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Fujita, Yuichi ; Nagashima, Hiroaki ; Tanaka, Kazuhiro ; Hashiguchi, Mitsuru ; Hirose, Takanori ; Itoh, Tomoo ; Sasayama, Takashi

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The histopathological and radiological features of T2-FLAIR mismatch sign in IDH-

mutant 1p/19q non-codeleted astrocytomas

Yuichi Fujita, MD, PhD 1; Hiroaki Nagashima, MD, PhD 1\*; Kazuhiro Tanaka, MD, PhD 1;

Mitsuru Hashiguchi, MD <sup>1</sup>; Takanori Hirose, MD, PhD <sup>2,3</sup>; Tomoo Itoh, MD, PhD <sup>4</sup>; Takashi

Sasayama, MD, PhD <sup>1</sup>

<sup>1</sup> Department of Neurosurgery, Kobe University Graduate School of Medicine, Kobe, Hyogo,

Japan

<sup>2</sup> Department of Pathology for Regional Communication, Kobe University Graduate School

of Medicine, Kobe, Hyogo, Japan

<sup>3</sup> Department of Diagnostic Pathology, Hyogo Cancer Center, Akashi, Hyogo, Japan

<sup>4</sup> Department of Diagnostic Pathology, Kobe University Graduate School of Medicine, Kobe,

Hyogo, Japan

\*Corresponding author: Hiroaki Nagashima

Department of Neurosurgery, Kobe University Graduate School of Medicine

7-5-1 Kusunoki-cho, Chuo-ku, Kobe, Hyogo 650-0017, Japan

Phone: +81-78-382-5966; Fax: +81-78-382-5979

E-mail: hn0628hn@med.kobe-u.ac.jp

**Key words:** Glioma; Astrocytoma; IDH mutation; 1p/19q codeletion; T2-FLAIR mismatch

sign; Apparent diffusion coefficient

**Short title:** Patho-radiological feature of mismatch sign

- 1 The histopathological and radiological features of T2-FLAIR mismatch sign in IDH-
- 2 mutant 1p/19q non-codeleted astrocytomas

- 4 Abstract
- 5 **Objective:** The T2-FLAIR mismatch sign is a useful imaging sign in clinical MRI studies for
- 6 detecting isocitrate dehydrogenase (IDH)-mutant 1p/19q non-codeleted astrocytomas.
- 7 However, the association between the mismatch sign and pathological findings is poorly
- 8 understood. Therefore, the aim of this study was to elucidate the relationship of
- 9 histopathological and radiological features with the mismatch sign in IDH-mutant 1p/19q
- 10 non-codeleted astrocytomas.
- Methods: We divided 17 IDH-mutant 1p/19q non-codeleted patients into two groups
- according to mismatch sign presence (WITH, n = 9; WITHOUT, n = 8) and retrospectively
- analyzed their pathological findings and apparent diffusion coefficient (ADC) values. We also
- compared these findings between the tumor Core (central area) and Rim (marginal area).
- 15 **Results:** In the pathological analysis, Core of the WITH group contained numerous
- microcysts whereas Rim had abundant neuroglial fibrils and cellularity. In contrast, Core of
- the WITHOUT group had highly concentrated neuroglial fibrils. In ADC analysis, Core of the
- WITH group had significantly higher ADC values compared with Rim (p < 0.001). However,
- there was no significant difference between Core and Rim in the WITHOUT group (p =
- 20 0.12). The WITH group had a significantly higher Core/Rim ratio of ADC values compared
- with the WITHOUT group (p < 0.001).
- 22 **Conclusions:** This study provides evidence that a region-dependent microstructural
- 23 difference could reflect the mismatch sign in IDH-mutant 1p/19q non-codeleted
- 24 astrocytomas. Core of the mismatch sign characteristically had microcystic changes
- accompanied by higher ADC values, whereas Rim had abundant neuroglial fibrils and

cellularity accompanied by lower ADC values.

