



# Intersexual flower differences in an andromonoecious species: small pollen-rich staminate flowers under resource limitation

Murakami, Kana  
Katsuhara, Koki R.  
Ushimaru, Atushi

---

(Citation)

Plant Biology, 24(2):259-265

(Issue Date)

2022-03

(Resource Type)

journal article

(Version)

Accepted Manuscript

(Rights)

This is the peer reviewed version of the following article: [Murakami, K., Katsuhara, K.R. and Ushimaru, A. (2022), Intersexual flower differences in an andromonoecious species: small pollen-rich staminate flowers under resource limitation. Plant Biol J, 24: 259-265.], which has been published in final form at...

(URL)

<https://hdl.handle.net/20.500.14094/90008984>



1  
2 **Intersexual flower differences in an andromonoecious species: small**  
3 **pollen-rich staminate flowers under resource limitation**  
4

5 Kana Murakami, Koki R. Katsuhara<sup>\*†</sup> and Atushi Ushimaru

6 Graduate School of Human Development and Environment, Kobe University, 3-11

7 Tsurukabuto, Kobe 657-8501, Japan;

8 <sup>†</sup>Present address: Graduate School of Environmental and Life Science, Okayama

9 University, Okayama, Japan.

10 <sup>\*</sup> Corresponding author: k.katsuhara0228@gmail.com; +81-086-251-8870

11  
12 K.M. and K.R.K. contributed equally to this study  
13

14 **Running head:** Intersexual differences in andromonoecious *Commelina communis*  
15

16 **Key-words:** *Commelina*, floral allometry, flower–pollinator matching, pollen–pistil  
17 interference, pollinator attraction, resource allocation, sexual dimorphism, stabilizing  
18 selection  
19  
20

## Abstract

- Andromonoecy, the presence of perfect and staminate flowers in the same individual, has evolved repeatedly in angiosperms. The staminate flowers are generally smaller than the perfect flowers in species that produce staminate flowers plastically when resources are limited. The smaller staminate flowers are expected to be less attractive to pollinators and have reduced size-matching with pollinators than perfect flowers. We hypothesized that these potential disadvantages of staminate versus perfect flowers facilitate the evolution of sex-specific floral morphology, such as allometric relationship between flower size and male reproductive organ, in andromonoecious species.
- We compared six floral morphology traits, pollen production, pollinator visits, and pollen removal from anthers between staminate and perfect flowers in several natural *Commelina communis* populations. Nectarless and zygomorphic *C. communis* flowers had polymorphic stamens with attracting, feeding, and pollinating anthers and were visited by diverse pollinators.
- Staminate flowers were significantly smaller than perfect flowers, despite a large overlap in size between sexes. The lengths of pollinating stamens did not differ between staminate and perfect flowers, and staminate flowers produced significantly more pollen. We observed significantly more pollinator visits to perfect flowers than to staminate flowers. By contrast, pollen removal from pollinating stamens was significantly higher in staminate flowers than in perfect flowers.
- There is sexual dimorphism in flower morphology in *C. communis*. Staminate flowers with smaller attraction organs, similar pollinating stamens, and higher

45 pollen production assured higher pollen donor success relative to perfect flowers.

46 Our results suggest that the morphological changes in staminate flowers enhance

47 pollination success, even with limited resources.

48

49

## INTRODUCTION

The diversity and evolution of sexual systems in flowering plants are central topics of evolutionary biology (Darwin, 1877; Barrett, 2002a). The diverse plant sexual systems involve various combinations of perfect and unisexual (staminate and pistillate) flowers (Bawa and Beach, 1981; Dellaporta and Calderon-Urrea, 1993). Approximately 70% of angiosperm species have only perfect flowers, and the rest have multiple sexual flower types within and among individuals (Yampolsky and Yampolsky, 1922; Richards, 1997; Torices et al., 2011). Various selection pressures have been proposed to explain the evolution and maintenance of unisexual flowers, such as those optimizing resource allocation to male and female function, facilitating outcrossing, or increasing display size, although their selective roles have still not been explored fully in many species with different sexual systems (Barrett, 2002a; Barrett and Hough, 2013; Goldberg et al., 2017).

Andromonoecy, the production of both perfect and staminate flowers on the same individual, is thought to have evolved repeatedly and occurs in approximately 4,000 angiosperm species (ca. 2%) across several families (Yampolsky and Yampolsky, 1922; Bawa and Beach, 1981; Richards, 1997). Three mutually non-exclusive hypotheses explain the conditions that favor staminate flower production in andromonoecious plants. The resource reallocation hypothesis predicts that the production of non-fruiting staminate flowers can save resources, which are reallocated to other fitness-enhancing traits (Bertin, 1982; Spalik, 1991; Emms, 1993; Vallejo-Marín and Rausher, 2007a). The increased pollen donation hypothesis emphasizes the superiority of staminate flowers as pollen donors compared to perfect flowers because staminate flowers can produce more pollen grains and avoid pollen–pistil interference

(Solomon, 1987; Elle and Meagher, 2000; Schlessman et al., 2004; Dai and Galloway, 2012). Finally, the pollinator attraction hypothesis holds that staminate flower production increases pollinator attraction by giving rise to a larger display in a given individual under resource limitation and can enhance female reproductive success (Podolsky, 1992; Vallejo-Marín and Rausher, 2007b). In most cases, staminate flower production in andromonoecious plants is regarded as a plastic response to resource limitation or stress, as a staminate flower is less costly than a perfect flower because it lacks ovules, large pistils, and fruit production (Solomon, 1985; Diggle, 1993; Granado et al., 2017). This hypothetical relationship between staminate flower production and resource limitation is supported by the fact that pistils and other floral organs, such as the petals and sepals of staminate flowers, are typically smaller and lighter than those of perfect flowers in andromonoecious species (*e.g.*, Primack and Lloyd, 1980; Schlessman, 1982; Solomon, 1986; Emms, 1993; Cuevas and Polito, 2004; Narbona et al., 2005; Vallejo-Marín and Rausher, 2007a; Zhang and Tan, 2009; but see Huang, 2003).

Smaller staminate flowers may have some disadvantages in terms of pollination success compared to perfect flowers in zoophilous andromonoecious plants. First, staminate flowers with smaller petals (or sepals) can be less attractive to pollinators and may be visited less often (Galen and Newport, 1987; Conner and Rush, 1996; Blarer et al., 2002), although pollinators do not discriminate between perfect and staminate flowers in some andromonoecious species (*Aesculus pavia*, Bertin, 1982; *Solanum carolinense*, Vallejo-Marín and Rausher, 2007a; *Passiflora incarnata*, Dai and Galloway, 2012). Second, the lower investment in stamens (filaments) may change the position of anthers in a staminate flower relative to that in a perfect flower and reduce

physical matching between the anthers and pollinators, thus decreasing efficient pollen transfer to the stigmas of perfect flowers (Harder, 1990; Tong and Huang, 2018). Note that the within-flower anther position, which is generally determined by filament length, is thought to be consistent across individuals within a population under strong pollinator-mediated stabilizing selection (Cresswell, 1998; Conner et al., 2009; Ushimaru et al., 2003a; Opedal, 2019). These potential disadvantages of staminate versus perfect flowers would facilitate the evolution of sex-specific floral morphology (*i.e.*, sexual dimorphism in floral morphology) in staminate flowers of andromonoecious species, although the idea has not been tested.

In this study, we compared the sexual differences in floral traits, pollinator visits, and pollen removal success between staminate and perfect flowers in the annual andromonoecious herb *Commelina communis*. The bilaterally symmetrical (zygomorphic) flowers of *C. communis* are self-compatible but still attract diverse pollinators such as syrphid flies and bumble and honey bees with their showy blue petals and yellow stamens (Ushimaru and Hyodo, 2005; Ushimaru et al., 2007). In zygomorphic species, physical matching between flowers and pollinators is likely important for efficient pollen transfer, as it increases the precision of pollen placement (Neal et al., 1998; Wolfe and Krstolic, 1999; Ushimaru and Hyodo, 2005; Cosacov et al., 2014; Nikkeshi et al., 2015). First, we confirmed that staminate flowers were significantly smaller than perfect flowers in *C. communis*, because these flowers look very similar in size and there is no reported size difference between them (Fig. 1; Morita and Nigorikawa, 1999). Then, we addressed the following questions. Do staminate flowers receive fewer pollinator visits than perfect flowers? Is there sexual dimorphism in floral traits other than size between the two flower types? Can floral traits in

staminate flowers compensate for the disadvantage in the pollination process caused by resource limitation? Finally, we discuss the selective forces acting on the morphology of staminate flowers and the maintenance of andromonoecy in flowering plants.

