophil IgE expression as MFI in CSU patients.

A. Basophil IgE expression

B. Basophil CRA1 receptor expression

Statistical analyses were performed using Spearman's rank correlation coefficient.

MFI, mean fluorescence intensity; CSU, chronic spontaneous urticaria.

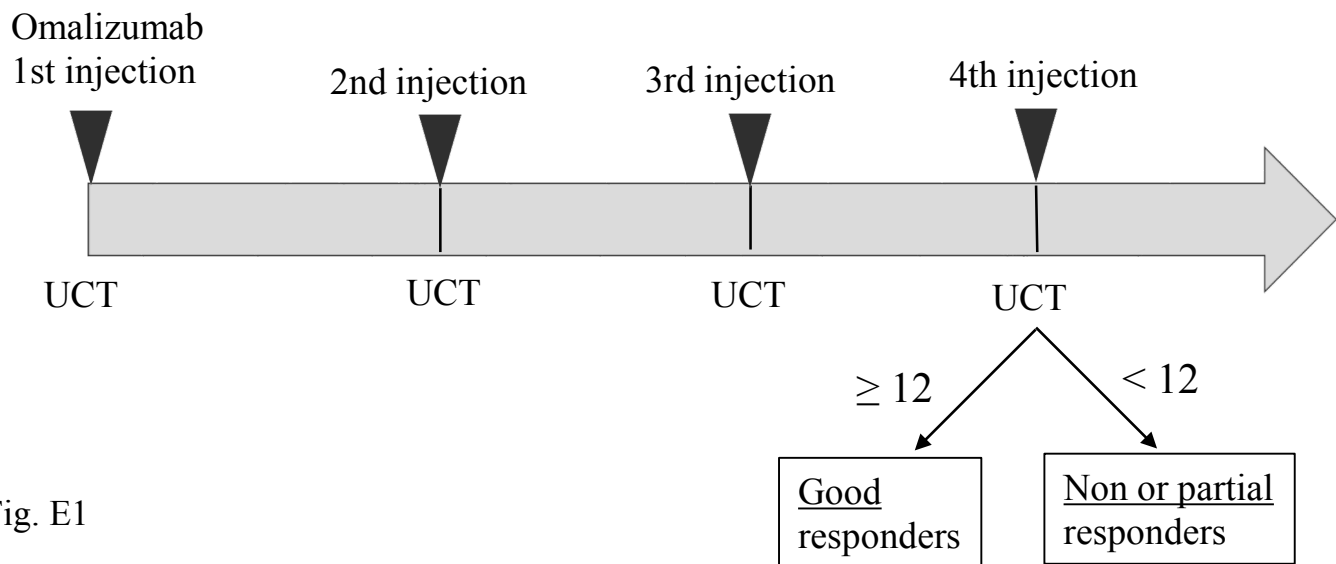
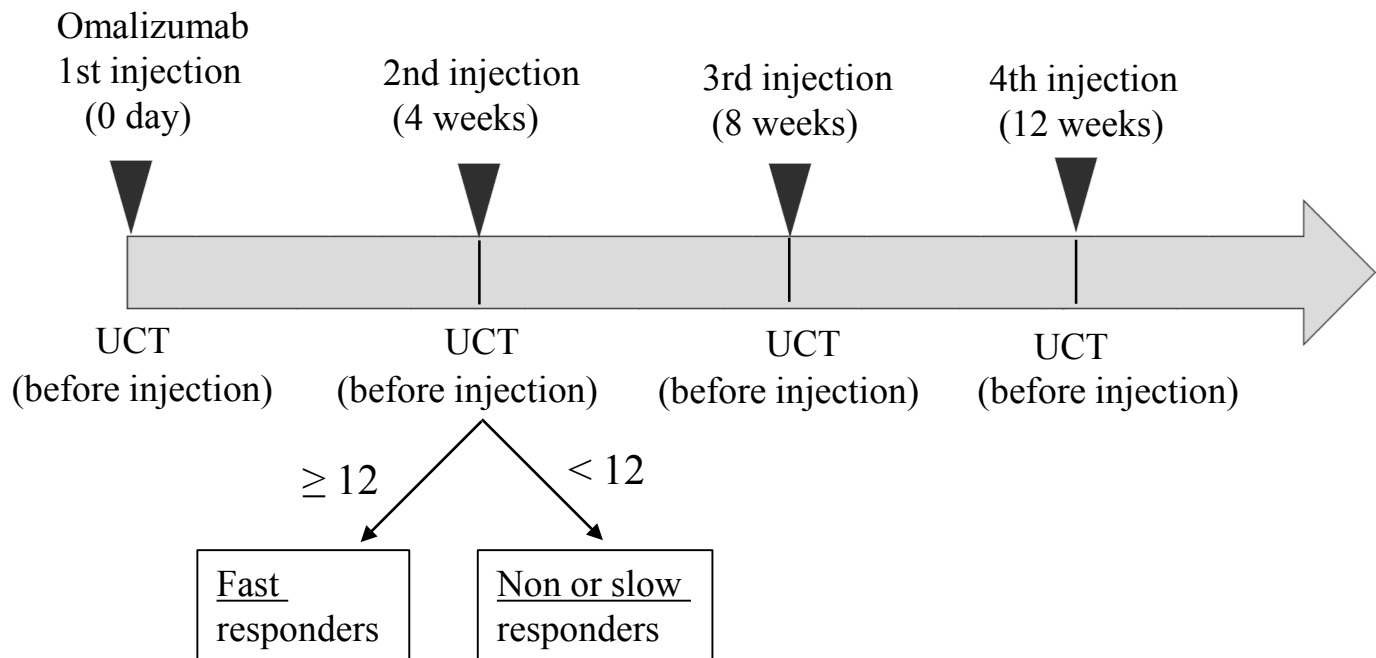