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### INTRODUCTION

The World Health Organization (WHO) Classification of Tumors of the Central Nervous 29 System was updated in 2016, leading to the present era of diagnosis based on molecular 30 characteristics rather than histopathology alone. These molecularly oriented groups more 31 accurately reflect the biological characteristics and clinical behavior of tumors.<sup>2–5</sup> Isocitrate 32 dehydrogenase (IDH) gene status and 1p/19q status in particular are recognized as key 33 genetic aberrations in lower-grade gliomas. 4,6 Preoperative detection of these molecular 34 characteristics might enable more favorable treatment strategies for patients. Several studies 35 have attempted preoperative prediction based on various imaging features, including location, 36 37 presence of calcification, contrast-enhancing pattern, perfusion imaging, magnetic resonance spectroscopy, and positron emission tomography.<sup>7–14</sup> Indeed, a multiparametric approach 38 combining these imaging features has been shown to improve diagnostic accuracy. Possible 39 clinical applications of radiogenomics, known as imaging genomics, have also been 40 discussed, including deep learning, but the field is still under development.<sup>15</sup> 41 T2-FLAIR mismatch sign was recently identified and found to be specific to IDH-mutant 42 1p/19q non-codeleted astrocytomas as a single parameter. 16-18 The radiological feature of the 43 mismatch sign is defined as complete/near-complete homogeneous hyperintense signal on 44 45 T2-weighted imaging (T2WI) and simultaneously complete/near-complete homogeneous hypointense signal on FLAIR with hyperintense peripheral region on FLAIR. The mismatch 46 sign is clinically useful because it is a simple feature that is obtainable with conventional 47 magnetic resonance imaging (MRI) sequences. However, its main limitation is its lower 48 sensitivity. Indeed, nearly half of patients with IDH-mutant 1p/19q non-codeleted 49 astrocytomas do not have the mismatch sign. It is still unclear what the presence or absence 50

51 of the mismatch sign within IDH-mutant 1p/19q non-codeleted astrocytomas represents. Moreover, the mismatch sign is not a prognostic factor for IDH-mutant 1p/19g non-codeleted 52 astrocytomas. 16,19 In addition, the relationship between the mismatch sign and the 53 microenvironment in IDH-mutant 1p/19q non-codeleted astrocytomas is unknown. Therefore, 54 to resolve these limitations, elucidation of the background resulting in the mismatch sign is 55 necessary. Another mismatch sign is cerebrospinal fluid, which shows a hyperintense signal 56 on T2WI and a hypointense signal on FLAIR. 57 Accordingly, we hypothesized that tumors with the mismatch sign might have abundant water 58 59 content in the form of such as microcysts, which meant larger intercellular space or lower cellularity in the core region that is hypointense on FLAIR. Furthermore, we surmised the 60 existence of a region-dependent microstructural difference between the central (hypointense 61 62 on FLAIR) and marginal (hyperintense on FLAIR) regions within tumors with the mismatch sign. The apparent diffusion coefficient (ADC) value, which reflects the diffusivity of water 63 molecules in tissue, can capture the microstructural difference underlying the mismatch 64 sign.<sup>20,21</sup> To test these hypotheses, we analyzed the histopathological and radiological features 65 of the mismatch sign in IDH-mutant 1p/19q non-codeleted astrocytomas. 66 67 MATERIAL AND METHODS 68 Study design and patients 69 70 The study was approved by the institutional review board at our institution (protocol number 180359) and conducted according to the institutional and national ethical guidelines and in 71 accordance with the Helsinki Declaration. 72 Between January 2009 and July 2020, 106 of 325 consecutive patients with glioma surgically 73 treated at our institution were histopathologically confirmed to have lower-grade glioma. Of 74 these 106 patients, 80 were selected using the following criteria: newly diagnosed WHO 75

76 grade II or III intracranial glioma confirmed by histopathology according to 2016 WHO guidelines<sup>1</sup>, full information on IDH and 1p/19q status, and adequate preoperative MRI scans 77 (T2WI, T2-weighted FLAIR, and diffusion-weighted imaging [DWI]). The remaining 26 78 patients were excluded because of a lack of pathological and molecular information (n = 14), 79 recurrence (n = 9), lack of preoperative T2WI or FLAIR (n = 2), or predominant lesion 80 location in the spinal cord (n = 1). Of the 80 patients, 17 were confirmed to have IDH-mutant 81 and 1p/19q non-codeleted astrocytomas and were enrolled in this study. A flow chart 82 outlining the patient selection process is shown in Figure 1. 83 84 We analyzed the presence of the mismatch sign and divided the IDH-mutant 1p/19q noncodeleted astrocytomas into two groups—WITH and WITHOUT—according to its presence. 85 The mismatch sign was identified according to the following criteria: the presence of an 86 87 almost homogeneous hyperintense signal on T2WI with a conversely hypointense signal on FLAIR, except for a hyperintense peripheral region. 16-18 If there was disagreement 88 concerning the mismatch sign between two independent reviewers (Y.F. and T.S.), who were 89 90 blinded to clinical information, a third reviewer (H.N.) was involved in the assessment. Finally, we analyzed the region-dependent changes in histopathological features and ADC 91 values, as described in the following subsections. In the WITH group, we defined "Core" as a 92 region showing a hyperintense signal on T2WI with a conversely hypointense signal on 93 FLAIR and "Rim" as a region showing a hyperintense signal on both T2WI and FLAIR. In 94 the WITHOUT group, we defined Core as the central region and Rim as the marginal region. 95 96 Histopathological analysis 97 The histopathological diagnoses of patients made between 2009 and 2015 were reevaluated 98 according to 2016 WHO guidelines. 1 IDH status was confirmed by immunohistochemistry 99 and DNA sequencing as previously described. 22,23 IDH status was first screened by an IDH1 100