## **MATERIALS AND METHODS**

**Study species**—*Commelina communis* L. (Commelinaceae) grows near rice fields and roadsides in temperate Northeast Asia. One plant has many inflorescences, each usually bearing one to five flowers, with each flower opening from sunrise to noon. The first flower in each inflorescence is usually perfect and those opening later can be staminate depending on conditions (Morita and Nigorikawa, 1999). Fruit production (or fertilization success) by early perfect flower(s) within a given inflorescence makes later flowers staminate, suggesting the resource conditions of each inflorescence influence the sex expression of later flowers (Morita and Nigorikawa, 1999; Katsuhara et al., in prep.). Both flower types are zygomorphic with two showy blue petals and have two long, one medium, and three short stamens, which have different functions. The anthers of long and medium stamens (L- and M-anthers, respectively) produce fertile pollen, which mainly contributes to pollination and serves as a reward for pollinators, respectively (Ushimaru et al., 2007). The anthers of short stamens (S-anthers) produce some sterile pollen and function as advertising for pollinators (Ushimaru et al., 2007). The long stamens provide a landing site for pollinators and the medium and short stamens function as floral guides (Ushimaru et al., 2007). Perfect flowers have four fertile ovules and a style of similar length to that of the long stamens, whereas staminate flowers have atrophied ovules.



Diverse insect groups including several syrphid flies and social and solitary bees frequently visit and forage or collect pollen from flowers of *C. communis*, which can also reproduce through bud pollination and delayed self-pollination (Morita and Nigorikawa, 1999; Katsuhara and Ushimaru, 2019). The existence of staminate flowers and the relatively high pollen-to-ovule ratio (1000–3000) in perfect flowers suggest the importance of outcrossing in this species (Cruden, 1977; Morita and Nigorikawa, 1999; Ushimaru et al., 2014). In addition, anther polymorphism (heteranthery) likely promotes outcrossing by controlling pollinator orientation in *Commelina* flowers (Ushimaru et al., 2007; Solís-Montero and Vallejo-Marín, 2017).

**Floral morphology and pollen production**—To examine differences in floral organ sizes between staminate and perfect flowers, we measured floral traits in six natural populations in Hyogo and Osaka prefectures, Japan from 5 September to 1 October 2017 (Table S2). Each population was investigated for 1–3 days, during which we arbitrarily selected 4–11 perfect and staminate flowers to measure the following six floral traits using digital calipers: petal length and width with the petal between glass slides, S-anther length and width with the anther on a glass slide; and the filament lengths of medium (M-stamen) and long (L-stamen) stamens. The petals and S-anthers are visual attractants; their sizes are important for pollination success (Ushimaru et al., 2007). Filament length is important in M- and L-stamens because it influences flower–pollinator matching (Ushimaru et al., 2003a, 2007; Cosacov et al., 2014).

To quantify pollen production, we arbitrarily selected 8–9 newly opened, unvisited flowers of each flower type at 5:30–6:00 in four populations (Table S2) and collected the M- and L-anthers from each. Each anther type of each flower was stored

separately in 1.0 mL of 99% ethanol. We estimated the total number of pollen grains per anther type by counting the pollen grains in three 10.0- $\mu$ L droplets per sample under a microscope (Ushimaru et al., 2014).

***Pollinator visits and pollination success***—From 5 September to 1 October in 2017, we compared differences in pollinator visits and pollen removal from anthers with fertile pollen grains between perfect and staminate flowers over 1–3 days in each of seven natural populations (Table S2). In each population, we set three 1  $\times$  1-m<sup>2</sup> plots in which pollinator visits to 6–12 arbitrarily selected perfect and staminate flowers were observed for 1 h (three 20-min observations per flower) during 6:30–11:00 each observation day. The pollinators were divided into four groups: *Episyrphus balteatus*, *Bombus diversus diversus*, other bees (including small solitary bees and some honey bees), and small syrphid flies (cf. Ushimaru et al., 2014; Katsuhara and Ushimaru, 2019).

We arbitrarily selected 8–9 flowers for observation in each of the four populations to estimate pollen removal by pollinators (Table S2). The M- and L-anthers of each selected flower were collected and stored separately in 1.0 mL of 99% ethanol just before the flower closed (12:00), and the pollen grains remaining on the anthers were counted in the laboratory using the same pollen-counting method. Pollen removal from M- and L- anthers in each flower was estimated as the population mean total pollen production minus the number of pollen grains remaining on each flower for each anther type.

## ***Data analyses***

*Floral traits*—First, we conducted principal component analysis (PCA) using the six floral parameters (petal length and width, S-anther length and width, and M-stamen and L-stamen lengths) to summarize the floral morphology of each flower type. We compared the mean value of each floral trait separately between staminate and perfect flowers because the six floral traits have different functions and might be influenced differently by resource limitation. Then, we constructed a linear mixed model (LMM), including the first principal component axis (PC1, which was positively correlated with all floral measurements and regarded as an index of flower size; Table S3) as the response variable, flower type (perfect = 0, staminate = 1) and days after 1 September as the explanatory variables, and population identity as a random term to examine sexual differences in flower size. If staminate flowers are produced under resource limitation in *C. communis*, we expect that staminate flowers would be smaller overall than perfect flowers that bloomed the same day. Second, to compare each floral measurement and pollen production between perfect and staminate flowers, we constructed six LMMs, each of which included one of the six floral measurements as the response variable and population identity and observation date as random terms, and two LMMs that included the pollen number for the M- and L anthers as the response variable and population identity as a random term. All LMMs incorporated flower type as the explanatory variable. Finally, to compare the size variation in each trait between flower types, we calculated the coefficient of variation (CV) and conducted an F-test for each measurement. We compared the size variation of each floral trait separately between staminate and perfect flowers because resource limitation might influence the variation in the six traits differently. We expect that organs related to pollinator attraction would

be more variable than those associated with flower–pollinator matching in staminate flowers, which are often produced under different levels of resource limitation (Ushimaru and Nakata 2001; Ushimaru et al. 2003a).

*Pollination success*—To compare pollinator visitation frequency between perfect and staminate flowers, we constructed a generalized LMM (Poisson error and log link) that incorporated the total number of visits per hour per flower as the response variable, flower type as the explanatory variable, and population identity and observation date as independent random terms. We also constructed LMMs that included pollen removal from M- or L- anthers and flower type as the response and explanatory variables, respectively, with population identity and observation date as random terms.

We examined the significance of each explanatory variable in the models using the Wald test. We performed all analyses using the glmmADMB package in R software (ver. 4.0.2; R Core Team, 2020; Fournier et al., 2012).

## RESULTS

*Floral traits*—In the PCA with floral measurements, PC1 explained 77.9% of the total variance, whereas the second axis (PC2) explained 9.5% and was negatively correlated with all measurements, except for M- and L-stamen lengths (Table S3). The PCA scatterplot did not show a clear difference between perfect and staminate flowers, possibly because floral organs other than the pistils appear not to differ in size or morphology between the sexes *prima facie* (Figs. 1 and 2). However, the LMM analysis revealed that the PC1 loadings were significantly lower in staminate flowers than in

perfect flowers when we considered a significant decrease in the variable with observation days in the analysis (Fig. 2, Table S4).

The LMM analyses of floral traits revealed that five measurements (petal length and width, S-anther length and width, and M-stamen length) were significantly smaller in staminate versus perfect flowers (Fig. 3, Table S4). The L-stamen length did not differ between staminate and perfect flowers (Fig. 3, Table S4). By contrast, staminate flowers produced significantly more pollen on both M- and L-anthers than did perfect flowers (Fig. 3, Table S4). Note that our preliminary experiment revealed that pollen ability (pollen fertility and pollen tube growth) did not differ between perfect and staminate flowers (Table S1).

Compared to perfect flowers, staminate flowers had larger CVs for the six floral measurements and smaller CVs for pollen production on M- and L-anthers (Table 1), although significant differences were only found for petal and S-anther lengths (Table 1). The CV of L-stamen length was the smallest among all traits in staminate flowers, and the value was comparable with that of perfect flowers.