Fig. E1

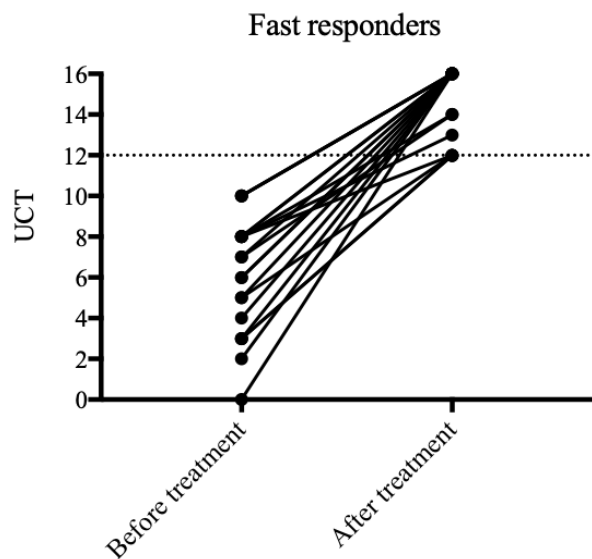


Fig. E2, A

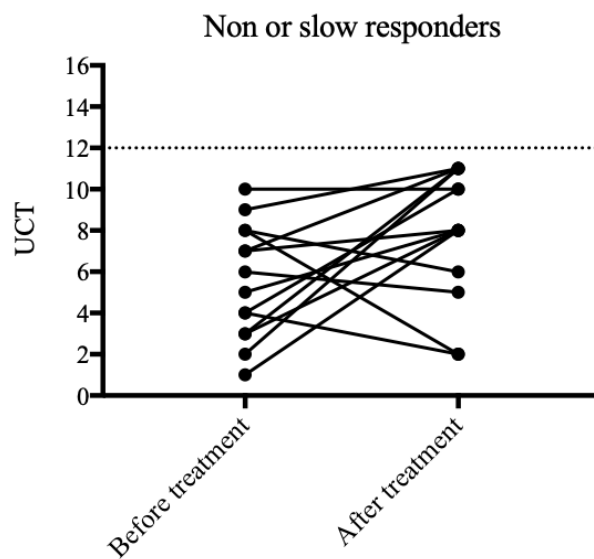


Fig. E2, B

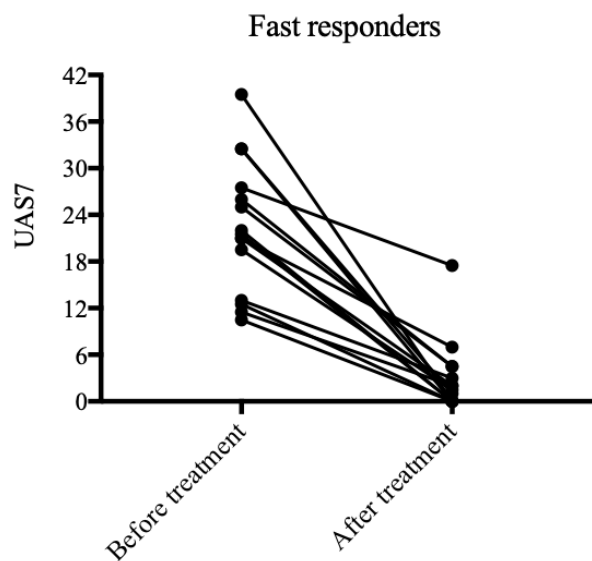


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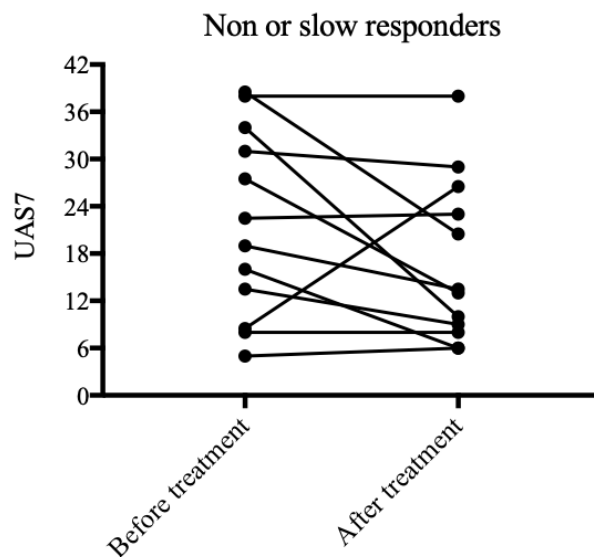


