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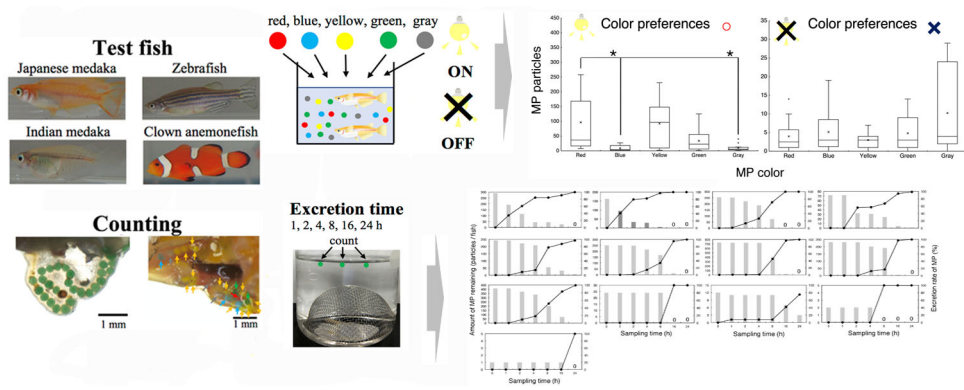
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Highlights

- Clown anemonefish ingested the most MP particles, zebrafish ingested the second-most
- The most frequently ingested MP colors were red, yellow, and green in fish
- Clown anemonefish rely on color vision to recognize for certain MP colors
- MP excretion times varied widely among individuals of the same species



1 **Color preferences and gastrointestinal-tract retention times of microplastics by**
2 **freshwater and marine fishes**

3

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15 **Abstract (196 words)**

16 We examined ingestion and retention rates of microplastics (MPs) by two freshwater
17 (Japanese medaka and zebrafish) and two marine fish species (Indian medaka and clown
18 anemonefish) to determine their color preferences and gastrointestinal-tract retention
19 times. In our ingestion experiments, clown anemonefish ingested the most MP particles,
20 followed by zebrafish, and then Japanese and Indian medaka. Next, we investigated color
21 preferences among five MP colors. Red, yellow, and green MP were ingested at higher
22 rates than grey and blue MPs for all tested fish species. To test whether these differences
23 truly reflect a recognition of and preference for certain colors based on color vision, we
24 investigated the preferences of clown anemonefish for MP colors under light and dark
25 conditions. Under dark conditions, ingestion of MP particles was reduced, and color
26 preferences were not observed. Finally, we assessed gastrointestinal-tract retention times
27 for all four fish species. Some individuals retained MP particles in their gastrointestinal
28 tracts for over 24h after ingestion. Our results show that fish rely on color vision to
29 recognize and express preferences for certain MP colors. In addition, MP excretion times
30 varied widely among individuals. Our results provide new insights into accidental MP
31 ingestion by fishes.

32

33 ***Keywords:* color preferences, microplastic, ingestion, fish**

34

35 **Introduction**

36 In recent years, microplastic (MP) pollution (i.e., pollution involving plastic particles of
37 diameter < 5 mm) has become a serious environmental problem around the world
38 (reviewed by Bagaev et al., 2021; Galarpe et al., 2021; Li et al., 2016). Pinherio et al.
39 (2021) reviewed MP concentrations in marine and freshwater environments in various
40 countries including the United Kingdom, United States, Brazil, China, France,
41 Germany, India, Italy, Australia, Indonesia, and Japan. Furthermore, MP contamination
42 has been repeatedly identified in marine (reviewed by Hidalgo-Ruz et al., 2012) and
43 freshwater (reviewed by Yang et al., 2021) environments worldwide. Therefore, the
44 ecotoxicity of MP in aquatic organisms has become a topic of increasing concern
45 (Galloway and Lewis, 2016).

46 To date, various field and laboratory studies have examined the influence of
47 MP on fishes. In the laboratory, MP ingestion and uptake into the gastrointestinal tract
48 has been observed in goldfish (*Carassius auratus*) (Grigorakis et al., 2017), zebrafish
49 (*Danio rerio*) (Kim et al., 2019), mummichog (*Fundulus heteroclitus*) (Ohkubo et al.,
50 2020), and red seabream (*Pagrus major*) (Ohkubo et al., 2020). In addition, MP has
51 been identified in wild fish captured in the Lijiang River in China (Zhang et al., 2021),
52 the Mondego Estuary in Portugal (Bessa et al., 2018), the Xingu River in Brazil
53 (Andrade et al., 2019), the Pacific coast of Ecuador (Alfaro-Núñez et al., 2021), and the

54 northern Bay of Bengal in Bangladesh (Hossain et al., 2020). These results indicate that
55 MP ingestion by fishes is widespread.