**Pollination success**—We observed 90 pollinator visits to 118 perfect flowers and 48 to 108 staminate flowers (44 and 14 *Episyrphus balteatus*, 3 and 1 *Bombus diversus*, 14 and 11 other bee, and 29 and 22 other small syrphid fly visits, respectively). The pollinator composition did not differ between flower types (Fisher's exact test,  $p = 0.11$ ). Pollinator visits per flower per hour was significantly less frequent for staminate flowers (mean 0.76 for perfect flowers and 0.44 for staminate flowers; Fig. 4, Table S4). Pollen removal from the L-anthers was significantly higher in staminate flowers,

whereas pollen removal from the M-anthers did not differ significantly between flower types (Fig. 4, Table S4).

## DISCUSSION

Our results demonstrated that although there was a large overlap between flower types, staminate flowers were significantly smaller than perfect flowers upon comparison on the same observation dates at the same site. This suggests that staminate flowers were under resource limitation. Staminate flowers were significantly more pollinator limited compared to perfect flowers, likely because of their reduced attractiveness due to the relatively smaller petals (Fig. S1). By contrast, the lengths of pollinating L-stamens in staminate flowers were comparable to those in perfect flowers and were the most stable trait. Therefore, the L-stamen length is under strong stabilizing selection to enable precise pollen placement on pollinator bodies corresponding to the stigma position of perfect flowers (Ushimaru et al., 2003a,b). This intersexual difference in the allometric relationship between flower size and L-stamen length may ensure pollen donor success in staminate flowers under resource limitation and likely contributes to the maintenance of andromonoecy in *C. communis*.

We found that pollen removal from the L-anthers was higher in staminate flowers than in perfect flowers, although the staminate flowers received fewer pollinator visits (ca. 60% of the visits to perfect flowers). The first possible explanation for this discrepancy is that reduced pollen–pistil interference promotes pollen removal per visit in staminate flowers (Lloyd and Webb, 1986; Fetscher, 2001; Barrett, 2002b), as reported for andromonoecious *Solanum carolinense* (Elle and Meagher, 2000). However, further examination of this is required because pollen–pistil interference was

not found in another study of *S. carolinense* (Vallejo-Marín and Rausher, 2007a). As a second possibility, increased pollen foraging from L-anthers by syrphid flies could have resulted in greater pollen removal from staminate flowers (Ushimaru and Hyodo, 2005; Ushimaru et al., 2007). However, this is unlikely because the relative abundance of syrphid flies and their foraging behavior did not differ between flower types. A future study of pistil-removed perfect flowers and detailed observations of pollinator behaviors are needed to elucidate the advantages/disadvantages of staminate flowers in terms of pollination success.

The lowest size variation in L-stamens among all the measurements of staminate flowers is consistent with a study of eight natural *C. communis* populations that demonstrated that this trait was conserved within and among populations (Ushimaru et al., 2003a). By contrast, the attraction-related organs (petals and S-anthers) of staminate flowers exhibited significantly larger size variation compared to perfect flowers. These results suggest that the size of attraction-related organs is flexible and likely responds plastically to resource conditions. These findings support the hypothesis that pollinators impose stronger stabilizing selection on traits related to flower–pollinator matching than on those involved in attraction (Cresswell, 1998; Worley et al., 2000; Ushimaru et al., 2003a). Staminate flowers may preferentially allocate available resources to pollination-related organs and then invest the surplus in attraction-related traits under resource-limited conditions. Elucidating the developmental process of staminate flowers in *C. communis* and other andromonoecious species will improve our understanding of the evolution and maintenance of andromonoecy in flowering plants.

We found that the M- and L-anthers of staminate flowers produced slightly, but significantly, more pollen grains than those of perfect flowers (ca. 1.17- and 1.27-fold

309 compared to perfect flowers, respectively), and pollen fertility did not differ between  
310 flower types (Table S1). The findings support the increased pollen donation hypothesis.  
311 Patterns of sexual dimorphism in pollen production vary among andromonoecious  
312 species—compared to perfect flowers, staminate flowers of *Sagittaria*  
313 *guyanensis* subsp. *lappula* produce more pollen (Huang et al., 2000), those of *Solanum*  
314 *carolinense* (Solomon, 1986) and *Olea europaea* produce similar amounts of pollen  
315 (Cuevas and Polito, 2004), and those of *Anthriscus sylvestris* produce less pollen  
316 (Spalik and Woodell, 1994). There was no common trend (Vallejo-Marín and Rausher,  
317 2007a). In *C. communis*, increased pollen production in staminate flowers likely  
318 compensates for fewer pollinator visits compared to perfect flowers to assure pollen  
319 donation success. Although we compared only pollen removal between flower types, it  
320 is necessary to examine the relationship between paternity success and pollen  
321 production with molecular markers in andromonoecious species exhibiting sexual  
322 dimorphism in pollen production.

323         In conclusion, we found cryptic sexual dimorphism in flower morphology in  
324 andromonoecious *C. communis*. The results from pollinator observations and the  
325 assessment of pollen removal success suggest that staminate flowers with smaller  
326 attraction organs compared to perfect flowers have similar pollinating stamens and  
327 higher rates of pollen production to ensure higher chances of pollen donor success.  
328 Because this is a case study of a single species, more andromonoecious species should  
329 be examined to generalize our findings.

330



## **ACKNOWLEDGMENTS**

We thank A. Nikkeshi and N. Ohmido for their advice on the pollen counting and pollen tube observation methods, respectively. We also thank the members of the Biodiversity, Evolutionary Ecology, and Minamoto Laboratories of Kobe University for their valuable comments on our study. We are grateful to the land owners who allowed our field surveys to be carried out on their properties. This work was supported by Grants-in-Aid for Scientific Research (nos. 16K07517, 17J01902, 19H03303, 19K06855, and 20J01271) from the Japan Society for the Promotion of Science.

## **AUTHOR CONTRIBUTIONS**

All authors conceived the ideas and designed the methodology;  
KM collected and analyzed the data;  
KM and KRK led the writing of the manuscript;  
All authors contributed critically to the drafts and gave final approval for publication.

## **DATA AVAILABILITY**

We are going to archive our data in the Dryad Digital Repository.

## **SUPPORTING INFORMATION**

Additional supporting information may be found online in the Supporting Information section at the end of the article.

353   **REFERENCE**

- 354   Barrett S.C.H. (2002a) The evolution of plant sexual diversity. *Nature Reviews Genetics*  
355    **3**:274–284.
- 356   Barrett S.C.H. (2002b) Sexual interference of the floral kind. *Heredity* **88**:154–159.
- 357   Barrett S.C.H., Hough J. (2013) Sexual dimorphism in flowering plants. *Journal of*  
358    *Experimental Botany* **64**:67–82.
- 359   Bawa K.S., Beach J.H. (1981) Evolution of Sexual Systems in Flowering Plants. *Annals*  
360    *of the Missouri Botanical Garden* **68**:254–274.
- 361   Bertin R.I. (1982) The ecology of sex expression in red buckeye. *Ecology* **63**:445–456.
- 362   Blarer A., Keasar T., Shmida A. (2002) Possible mechanisms for the formation of  
363    flower size preferences by foraging bumblebees. *Ethology* **108**:341–351.
- 364   Conner J.K., Rush S. (1996) Effects of flower size and number on pollinator visitation  
365    to wild radish, *Raphanus raphanistrum*. *Oecologia* **105**:509–516.
- 366   Conner J.K., Sahli H.F., Karoly K. (2009) Tests of adaptation: functional studies of  
367    pollen removal and estimates of natural selection on anther position in wild radish.  
368    *Annals of Botany* **103**:1547–1556.
- 369   Cosacov A., Cocucci A.A., Sérsic A.N. (2014) Geographical differentiation in floral  
370    traits across the distribution range of the Patagonian oil-secreting *Calceolaria*  
371    *polyrhiza*: do pollinators matter? *Annals of Botany* **113**:251–266.
- 372   Cresswell J.E. (1998) Stabilizing selection and the structural variability of flowers  
373    within species. *Annals of Botany* **81**:463–473.
- 374   Cruden R.W. (1977) Pollen-ovule ratios: a conservative indicator of breeding systems in  
375    flowering plants. *Evolution* **31**:32–46.