Fig. E2, D

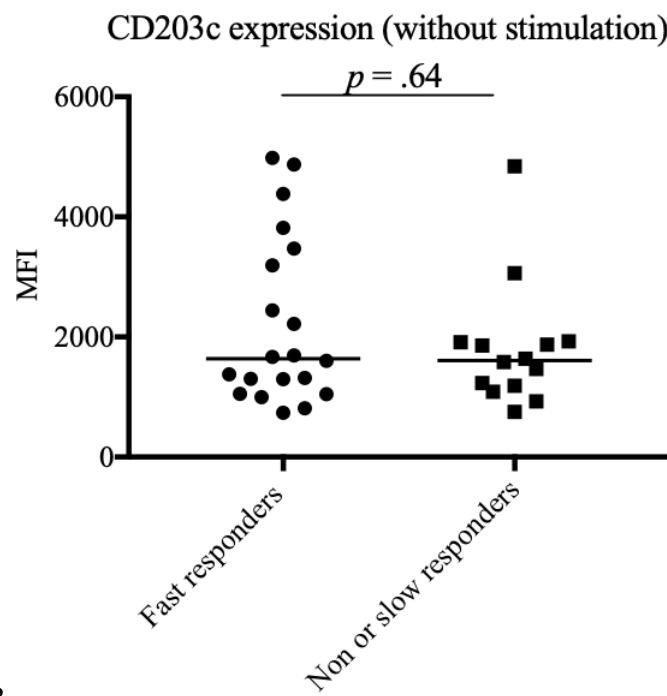


Fig. E3

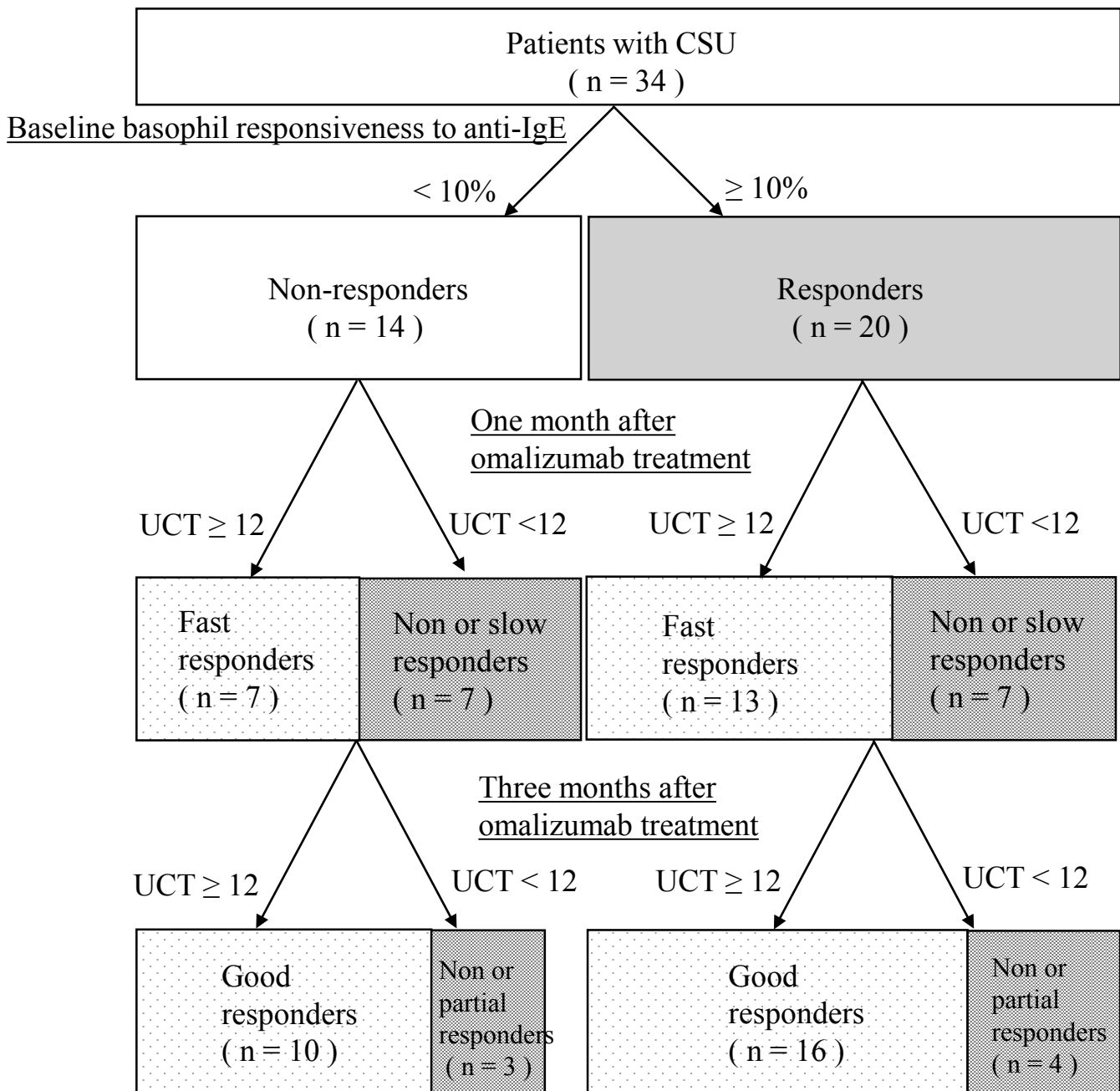


Fig. E4

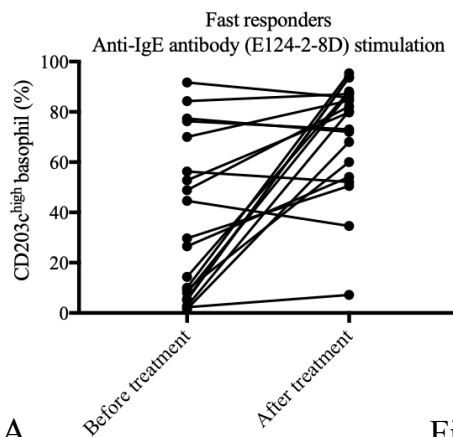


Fig. E5, A

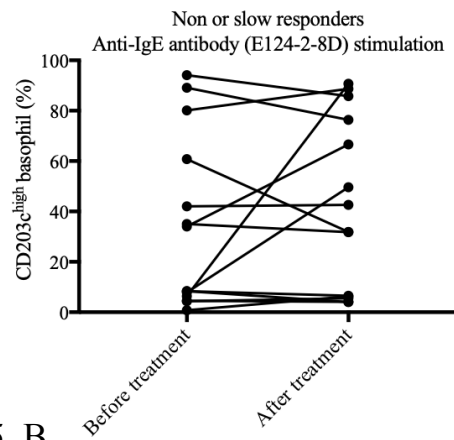


Fig. E5, B

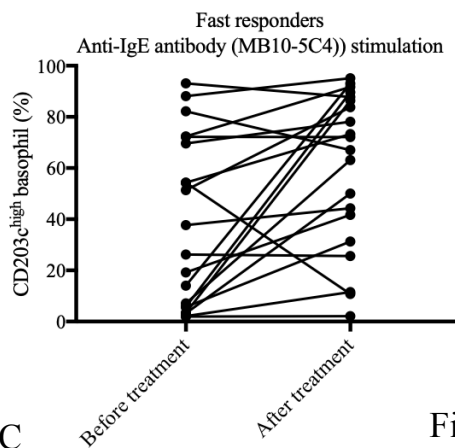


Fig. E5, C

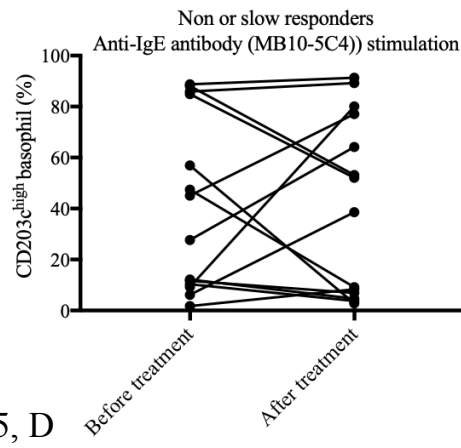


Fig. E5, D

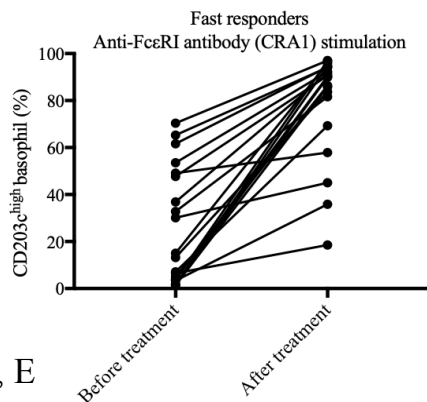


Fig. E5, E

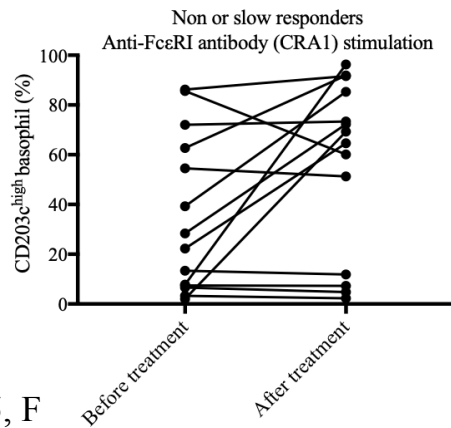


Fig. E5, F

Fig. E6, A

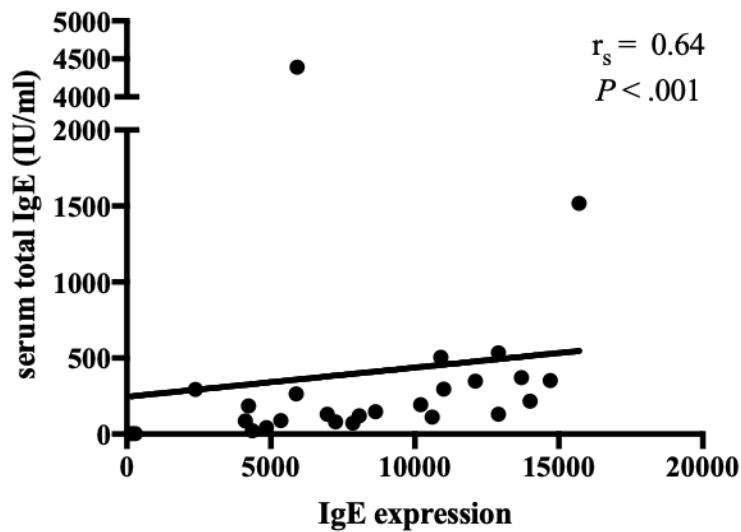