R132H mutation-specific antibody. In negative cases, IDH mutation status was assessed by a clinically validated DNA sequencing. Fluorescence in situ hybridization (FISH) was applied to assess 1p/19q status as previously described <sup>22,24</sup>. The FISH probes 1p36/1q25 and 19q13/19p13 were used to target 1p and 19q, respectively. A probe ratio < 0.75 was diagnosed as a loss.

All surgeries were performed using a neuronavigation system. The site corresponding to the evaluation tissue was confirmed on the MRI data intraoperatively registered for the patient using iPlan Cranial (BrainLAB AG, Feldkirchen, Germany). The tissue corresponding to Core was assessed by two independent reviewers (Y.F. and T.S.). Moreover, we obtained multiple samples from Core and Rim in representative patients with the mismatch sign and separately analyzed their features.

# **Imaging analysis**

Conventional MRI sequences were obtained with a 3.0-T MRI scanner (Achieva; Philips Medical Systems, Eindhoven, The Netherlands) and included T2WI (repetition time [TR]/echo time [TE], 4200/100 ms; field of view [FOV], 230 mm; matrix, 320 × 400; slice thickness, 5.0 mm); T2-weighted FLAIR (TR/TE/inversion time [TI], 11000/125/2800 ms; FOV, 230 mm; matrix, 204 × 256; slice thickness, 5.0 mm); DWI (b-values, 0 and 1000 s/mm<sup>2</sup>; TR/TE 4500/75 ms; FOV, 240 mm; slice thickness, 4 mm; slice gap, 1 mm; matrix, 109 × 128; flip angle, 90°); and three-dimensional T1-weighted imaging (T1WI; TR/TE, 5.0/2.2 ms; FOV, 240 mm; matrix,  $304 \times 304$ ; slice thickness, 0.8 mm) before and after injection of intravenous gadolinium contrast agent (0.2 ml/kg; Magnescope, Guerbet, Paris, France). The ADC values from DWI of two types (b-values 0 and 1000 s/mm<sup>2</sup>) were calculated per voxel to create the ADC map. An region of interest (ROI) measuring 10 mm<sup>2</sup> was manually drawn on a representative FLAIR slice and then reflected onto the ADC map. 

Ten ROIs were each set with a wide and balanced distribution in Core and Rim. The means of the ADC values within the ROIs were used for the statistical analysis. The ROIs were drawn by a single neurosurgeon (Y.F.) and confirmed by two independent reviewers (T.S. and H.N.).

## Statistical analysis

The inter-reviewer agreement concerning the mismatch sign was assessed using Cohen's Kappa statistic ( $\kappa \le 0.4$ , poor agreement;  $0.4 < \kappa \le 0.6$ , moderate agreement;  $0.6 < \kappa \le 0.8$ , good agreement;  $0.8 < \kappa$ , substantial agreement). The characteristics of each group were compared using the Mann–Whitney U test and Fisher's exact test. The Mann–Whitney U test was used to compare ADC values between groups (WITH and WITHOUT) and regions (Core and Rim). All statistical analyses were performed with EZR (Saitama Medical Center, Jichi Medical University, Saitama, Japan), which is a graphical user interface for R (The R Foundation for Statistical Computing, Vienna, Austria). A two-sided p-value < 0.05 was considered statistically significant.