376 Cuevas J., Polito V.S. (2004) The role of staminate flowers in the breeding system of  
 377 *Olea europaea* (Oleaceae): an andromonoecious, wind-pollinated taxon. *Annals of*  
 378 *Botany* **93**:547–553.

379 Dai C., Galloway L.F. (2012) Male flowers are better fathers than hermaphroditic  
 380 flowers in andromonoecious *Passiflora incarnata*. *New Phytologist* **193**:787–796.

381 Darwin C. (1877) The different forms of flowers on plants of the same species. Reprint,  
 382 1986. University of Chicago Press, Chicago, IL

383 Dellaporta S.L., Calderon-Urrea A. (1993) Sex determination in flowering plants. *The*  
 384 *Plant Cell* **5**:1241–1251.

385 Diggle P.K. (1993) Developmental plasticity, genetic variation, and the evolution of  
 386 andromonoecy in *Solanum Hirtum* (solanaceae). *American Journal of Botany* **80**:967–  
 387 973.

388 Elle E., Meagher T.R. (2000) Sex allocation and reproductive success in the  
 389 andromonoecious perennial *Solanum carolinense* (Solanaceae). II. Paternity and  
 390 functional gender. *The American Naturalist* **156**: 622–636.

391 Emms S.K. (1993) Andromonoecy in *Zigadenus paniculatus* (Liliaceae): spatial and  
 392 temporal patterns of sex allocation. *American Journal of Botany* **80**:914–923.

393 Fetscher A.E. (2001) Resolution of male-female conflict in an hermaphroditic flower.  
 394 *Proceedings: Biological Sciences* **268**:525–529.

395 Fournier D.A., Skaug H.J., Ancheta J., Ianelli, J., Magnusson A., Maunder M., Nielsen  
 396 A., Sibert J. (2012) AD Model Builder: using automatic differentiation for statistical  
 397 inference of highly parameterized complex nonlinear models. *Optim. Methods Softw.*  
 398 **27**: 233–249.

399 Galen C., Newport M.E.A. (1987) Bumble bee behavior and selection on flower size in  
400 the sky pilot, *Polemonium viscosum*. *Oecologia* **74**:20–23.

401 Goldberg E.E., Otto S.P., Vamosi J.C., Mayrose I., Sabath N., Ming R., Ashman T.-L.  
402 (2017) Macroevo­lutionary synthesis of flowering plant sexual systems. *Evolution*  
403 **71**:898–912.

404 Gong Y.-B., Huang S.-Q. (2009) Floral symmetry: pollinator-mediated stabilizing  
405 selection on flower size in bilateral species. *Proceedings of the Royal Society B:*  
406 *Biological Sciences* **276**:4013–4020.

407 Granado-Yela C., Balaguer L., Cayuela L., Méndez M. (2017) Unusual positional  
408 effects on flower sex in an andromonoecious tree: Resource competition, architectural  
409 constraints, or inhibition by the apical flower? *American Journal of Botany* **104**:608–  
410 615.

411 Harder L.D. (1990) Pollen removal by bumble bees and its implications for pollen  
412 dispersal. *Ecology* **71**:1110–1125.

413 Huang S.-Q. (2003) Flower dimorphism and the maintenance of andromonoecy in  
414 *Sagittaria guyanensis* ssp. *lappula* (Alismataceae). *New Phytologist* **157**:357–364.

415 Huang S.Q., Song N., Wang Q., Tang L.L., Wang X.F. (2000) Sex expression and the  
416 evolutionary advantages of male flowers in an andromonoecious species, *Sagittaria*  
417 *guyanensis* subsp. *lappula* (Alismataceae). *Journal of Integrative Plant Biology* **42**:  
418 1108–1114.

419 Katsuhara K.R., Ushimaru A. (2019) Prior selfing can mitigate the negative effects of  
420 mutual reproductive interference between coexisting congeners. *Functional Ecology*  
421 **33**:1504–1513.

422 Lloyd D.G., Webb C. J. (1986) The avoidance of interference between the presentation  
 423 of pollen and stigmas in angiosperms. I. Dichogamy. *New Zealand Journal of Botany*  
 424 **24**: 135–162.

425 Morita T., Nigorikawa T. (1999) *Phenotypic plasticity of floral sex*. In M. Ohara (Eds.),  
 426 Natural history of flowers (pp. 227–242). Sapporo: Hokkaido University Press. [in  
 427 Japanese]

428 Narbona E., Ortiz P.L., Arista M. (2005) Dichogamy and sexual dimorphism in floral  
 429 traits in the andromonoecious *Euphorbia boetica*. *Annals of Botany* **95**:779–787.

430 Neal P.R., Dafni A., Giurfa M. (1998) Floral symmetry and its role in plant-pollinator  
 431 systems: terminology, distribution, and hypotheses. *Annual Review of Ecology and*  
 432 *Systematics* **29**:345–373.

433 Nikkeshi A., Kurimoto D., Ushimaru A. (2015) Low flower-size variation in bilaterally  
 434 symmetrical flowers: Support for the pollination precision hypothesis. *American*  
 435 *Journal of Botany* **102**:2032–2040.

436 Opedal Ø.H. (2019) The evolvability of animal-pollinated flowers: towards predicting  
 437 adaptation to novel pollinator communities. *New Phytologist* **221**:1128–1135.

438 Podolsky R.D. (1992) Strange floral attractors: pollinator attraction and the evolution of  
 439 plant sexual systems. *Science* **258**:791–793.

440 Primack R.B., Lloyd D.G. (1980) Andromonoecy in the New Zealand montane shrub  
 441 manuka, *Leptospermum Scoparium* (Myrtaceae). *American Journal of Botany* **67**:361–  
 442 368.

443 Richards A.J. (1997) *Plant Breeding Systems*. Chapman & Hall, London.

444 R Core Team. (2020) R: A language and environment for statistical computing. R  
 445 Foundation for Statistical Computing, Vienna, Austria. URL [https://www.R-](https://www.R-project.org/)  
 446 [project.org/](https://www.R-project.org/).  
 447 Schlessman M.A., Underwood N., Watkins T., Graceffa L.M., Cordray D. (2004)  
 448 Functions of staminate flowers in andromonoecious *Pseudocymopterus montanus*  
 449 (Apiaceae, Apioideae). *Plant Species Biology* **19**:1–12.  
 450 Solís-Montero L., Vallejo-Marín M. (2017) Does the morphological fit between flowers  
 451 and pollinators affect pollen deposition? An experimental test in a buzz-pollinated  
 452 species with anther dimorphism. *Ecology and Evolution* **7**:2706–2715.  
 453 Solomon B.P. (1985) Environmentally influenced changes in sex expression in an  
 454 Andromonoecious plant. *Ecology* **66**:1321–1332.  
 455 Solomon B.P. (1986) Sexual allocation and andromonoecy: resource investment in male  
 456 and hermaphrodite flowers of *Solanum carolinense* (Solanaceae). *American Journal of*  
 457 *Botany* **73**:1215–1221.  
 458 Solomon B.P. (1987) The role of male flowers in *Solanum carolinense*: pollen donors or  
 459 pollinator attractors? *Evolutionary Trends Plants* **1**:89–93.  
 460 Spalik K. (1991) On evolution of andromonoecy and ‘overproduction’ of flowers: a  
 461 resource allocation model. *Biological Journal of the Linnean Society* **42**:325–336.  
 462 Spalik K., Woodell S.R.J. (1994) Regulation of pollen production in *Anthriscus*  
 463 *sylvestris*, an Andromonoecious species. *International Journal of Plant Sciences*  
 464 **155**:750–754.  
 465 Tong Z.-Y., Huang S.-Q. (2018) Safe sites of pollen placement: a conflict of interest  
 466 between plants and bees? *Oecologia* **186**:163–171.

467 Torices R., Méndez M., Gómez J.M. (2011) Where do monomorphic sexual systems fit  
 468 in the evolution of dioecy? Insights from the largest family of angiosperms. *New*  
 469 *Phytologist* **190**:234–248.

470 Ushimaru A., Hyodo F. (2005) Why do bilaterally symmetrical flowers orient  
 471 vertically? Flower orientation influences pollinator landing behaviour. *Evolutionary*  
 472 *Ecology Research* **7**: 151–160.