Fig. E6, B

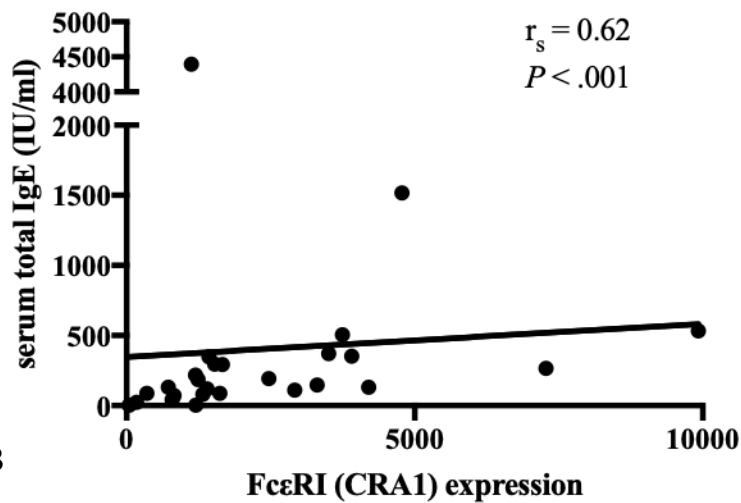




Table EI. Demographic characteristics based on the good therapeutic effect of omalizumab

	Good responders (n = 26)	Non or partial responders (n = 7)	<i>P</i> value
Age, years	45.3 ± 3.6	59.4 ± 7.7	.08
Female, n (%)	18 (69%)	3 (42%)	.37
Disease duration, years	3 (0.2–33)	4 (0.5–25)	.61
Total IgE (IU/ml)	139.5 (22.9–4393)	163.2 (3–505)	.40
Basophil counts (/μl)	18 (0–98)	28 (0–73)	.47
ASST positive rate, n (%)	5/12 (41%)	2/3 (66%)	.56
UCT	6.1 ± 2.8	5.5 ± 2.0	.63
UAS7	23.0 ± 9.1	19.6 ± 12.4	.45
DLQI	7 (2–24)	5.5 (2–14)	.51

Data are given as the mean ± SD for age, UCT and UAS7; n (%) for sex, ASST positive rate; median (range) for disease duration, serum total IgE, basophil counts, and DLQI.

Statistical differences between two groups were analyzed by unpaired *t*-test for age, UCT, and UAS7, Fisher's exact test for female and ASST positive rate, and Mann-Whitney *U*-test for disease duration, serum total IgE, basophil counts, and DLQI.