### **RESULTS**

### **Patient characteristics**

In total, 17 patients with IDH-mutant and 1p/19q non-codeleted astrocytomas were included in this study. The mismatch sign was found in 9 of the 17 (53%). The inter-reviewer consensus for the mismatch sign indicated substantial agreement ( $\kappa$  = 0.88). There were 9 patients (7 men, 2 women; median age, 36 [range, 19–41] years) in the WITH group and 8 patients (1 man, 7 women; median age, 68 [range, 29–76] years) in the WITHOUT group. Patient characterics in both groups are summarized in Table 1. Patients with the mismatch sign were significantly younger than those without it (p = 0.007). The mismatch sign was more common in men (p = 0.01). There were no significant between-group differences in

151	their other characteristics: preoperative Karnofsky Performance Status score, tumor location,
152	tumor size, contrast-enhancing rate, extent of resection, WHO grade, and MIB-1 index at
153	baseline.
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155	Histopathological features of the mismatch sign
156	First, regarding the tissue corresponding to Core, the tumor matrix of the WITH group
157	contained numerous and variably sized microcysts. In contrast, the tumor matrix of the
158	WITHOUT group was predominantly composed of abundant neuroglial fibrils (Fig. 2).
159	Second, in terms of the region-dependent changes in the WITH group, Rim was composed of
160	abundant neuroglial fibrils and cellularity, which contrasted with the numerous microcysts in
161	Core (Fig. 3).
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163	Correlation between the mismatch sign and ADC values
164	The ADC values (expressed as $10^{-6}$ mm <sup>2</sup> /s), which were based on a complete evaluation that
165	included both Core and Rim, were significantly higher in the WITH group than in the
166	WITHOUT group (1703.5 $\pm$ 454.7 vs 1391.0 $\pm$ 182.9; p = 0.004). The region-dependent
167	changes in ADC values were also analyzed in each group. In the WITH group, Core had
168	significantly higher ADC values than Rim (1998.5 $\pm$ 118.1 vs 1135.0 $\pm$ 188.3; p < 0.001).
169	However, in the WITHOUT group, there was no significant difference between Core and Rim
170	$(1415.5 \pm 198.1 \text{ vs } 1388.0 \pm 163.0; p = 0.12)$ . Furthermore, the Core/Rim ratio of ADC
171	values was analyzed. The WITH group had a significantly higher Core/Rim ratio than the
172	WITHOUT group $(1.8 \pm 0.2 \text{ vs } 1.1 \pm 0.1; p < 0.001)$ (Fig. 4).
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174	DISCUSSION
175	Detection of the molecular characteristics of gliomas based on preoperative imaging is a

"virtual biopsy". 15 Noninvasive preoperative diagnosis of molecularly oriented groups in lower-grade gliomas might enable more favorable treatment strategies for patients. The mismatch sign is so specific to IDH-mutant 1p/19q non-codeleted astrocytomas that it has great potential for clinical impact. 16-18,26-28 In this study, we determined that the microstructural difference in IDH-mutant 1p/19q non-codeleted astrocytomas was reflected in the mismatch sign. The tumor matrix corresponding to Core of the mismatch sign contained numerous microcysts, which contrasted with the abundant neuroglial fibrils and cellularity in Rim. Furthermore, Core had significantly higher ADC values than Rim within the mismatch sign. These histopathological and radiological region-dependent differences were unique to the mismatch sign. The histopathological feature of Core in the WITH group—a microcystic change—was comparable to that of conventional protoplasmic astrocytoma. Tay et al.<sup>29</sup> reported signal suppression on FLAIR in protoplasmic astrocytoma. Patel et al. 17, who discovered the mismatch sign from The Cancer Genome Atlas (TCGA)/ The Cancer Imaging Archive (TCIA) cohort, reported that tumors with the mismatch sign tended to have more microcysts. In this study, we confirmed that the microcystic changes were observed only in Core in all of the WITH group. Rim in the WITH group inversely had higher cellularity and more neuroglial fibrils than Core. Our findings suggest that the mismatch sign is linked to the region-dependent histopathological differences. Based on the previous edition of the WHO guidelines, tumors with the mismatch sign might be misdiagnosed as fibrillary astrocytoma by sampling from Rim when they should be accurately diagnosed as protoplasmic astrocytoma by sampling from Core. For this reason, epidemiologically, the mismatch sign with the protoplasmic feature was proportionally more common than the conventional protoplasmic astrocytoma. 16-18,28,29 The updated WHO guidelines overcame the problem of histopathological diagnosis and now advocate for diagnosis based on molecular