473 Ushimaru A., Itagaki T., Ishii H.S. (2003a) Variation in floral organ size depends on  
 474 function: a test with *Commelina communis*, an andromonoecious species. *Evolutionary*  
 475 *Ecology Research* **5**:615–622.

476 Ushimaru A., Itagaki T., Ishii H.S. (2003b) Floral correlations in an andromonoecious  
 477 *Commelina communis*. *Plant Species Biology* **18**: 103–100.

478 Ushimaru A., Kikuchi S., Yonekura R., Maruyama A., Yanagisawa N., Kagami M.,  
 479 Nakagawa M., Mahoro S., Kohmatsu Y., Hatada A., Kitamura S., Nakata K. (2004)  
 480 The influence of floral symmetry and pollination systems on flower size variation.  
 481 *Nordic Journal of Botany* **24**:593–598.

482 Ushimaru A., Kobayashi A., Dohzono I. (2014) Does urbanization promote floral  
 483 diversification? Implications from changes in herkogamy with pollinator availability in  
 484 an urban-rural area. *The American Naturalist* **184**:258–267.

485 Ushimaru A., Nakata K. (2001) Evolution of flower allometry and its significance on  
 486 pollination success in deceptive orchid, *Pogonia japonica*. *Internatinal Journal of*  
 487 *Plant Sciences* **162**:1307–1311.

488 Ushimaru A., Watanabe T., Nakata K. (2007) Colored floral organs influence pollinator  
 489 behavior and pollen transfer in *Commelina communis* (Commelinaceae). *American*  
 490 *Journal of Botany* **94**:249–258.

491 Vallejo-Marín M., Rausher M.D. (2007a) The role of male flowers in andromonoecious  
492 species: energetic costs and siring success in *Solanum Carolinense* L. *Evolution*  
493 **61**:404–412.

494 Vallejo-Marín M., Rausher M.D. (2007b) Selection through female fitness helps to  
495 explain the maintenance of male flowers. *The American Naturalist* **169**:563–568.

496 Wolfe L.M., Krstolic J.L., Ritland A.E.K. (1999) Floral symmetry and its influence on  
497 variance in flower size. *The American Naturalist* **154**:484–488.

498 Worley A.C., Baker A.M., Thompson J.D., Barrett S.C.H. (2000) Floral display in  
499 narcissus: variation in flower size and number at the species, population, and  
500 individual levels. *International Journal of Plant Sciences* **161**:69–79.

501 Yampolsky C., Yampolsky H. (1922) Distribution of sex forms in phanerogamic flora.  
502 *Bibliotheca Genetica* **3**:1–62.

503 Zhang T., Tan D.-Y. (2009) An examination of the function of male flowers in an  
504 andromonoecious shrub *Capparis spinosa*. *Journal of Integrative Plant Biology* **51**:316–  
505 324.

506



## Figure legends

**Fig. 1.** Perfect (left) and staminate (right) flowers of *Commelina communis*. Both were the second-opening flowers of each inflorescence on the same individual. The photograph was taken in the middle of the flowering season.

**Fig. 2.** Scatterplot of a principal component analysis of floral traits (a), and the relationship between principal component 1 (PC1) and the observation date for each flower type (b). PC1 explained 77.9% of the variation, was positively correlated with all measurements, and was taken to represent flower size (Table S3). Red circles, perfect flowers; blue circles, staminate flowers. Red and blue lines in (b) indicate the estimated mean values for perfect and staminate flowers, respectively, based on the results from generalized linear mixed model analysis. Boxplots in (b) indicate PC1 values for perfect (P) and staminate (S) flowers, and asterisks indicate a significant difference (\*\*\*)  $p < 0.001$ ).

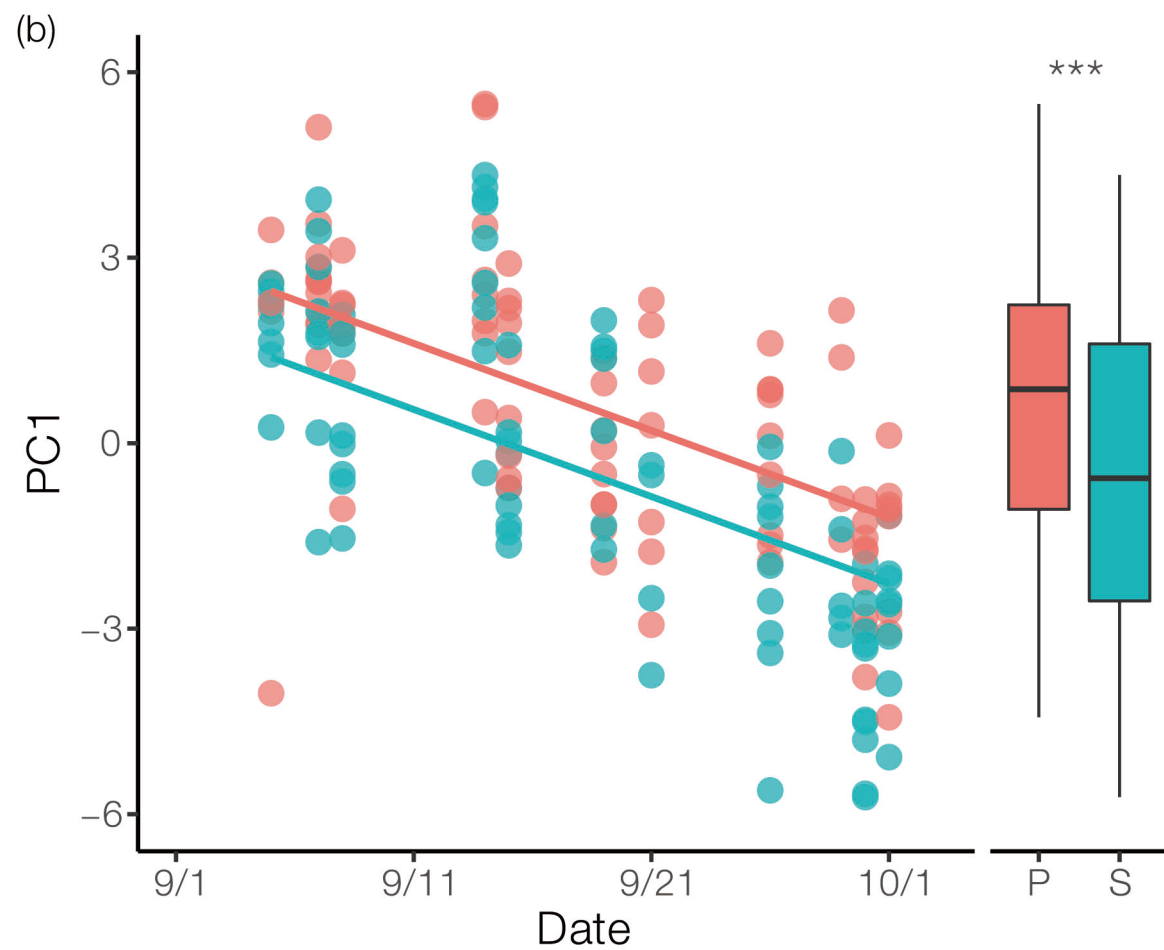
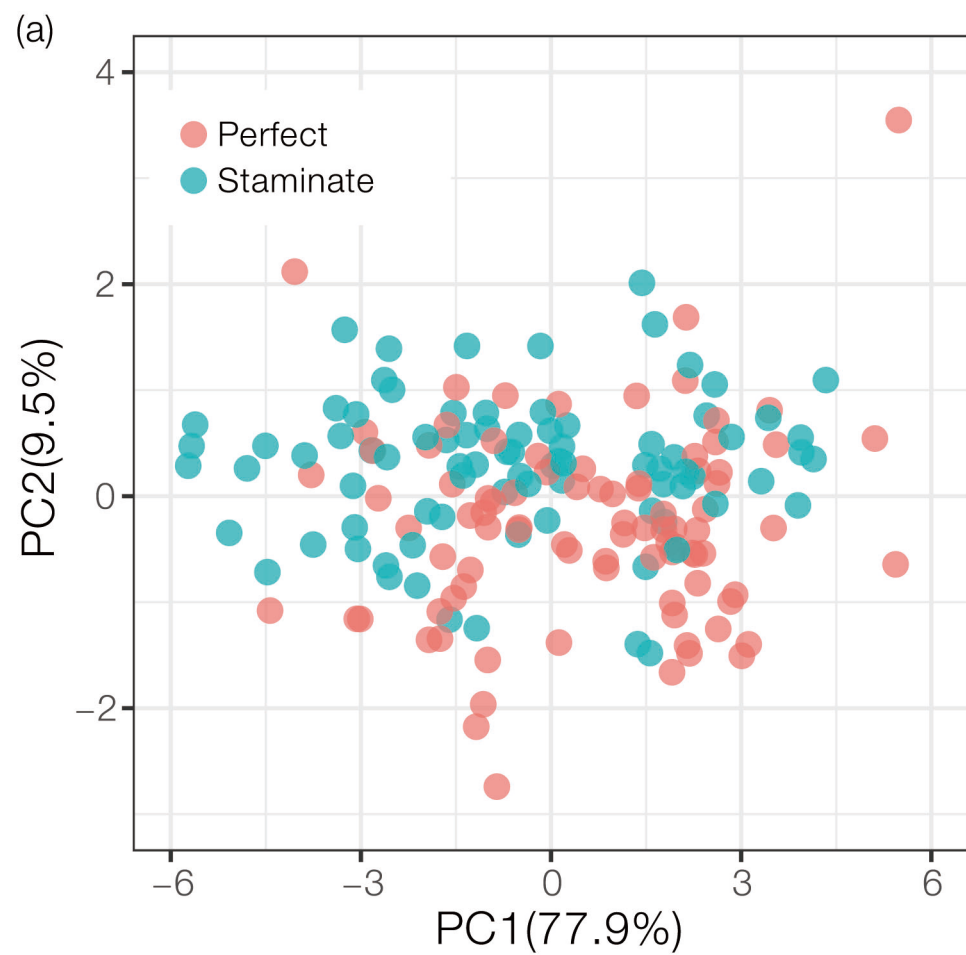
**Fig. 3.** Boxplots of floral traits of perfect (P) and staminate (S) flowers: petal length (a) and width (b), S-anther length (c) and width (d), M-stamen (e) and L-stamen (f) lengths, and the number of pollen grains on an M-anther (g) and L-anther (h). The Wald test result for each comparison is indicated as follows: \*\*\*  $p < 0.001$ ; \*\*  $p < 0.01$ ; \*  $p < 0.05$ ; ns, not significant.