56 Various colors of MP (e.g., transparent, black, blue, grey, green, red, purple,
57 and yellow) are present in marine (Courtene-Jones et al., 2017; Dai et al., 2018; Lusher
58 et al., 2014; Martin et al., 2017) and freshwater environments (Li et al., 2018; McNeish
59 et al., 2018; Su et al., 2016; Wu et al., 2020). Colored MP has also been detected in wild
60 fish (Bessa et al., 2018; Jawad et al., 2021; Kazour et al., 2020), suggesting that colored
61 MP can potentially be mistaken for natural prey. If various marine and freshwater fishes
62 exhibit color preferences for MP ingestion, this would be important for understanding
63 the harmful influence of MP on wild fishes.

64 Another important factor for estimating the harmful influence of MP on fishes
65 is MP retention and excretion times. This information is needed to assess the extent to
66 which hazardous chemical substances adsorbed onto MPs can be absorbed into the body
67 after ingestion. However, to our knowledge, there are only a few laboratory reports that
68 currently provide this data (i.e., for goldfish [Grigorakis et al., 2017], mummichog
69 [Ohkubo et al., 2020], and red seabream [Ohkubo et al., 2020]). More data are needed
70 on retention and excretion times in various fishes including both freshwater and marine
71 species, because MP is detected in both freshwater and marine environments.

72 In our study, we examine two freshwater fish species (Japanese medaka
73 *Oryzias latipes* and zebrafish *Danio rerio*) and two marine species (Indian medaka

74 *Oryzias melastigma* and clown anemonefish *Amphiprion ocellaris*). Japanese medaka
75 typically inhabits gently flowing rivers and waterways in Japan (Sakaizumi 1986;
76 Sakaizumi and Jeon 1987), and zebrafish inhabits moderately flowing to stagnant clear
77 waters in India, Pakistan, Bangladesh, Nepal, and Bhutan (Lawrence, 2007). Japanese
78 medaka and zebrafish are widely used as model freshwater fishes for ecotoxicological
79 studies under OECD test guidelines (OECD, 2012, 2013, 2019). Indian medaka inhabits
80 coastal marine waters and freshwater bodies in Pakistan, India, Myanmar, and Thailand
81 (Dong et al., 2014). Recently, Indian medaka has also become a common choice as a
82 model marine fish for ecotoxicological studies (He et al., 2021; Horie et al., 2019;
83 Wang et al., 2020; Zheng et al., 2020). Clownfish is found throughout the Indo-Pacific
84 (Camp et al., 2016; Chen and Hsieh., 2017; Chen et al., 2018).

85 First, we identify any preferences for MP color among Japanese medaka,
86 zebrafish, Indian medaka, and clown anemonefish under laboratory conditions. Next,
87 we estimate MP retention times for all four species. The findings of this study will be
88 useful for further estimation of MP ecotoxicity in these species.

89

90 **2. Materials and methods**

91 All animal experiments were conducted according to the relevant national guidelines
92 (Act on Welfare and Management of Animals, Ministry of the Environment, Japan) and
93 the fish used in the present study were handled according to the animal care and use

94 guidelines of Kobe University. All animal experiments were approved by the
95 institutional animal care and use committee, Research Center for Inland Sea, Kobe
96 University (Permission number, 2021-04). Our research was also performed in
97 accordance with the ARRIVE guidelines.