ASST, autologous serum skin test; UCT, urticaria control test; UAS7, 7-day urticaria activity score; DLQI, dermatology life quality index.

One patient among N/PRs did not have data of total serum IgE and basophil count before treatment and the patient was excluded from the GRs vs. N/PRs dataset.

Table EII. Basophil responsiveness with stimulation before and after treatment with omalizumab in chronic spontaneous urticaria patients

	No	Age	Sex	Total IgE (IU/ml)	Stimulation (proportion of CD203c <sup>high</sup> basophil)					
					Anti-IgE antibody (%) (E124-2-8D)		Anti-IgE antibody (%) (MB10-5C4)		Anti- FcεR1 antibody (%) (CRA1)	
					Before treatment	After treatment	Before treatment	After treatment	Before treatment	After treatment
Fast responders	1	37	M	121	9.28	60	5.68	31.3	7.14	69.3
	2	20	F	131	2.33	7.23	2.14	11.5	3.37	35.9
	3	57	F	148	52.8	79.8	72.2	72.1	32.8	81.6
	4	17	M	111	44.6	34.6	54.3	10.8	49.1	57.9
	5	32	F	73	14.4	88.1	19.2	41.7	15	95.9
	6	49	F	294	29.7	50.5	51.4	83.8	36.9	91.5
	7	35	M	216	87.1	94.2	88.1	95.1	70.4	97
	8	40	F	193	10	95.4	14	93	1.82	94.7
	9	41	F	86.9	1.53	68	1.97	2.1	1.51	89.9
	10	58	F	371	70	84.5	72.4	91.7	53.5	92.3
	11	75	M	22.9	8.03	87.4	7.13	63.1	3.85	90.2
	12	39	M	17.1	77.4	72	69.6	78.1	4.99	86.4
	13	75	M	41.9	76.3	73	82.2	67	65.3	94.1
	14	45	F	88.8	5.13	93.9	3.37	86.4	1.43	85.9
	15	74	F	282	56.3	52.1	37.7	44.3	30.1	45
	16	45	M	347	91.7	85.7	93.1	87.8	61.6	94.3
	17	21	F	2535	2.67	80.3	5.83	89.6	3.69	96.9
	18	74	F	81.5	26.5	54.1	26.2	25.6	13.2	83.7
	19	20	F	n.d.	48.9	82.2	54.3	73.3	47.7	90.6
	20	59	F	n.d	5.63	93.6	3.17	50	6.52	18.5
Non or slow responders	1	73	M	n.d.	42	42.6	47.4	9.11	28.4	72.3
	2	73	M	265	80.1	88.7	84.9	52	62.7	91.9
	3*	23	F	295	34	66.6	45	77	39.3	85.3
	4	51	M	505	8.39	4.04	10.3	3.7	13.4	11.9
	5	63	F	<3	4.55	4.16	12.1	4.64	6.61	4.76
	6	49	M	<3	4.35	5.53	11.6	6.86	7.45	7.26
	7	23	F	534	94.1	85.8	85.9	89.2	86.2	91.6
	8*	61	F	80.2	6.38	90.7	9.21	80	7.82	96.3
	9*	36	F	1518	35	31.8	88	53.1	85.6	60.1
	10*	42	M	4392	8.37	6.52	27.6	64.1	22.3	54.6
	11*	68	F	131	7.91	49.6	6.16	38.5	2.05	69.3
	12*	38	F	185	89.1	76.4	88.7	91.3	72	73.4
	13	54	F	8.4	0.76	6.25	1.67	8.33	3.28	2.25
	14	84	F	7.4	60.8	31.9	56.9	2.78	54.5	51.3

CD203c expression after antibody stimulation before and after treatment with omalizumab is described as CD203c<sup>high</sup> basophils (%). FRs, fast responders (urticaria control test (UCT) scores  $\geq 12$  up to the end of week 4 after starting treatment); N/SR, non or slow responders (UCT scores  $< 12$  up to the end of week 4 after starting treatment).

\* Patients of the 14 N/SRs at 4 week who became GRs at week 12.