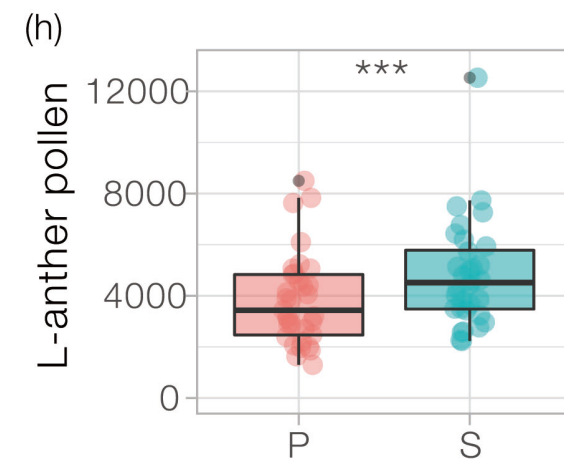
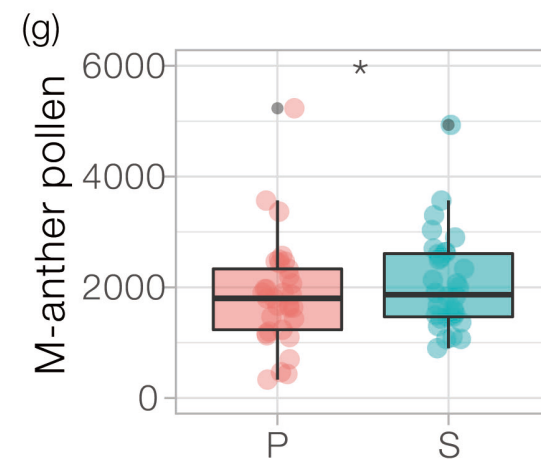
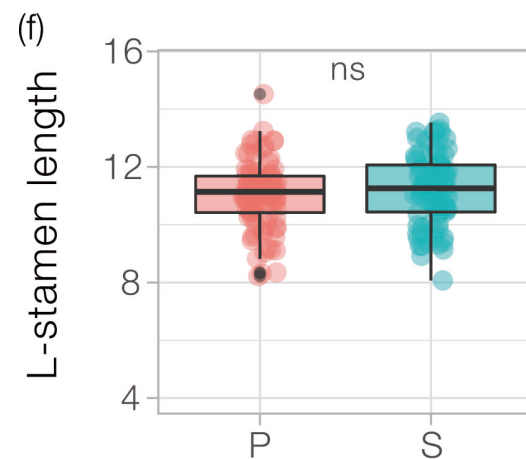
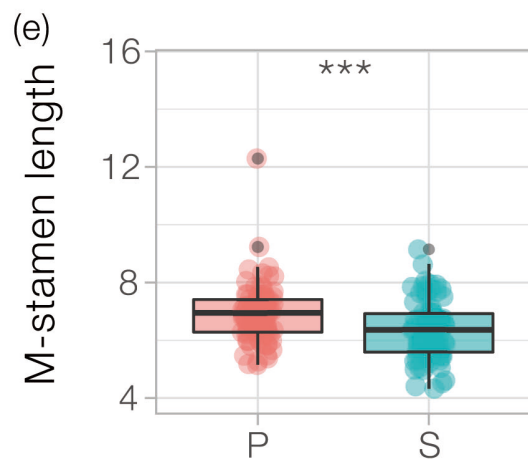
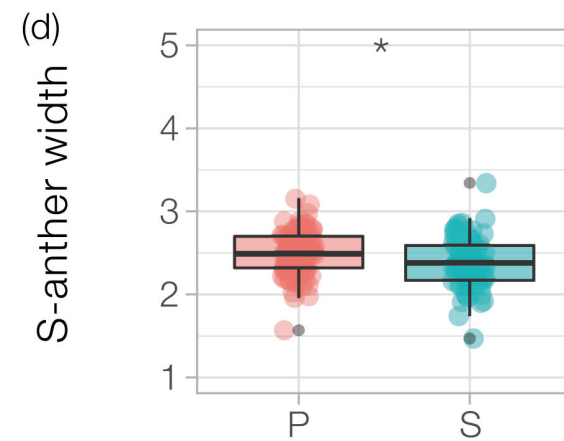
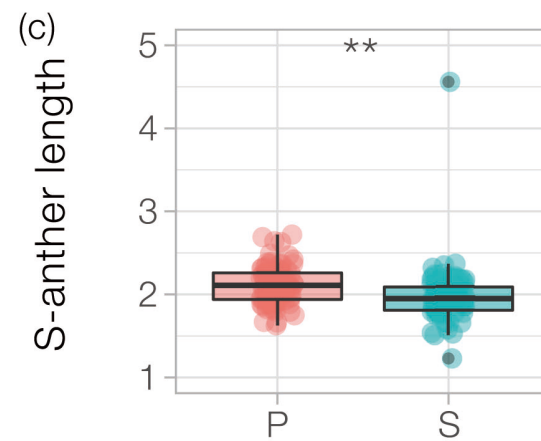
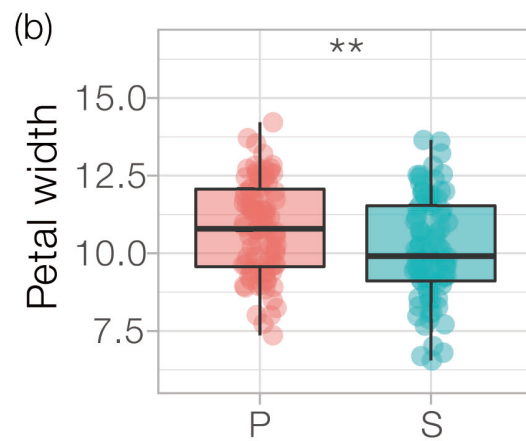
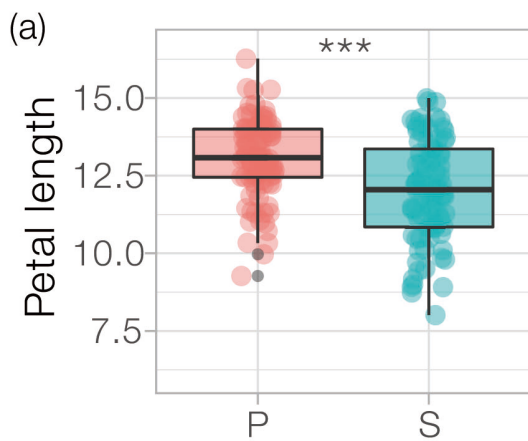
**Fig. 4.** Pollinator visitation frequency (a) and pollen removal from M-anthers (b) and L-anthers (c) in perfect (P) and staminate (S) flowers. The Wald test result for each comparison is indicated as follows: \*\*\*  $p < 0.001$ ; \*\*  $p < 0.01$ ; ns, not significant. Pollinator visitation data are presented as the means and standard errors (bars) in (a), and the medians are indicated with bold lines in the boxplots in (b) and (c).