98

99 2.1. Study organisms and MPs

100 We examined two freshwater fishes, Japanese medaka and zebrafish, and two marine
101 fishes, Indian medaka and clown anemonefish. Japanese medaka, zebrafish, Indian
102 medaka, and Clown anemonefish were bred at Kobe University (Hyogo, Japan). Both
103 marine and freshwater fishes were maintained at a water temperature of 25 ± 2 °C
104 (mean \pm SD) by using a recirculating system. Marine fishes were maintained at a
105 salinity of 32 PSU. Fish were raised under an artificial photoperiod of 12-h/12-h
106 light/dark. Marine species were kept in artificial seawater (Marine ART Hi; Osaka
107 Yakken Co. Ltd, Osaka, Japan). In this study, we used young fish and genetic sex was
108 unknown. Average total body lengths (mm) and wet body weights (mg) of each species
109 were as follows: 15.1 ± 0.1 mm and 30.0 ± 0.8 mg for Japanese medaka (age: 1-2
110 months after hatching); 12.6 ± 0.1 mm and 12.2 ± 0.4 mg for zebrafish (age: 1-2 months
111 after hatching); 16.7 ± 0.1 mm and 40.6 ± 0.8 mg for Indian medaka (age: 1-2 months
112 after hatching); and 31.1 ± 0.2 mm and 484.7 ± 11.9 mg for clown anemonefish (age: 4-
113 6 months after hatching).

114 Polyethylene microspheres were purchased from Cospheric (Santa Barbara, CA,
115 USA) in five colors (red, blue, yellow, green, and gray). The average diameter and
116 particle density of each color was as follows. Red: $219.2 \pm 22.6 \mu\text{m}$, 0.98 g/cc ; blue:
117 $279.0 \pm 17.0 \mu\text{m}$, 1.00 g/cc ; yellow: $256.9 \pm 21.2 \mu\text{m}$, 1.00 g/cc ; green: 253.6 ± 20.4
118 μm , 0.98 g/cc ; gray: $257.7 \pm 21.7 \mu\text{m}$, 1.00 g/cc (Supplementary Figure 1).

119

120 2.2. MP ingestion assays (Experiments 1 and 2)

121 A flow chart summarizing the experimental procedure for all experiments (Experiments
122 1–4) is shown in Figure 1. For MP ingestion, we conducted two experiments: in
123 Experiment 1, we exposed each of the four fish species to a single color (red, blue,
124 yellow, green, or gray) of MP; in Experiment 2, we exposed each species to a mix of the
125 five MP colors to test for the presence of a color preference. Seven fish were placed in
126 5-L glass tanks filled with 4 L of water, and two replicate tanks were used for each
127 exposure (i.e., $n = 14$ per exposure).

128 Before establishing the nominal MP concentrations for exposures, we assessed
129 the maximum MP ingestion amount for each fish species. The maximum MP ingestion
130 test lasted for 4 h (until feeding behavior was no longer observed for four fish species).
131 The Japanese medaka, zebrafish, and Indian medaka individuals that ingested the most
132 MP particles in our assessment each ingested fewer than 100 particles, and the clown
133 anemonefish that ingested the most MP particles ingested fewer than 1000 particles

134 (Figure 2). The weight of 1000 MP particles is approximately 10 mg. Because each 5-L
135 glass tank contained 4 L of water and seven fish in our exposure experiments, the
136 nominal MP concentration for Experiment 1 was set at 1.75 mg/L for Japanese medaka,
137 zebrafish, and Indian medaka, and 17.5 mg/L for clown anemonefish. For Experiment 2,
138 the concentration for each MP color was set at 1.75 mg/L (i.e., a total MP concentration
139 of 8.75 mg/L) for each of the three fish species. In case of clown anemonefish, the
140 concentration for each MP color was set at 17.5 mg/L (i.e., a total MP concentration of
141 87.5 mg/L).

142 Fish were placed into 5-L glass tanks 24 h prior to the start of each exposure
143 experiment, and were not fed during this time to ensure that their gastrointestinal tracts
144 would be empty. Each MP exposure test lasted for 4 h (until feeding behavior was no
145 longer observed for four fish species). After 4 h of exposure, fish were anesthetized
146 (MS-222 at a concentration of 200 mg/L) and their gastrointestinal tracts dissected, and
147 the number of MP particles in each gastrointestinal tract was counted under a
148 stereomicroscope (SZ61, Olympus).

149

150 2.3. MP color-preference assays (Experiments 3)

151 In Experiment 3, we examined clown anemonefish (which ingested the most MP
152 particles in pretests prior to Experiment 1) to clarify whether fish rely on their color
153 vision to recognize MP colors. Exposure conditions were similar to those in Experiment

154 2. Seven fish were placed in 5-L glass tanks, two replicate tanks were used for each
155 exposure (i.e., $n = 14$ per condition), and the exposure concentration for each MP color
156 was set to 17.5 mg/L (i.e., total MP concentration of 87.5 mg/L).