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characteristics. Although the mismatch sign does not cover all IDH-mutant 1p/19q noncodeleted astrocytomas, it can be used to accurately detect the same molecularly oriented groups, in line with the aim of the updated guidelines. We were also able to identify a significant difference in ADC values between Core and Rim in the WITH group. This radiological feature was not found in the WITHOUT group. ADC imaging accurately reflects the diffusivity of water molecules. Tumors with high cellularity have a narrower intercellular space, which restricts water movement. <sup>20,21</sup> In general, regions with higher cellularity show lower ADC values. Our ADC findings suggest that Core had less cellularity and a larger intercellular space compared with Rim within the mismatch sign. This finding was consistent with the histopathological findings. Thus, a mismatch sign reflecting microstructural differences was successfully identified using our histopathological and radiological findings. The mismatch sign accurately represented at least the microstructural subgroup of IDH-mutant 1p/19q non-codeleted astrocytomas. However, our study also has several limitations. First, the study was conducted at a single institution and the number of patients was small. The mismatch sign was observed in 53% of IDH-mutant 1p/19q non-codeleted astrocytomas in our study, which was about the same as previously reported values (39%–73%). 16–18 Interestingly, one new finding was that the mismatch sign was significantly more common in younger males within IDH-mutant 1p/19q non-codeleted astrocytomas. Johnson et al. 30 reported exceptional cases with the mismatch sign, and they were all younger males. Sex differences in glioblastoma were recently revealed by transcriptome analysis.<sup>31</sup> Although the sex differences in lower-grade gliomas are not sufficiently understood, the mismatch sign might be linked to sex-specific molecular subgroups of IDH-mutant 1p/19q non-codeleted astrocytomas. Second, the ROIs for the ADC values reflected some, but not all, of the regions. However, our ADC findings supported the histopathological findings. In particular, a Core/Rim ratio of the ADC value exceeding 1 in

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all cases of the WITH group was one of the supportive results and it was linked to the region-dependent histopathological differences. Third, this study could not fully examine false-positive mismatch signs. Onishi et al. 32 reported the mismatch sign in dysembryoplastic neuroepithelial tumor. The histopathological hallmark of dysembryoplastic neuroepithelial tumor is a so-called specific glioneuronal element with a mucoid-rich background. This feature closely resembles the central region of IDH-mutant 1p/19q non-codeleted astrocytomas with the mismatch sign. This indicated that tumors with similar histopathological microstructures (microcystic changes/mucoid-rich changes) might show the mismatch sign, although there was no histopathological evaluation of the marginal region of dysembryoplastic neuroepithelial tumor. Finally, further exploratory research is necessary to elucidate how to make the best use of the mismatch sign as one parameter of a multiparametric approach to virtual biopsy. Nevertheless, our finding that the mismatch sign reflected the microstructural difference in IDH-mutant 1p/19q non-codeleted astrocytomas is clinically important.

### **CONCLUSION**

This study provides evidence that the microstructural difference in IDH-mutant 1p/19q non-codeleted astrocytomas is reflected in the mismatch sign. The central region of the mismatch sign had the characteristic microcystic changes accompanied by higher ADC values, in contrast to the marginal region, which had abundant neuroglial fibrils and cellularity accompanied by lower ADC values. These changes were unique to IDH-mutant 1p/19q non-codeleted astrocytomas with the mismatch sign and were distinguishable from those of IDH-mutant 1p/19q non-codeleted astrocytomas without the mismatch sign.

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367	Figure legends
368	Fig. 1 Flow chart showing the patient selection process.
369	DIPG, diffuse intrinsic pontine glioma; IDH, Isocitrate dehydrogenase
370	
371	Fig. 2 Photomicrographs showing tissue corresponding to Core in the WITH group (a, b) and
372	the WITHOUT group (c, d). Hematoxylin and eosin staining; original magnification, ×200.
373	
374	Fig. 3 A representative case in the WITH group. The squares in the FLAIR image show the
375	sampling locations corresponding to Core (gray color) and Rim (black color).
376	Photomicrographs respectively show tissue corresponding to Core and Rim. Hematoxylin and
377	eosin staining; original magnification, ×200.
378	
379	Fig. 4 Representative cases in the WITH group (a) and the WITHOUT group (b). FLAIR
380	(middle) and the ADC map (right) demonstrate representative settings for the ROIs. The
381	small circles corresponding to Core (gray color) and Rim (black color) indicate the ROIs used
382	for analysis of the ADC values. Box plots of ADC values are shown for groups (c) and
383	locations in each group (d, e). The box plot of the Core/Rim ratio of the ADC values is shown
384	for the WITH and WITHOUT groups (f). ** $p < 0.01$ , *** $p < 0.001$ , Mann–Whitney $U$ test.
385	
386	Table 1. Patient characteristics
387	Abbreviations: WHO World Health Organization