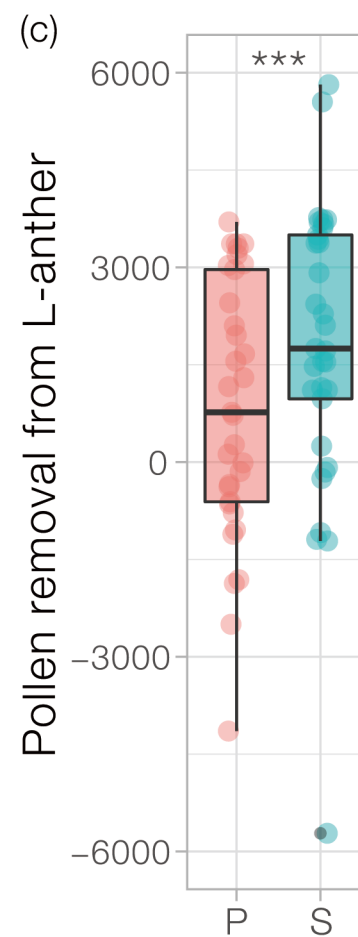
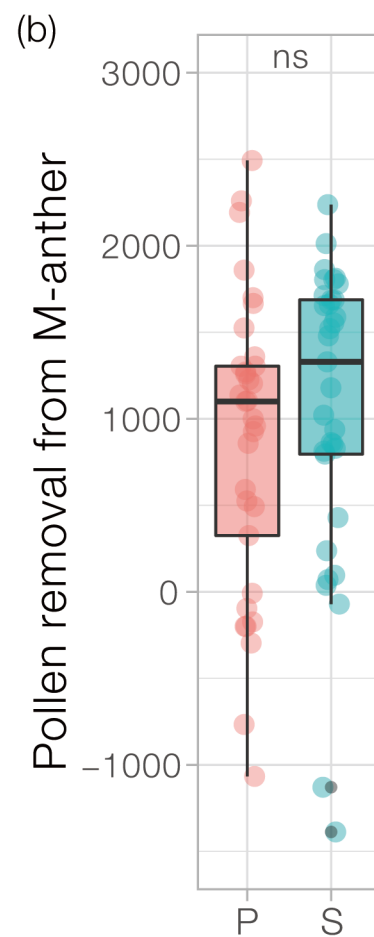
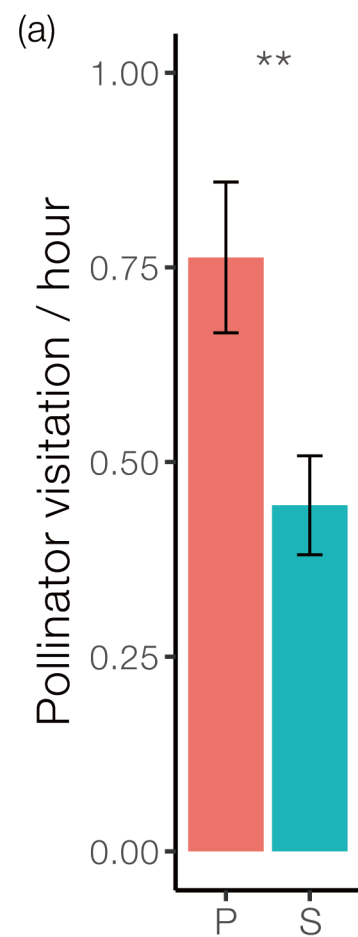
531 **Table 1.** Coefficients of variation (CVs) and variation test (F-test) results for each floral  
 532 trait. P, perfect flowers; S, staminate flowers; S-, M-, L-anther, anther on a short,  
 533 medium, or long stamen, respectively. Boldface indicates significant effects ( $p < 0.05$ ).

Trait	CV (%)		Variation test	
	P	S	F value	<i>p</i>
Petal length	10.0	13.8	1.93	<b>0.001</b>
Petal width	14.1	16.3	1.33	0.09
S-anther length	10.8	17.6	2.66	<b>&lt; 0.001</b>
S-anther width	10.6	12.4	1.38	0.070
M-stamen length	14.3	15.4	1.17	0.235
L-stamen length	10.6	10.9	1.07	0.383
No. of M-anther pollen	51.4	41.8	1.51	0.127
No. of L-anther pollen	46.7	43.5	1.15	0.348









**Table S1.** Pollen fertility and pollen tube growth in perfect (P) and staminate (S)  
flowers.

		P	S
Pollen fertility			
	Proportion of mature fruit	88% (21/24)	70% (19/27)
Pollen tube growth			
	Proportion of styles with pollen growth to the ovary (3 h after pollination)	39% (7/18)	58% (11/19)
	Proportion of styles with pollen growth to the ovary (6 h after pollination)	93% (14/15)	100% (14/14)

**Method:** In 2017, we performed two types of hand-pollination to examine differences in pollen fertility and pollen tube growth between perfect and staminate flowers in *Commelina communis*. To examine pollen fertility, we hand-pollinated the stigmas of perfect flowers with pollen grains collected from perfect and staminate flowers and examined the fruit set after anthesis in a population near Ogo01 (see Table S2). The petals of experimentally pollinated perfect flowers were removed to eliminate pollinator visitation. We conducted the experiment over 3 days and obtained fruit set data from 24 and 27 flowers pollinated using pollen from perfect and staminate flowers, respectively.

To examine pollen tube growth, we hand-pollinated petal-removed perfect flowers with pollen from each flower type in a similar way and collected and stored their pistils in 99% ethanol for 3 or 6 h after pollen application. Then, we rehydrated the samples with 0.5 N sodium hydroxide solution for 15 min at 60°C and stained them with 0.1% aniline blue solution for 24 h. After the treatment, we put samples on a glass slide, crushed them with a glass cover, and observed pollen tube growth to the ovary within the style via fluorescence microscopy. These experiments were conducted over

1–3 days at Ogo01 and in a population close to Ogo01 (Table S2). We observed 33  
pollinated pistils of each flower type (perfect versus staminate).

For pollen fertility, we constructed a generalized linear mixed model (GLMM;  
binomial error and logit link), including fruit set, pollen source (perfect/staminate), and  
observation date as the response variable, explanatory variable, and random term,  
respectively. For pollen tube growth, we constructed a GLMM (binomial error and logit  
link) in which the proportion of styles with pollen growth to the ovary was the response  
variable, the source of pollen (perfect/staminate) and time after pollen application were  
the explanatory variables, and observation date and population identity were random  
terms. We examined the significance of each explanatory variable in the models using  
the Wald test. We performed all analyses using R, with the glmmADMB package.

**Results:** According to the GLMM analyses, pollen source had no significant effect on  
fruit set ( $p = 0.11$ ). There were no significant differences in the proportions of styles  
with pollen growth to the ovary between the sources of pollen ( $p = 0.12$ ), whereas the  
proportion increased significantly with time after pollen application ( $p = 0.006$ ).



34 **Table S2.** Characteristics of the sampled populations: population identity, location, sample size for each observation, and pollen production  
35 on M- and L-anthers.

Prefecture			Pollinator observations	Pollinator observations	Pollen count after	Froral trait	Floral trait measurements	Pollen count before	Pollen on M-anthers	Pollen on L-anthers
Population	lat	long	(days)	(P/S flowers)	pollination	mesurements	(P/S flowers sampled)	pollination	before pollination	before pollination
			(days)	(P/S flowers)	(P/S flowers sampled)	(days spent)		(P/S flowers sampled)	(P/S flowers)	(P/S flowers)
Hyogo										
Yamada	34.7636	135.1302	2	20/19	8/8	2	18/16	8/8	1866.7 / 2237.5	3120.8 / 3679.2
Ogo01	34.8168	135.1708	1	12/8	na	1	12/8	na	na	na
Ogo02	34.8139	135.1918	2	18/18	na	2	13/16	na	na	na
Ogo03	34.8170	135.1940	1	10/10	9/9	1	9/9	8/8	1658.3 / 2229.2	3354.2 / 4708.3
Rokko	34.7372	135.2346	1	8/6	na	na	na	na	na	na
Osaka										
Turumi	34.7116	135.5799	2	21/19	8/8	2	14/16	9/8	2559.3 / 2145.8	5333.3 / 7050.0
Yodogawa	34.7312	135.5337	3	29/28	8/8	3	23/20	8/8	1304.2 / 1687.5	3362.5 / 3766.7

36 na, not applicable

37 **Table S3.** Results of principal component analysis (PCA). The values show the PC axis  
38 loadings obtained for each floral trait.

Trait	PC1 (77.9%)	PC2 (9.5%)
Petal length	0.59	-0.46
Petal width	0.62	-0.23
S-anther length	0.03	-0.07
S-anther width	0.05	-0.05
M-stamen length	0.32	0.41
L-stamen length	0.39	0.75

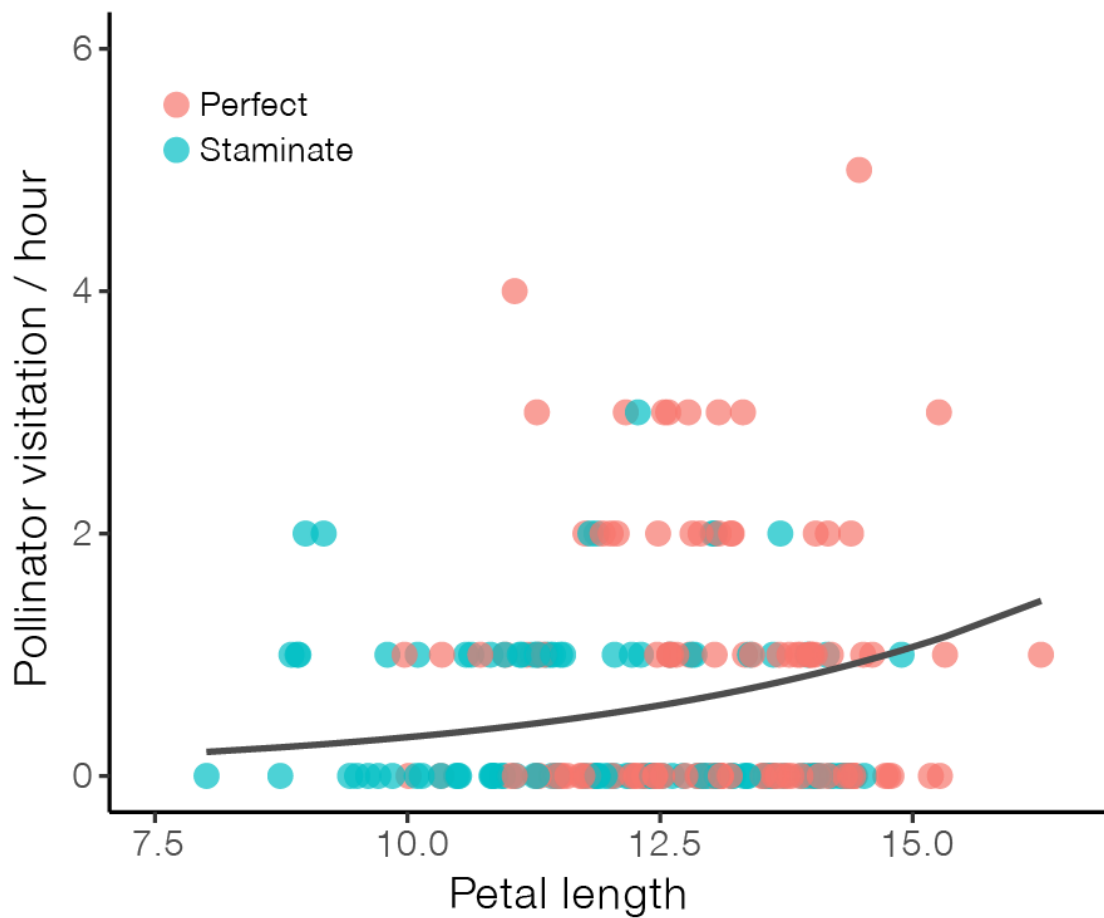
39

40

41 **Table S4.** Results of generalized linear mixed model analyses. The Wald test results are

Response variable / Explanatory variable	Estimated coefficient	Standard error	z-value	P	
<b>Flower size</b>					
PC1 (Flower size) /					
Intercept	3.02	0.527	5.73	< 0.001	***
Flower type - staminate	-0.14	0.025	-5.64	< 0.001	***
Days after September first	-1.07	0.244	-4.37	< 0.001	***
<b>Floral traits</b>					
Petal length /					
Intercept	12.98	0.416	31.18	< 0.001	***
Flower type - staminate	-0.99	0.175	-5.64	< 0.001	***
Petal width /					
Intercept	10.83	0.474	22.86	< 0.001	***
Flower type - staminate	-0.57	0.18	-3.17	0.002	**
S-anther length /					
Intercept	2.12	0.0484	43.78	< 0.001	***
Flower type - staminate	-0.13	0.0424	-3.07	0.002	**
S-anther width /					
Intercept	2.50	0.0478	52.22	< 0.001	***
Flower type - staminate	-0.10	0.0397	-2.55	0.011	*
M-stamen length /					
Intercept	6.93	0.271	25.6	< 0.001	***
Flower type - staminate	-0.57	0.113	-5	< 0.001	***
L-stamen length /					
Intercept	11.12	0.344	32.35	< 0.001	***
Flower type - staminate	0.20	0.134	1.52	0.13	
No. of M-anther pollen /					
Intercept	1858	164	11.29	< 0.001	***
Flower type - staminate	218	100	2.17	0.03	*
No. of L-anther pollen /					
Intercept	3783	557	6.79	< 0.001	***
Flower type - staminate	1018	100	10.17	< 0.001	***
<b>Pollination success</b>					
No. of pollinator visits /					
Intercept	-0.45	0.275	-1.63	0.104	
Flower type - staminate	-0.51	0.179	-2.86	0.004	**
Pollen removal of M-anther /					
Intercept	877	147.7	5.93	< 0.001	***
Flower type - staminate	181	99.3	1.82	0.069	
Pollen removal of L-anther /					
Intercept	1091	600	1.82	0.069	***
Flower type - staminate	982	101	9.74	< 0.001	***

42 indicated as follows: \*  $P < 0.05$ ; \*\*  $P < 0.01$ ; \*\*\*  $P < 0.001$ .



**Fig. S1.** Relationships between pollinator visits and petal length in perfect and staminate flowers. Red circles, perfect flowers; blue circles, staminate flowers; black line, mean value based on GLMM result.

**Method:** To test whether the smaller petals of staminate versus perfect flowers explain the reduced rate of pollinator visitation to staminate flowers, we performed a GLMM analysis (Poisson error and log link) that incorporated the total visits per hour per flower as the response variable, flower type (perfect/staminate), petal length, and their interaction as explanatory variables, and population identity and observation date as independent random terms. We used 113/118 perfect and 104/108 staminate flowers for

which floral traits were measured and pollinator visits were observed, respectively. We  
examined the significance of each explanatory variable in the models using the Wald  
test. We performed all analyses using R, with the glmmADMB package.

**Results:** From the GLMM analysis, pollinator visitation increased significantly with  
petal length ( $p = 0.02$ ), whereas flower type and the interaction between flower type and  
petal length were not significant ( $p = 0.14$  and  $0.08$ , respectively). These results suggest  
that staminate flowers receive fewer pollinator visits because staminate flowers have  
smaller blue petals compared to perfect flowers.