Fig. 1

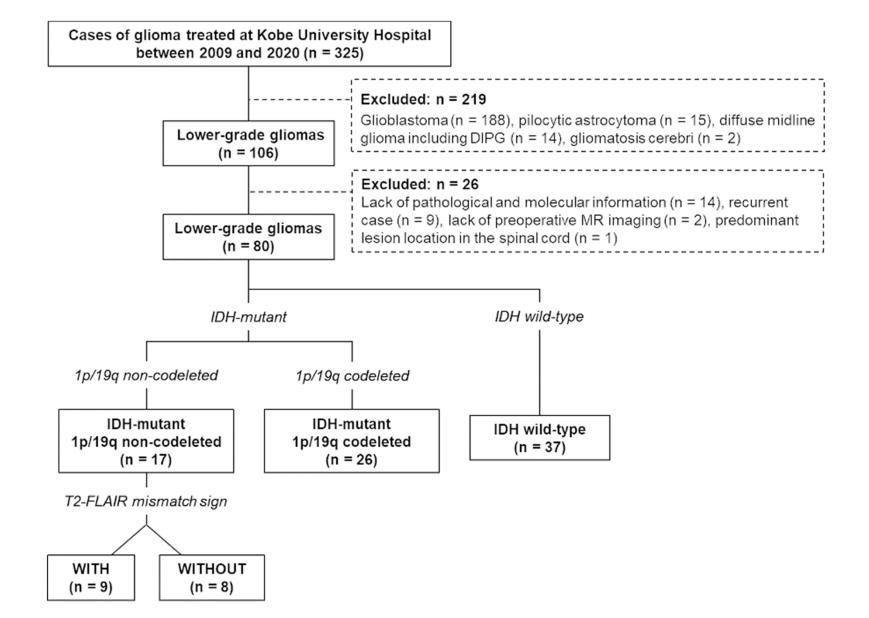


Fig. 2

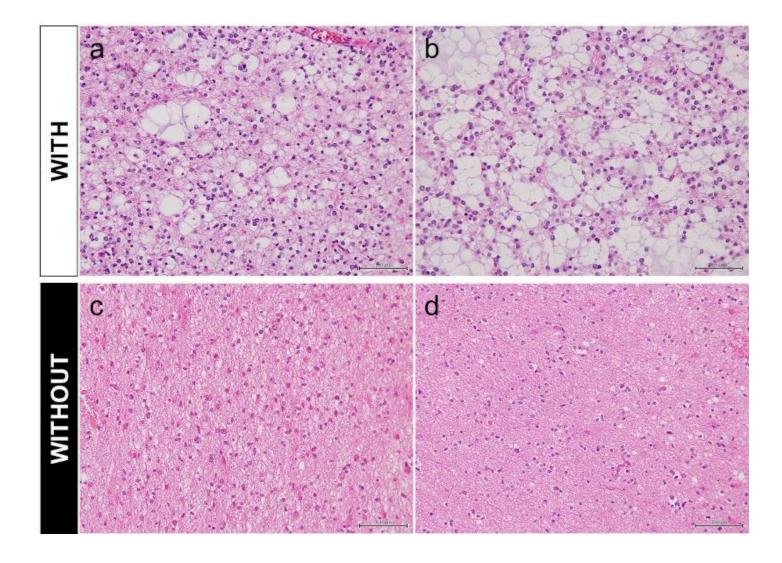


Fig. 3

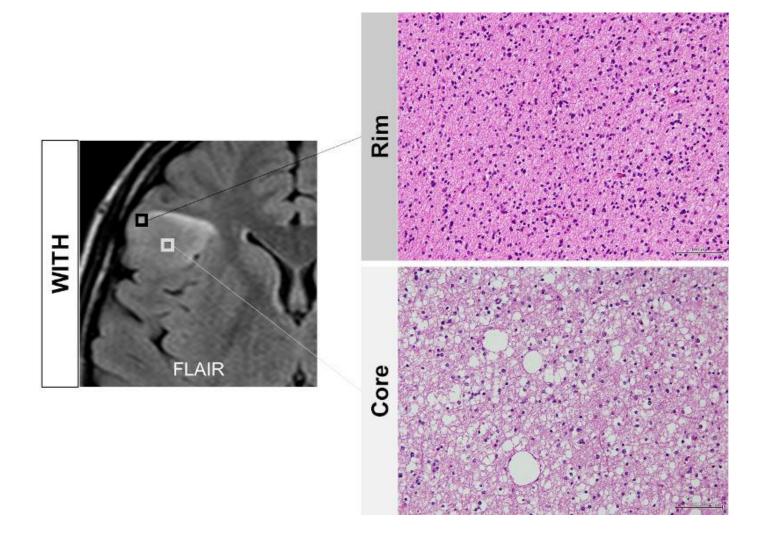


Fig. 4

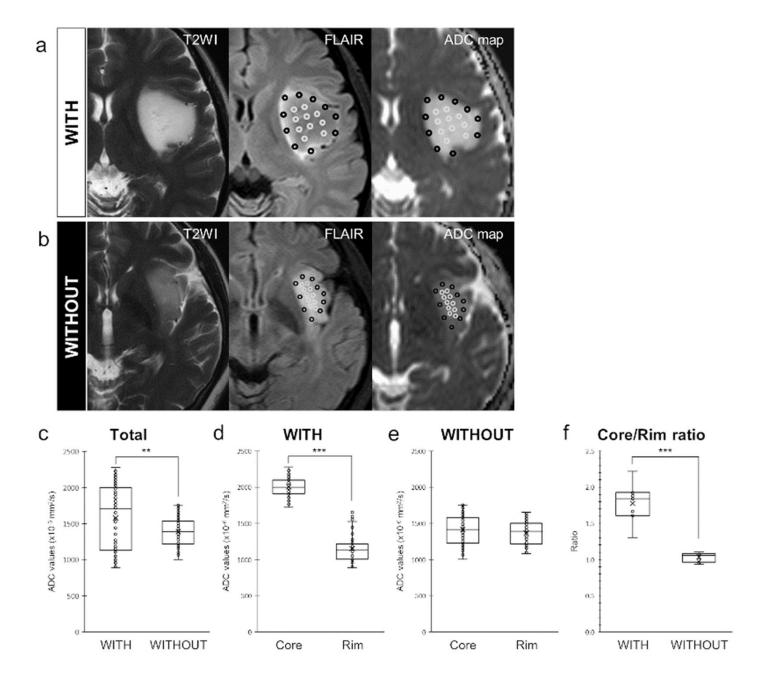


Table 1 Patient characteristics

	WITH	WITHOUT	D 1/ 1
Characteristic	(n = 9)	(n=8)	P Value
Age, years			
Median (range)	36 (19–41)	68 (29–76)	0.007
Sex, n (%)			
Male	7 (77.8)	1 (12.5)	0.01
Female	2 (22.2)	7 (87.5)	
Preoperative Karnofsky Performance Status			
Median (range)	100 (90–100)	90 (70–100)	0.13
Tumor location, n (%)			
Frontal	5 (55.6)	3 (37.5)	0.43
Temporal	3 (33.3)	3 (37.5)	
Insular	1 (11.1)	2 (25.0)	
Laterality, n (%)			
Right	7 (77.8)	5 (62.5)	0.54
Left	2 (22.2)	3 (37.5)	
Tumor size (FLAIR), mm			
Median (range)	55 (27–78)	55 (20–74)	0.92
< 6 cm	5 (55.6)	5 (62.5)	0.82
≥ 6 cm	4 (44.4)	3 (37.5)	
Crossing midline, n (%)	1 (11.1)	1 (12.5)	1.0
Contrast-enhancing, n (%)	0 (0)	0 (0)	1.0
Extent of resection, n (%)			
≥ 90%	6 (66.7)	5 (62.5)	0.91

< 90%	2 (22.2)	2 (25.0)	
Biopsy	1 (11.1)	1 (12.5)	
WHO grade, n (%)			0.91
Grade II: Diffuse astrocytoma	6 (66.7)	5 (62.5)	
Grade III: Anaplastic astrocytoma	3 (33.3)	3 (37.5)	
MIB-1 index, %			
Median (range)	3.8 (1.5–15)	4.0 (1.0–12)	0.92