157 Ten days prior to each exposure experiment, in order to reset the circadian
158 rhythm of the study animals, the husbandry schedules for the fish were reversed; i.e.,
159 feeding was performed in the dark, and water changes were performed in the light. Fish
160 were then placed into 5-L glass tanks 24 h prior to the start of each exposure
161 experiment, and were not fed during this time to ensure that their gastrointestinal tracts
162 would be empty. Next, MP exposure tests were carried out for 4 h (until feeding
163 behavior was no longer observed for clown anemonefish) under dark conditions. After
164 the 4-h exposure, fish were anesthetized (MS-222 at a concentration of 200 mg/L) and
165 their gastrointestinal tracts dissected, and the number of MP particles in each
166 gastrointestinal tract was counted under a stereomicroscope (SZ61, Olympus).

167

168 2.4. MP retention in and excretion from the gastrointestinal tract (Experiments 4)

169 In Experiment 4, we examined the gastrointestinal tract retention and excretion times of
170 green MP (which was the most commonly ingested MP color in Experiment 1) in each
171 fish species. Exposure conditions were the same as in Experiment 1. After 4 h of
172 exposure, each fish was transferred from its 5-L glass exposure tank to a 1-L glass
173 beaker with a stainless-steel screen at the bottom for the excretion experiment.

174 The fish were confined under the stainless-steel screen (mesh size: 2 mm (width)*2 mm
175 (height)) (to prevent re-feeding for the excretion MP) so that the MP, once excreted,
176 could swim freely to the surface due to their low density. Once at the surface, they were
177 collected by using glass pipett and were counted under a stereomicroscope in directly
178 (SZ61, Olympus). A total of 14 glass beakers were prepared to accommodate the 14
179 experimental fish. After 1, 2, 4, 8, 16, and 24 h, the number of MP particles excreted
180 from each fish was counted, and all MP were removed from the 1-L beaker. After the
181 excretion test, fish were anesthetized (MS-222 at a concentration of 200 mg/L) and their
182 gastrointestinal tracts dissected, and the number of MP particles in each gastrointestinal
183 tract was counted under a stereomicroscope (SZ61, Olympus).

184

185 2.5. Statistical analysis

186 All data were analyzed in Microsoft Excel. To analyze significant differences between
187 light and dark conditions, we used open source statistical software R ([http://www.R-](http://www.R-project.org/)
188 [project.org/](http://www.R-project.org/)) and the package *Rcmdr* (Fox and Bouchet-Valat 2018) to test for
189 homogeneity of variance using Bartlett's test (significance level, 5%), and tested for
190 significant differences using Steel's test. To analyze color preference in clown
191 anemonefish, we used Steel–Dwass multiple-comparison tests (significance level, 5%).

192

193 3. Results

194 3.1. MP uptake by the four fish species

195 Table 1 shows a summary of the number of fish of each species in Experiment 1 that
196 ingested MP of each color. Zebrafish and clown anemonefish readily ingested MP of all
197 colors, but Japanese medaka and Indian medaka mainly ingested green MP. The
198 proportion of individuals that ingested MP of any color was highest for clown
199 anemonefish, followed by zebrafish, Japanese medaka, and Indian medaka.

200 Figure 2 shows the number of MP particles ingested by the four fish species. In
201 Japanese medaka, almost all individuals ingested fewer than 10 MP particles, with the
202 exception of one individual that ingested 31 blue MP particles and three individuals that
203 ingested 28, 29, and 77 green MP particles, respectively (Fig. 2a). In zebrafish, almost
204 all individuals ingested between 10 and 30 MP particles, and the most MP particles
205 were ingested by an individual that consumed 63 yellow MP particles (Fig. 2b). In
206 Indian medaka, almost all individuals ingested fewer than 10 MP particles, with the
207 exception of two individuals that ingested 12 and 24 red MP particles, respectively and
208 one individual that ingested 15 green MP particles (Fig. 2c). Clown anemonefish
209 ingested the most MP in total. Almost all individuals ingested more than 100 MP
210 particles, including one individual that ingested 744 red MP particles (Fig. 2d). There
211 were no consistent differences in MP intake by color across the four species.

212

213 3.2. Color preferences for MP intake

214 The number of fish that ingested MP in Experiment 2 was as follows: Japanese medaka,
215 4 of 14; zebrafish, 6 of 14; Indian medaka, 5 of 14; and clown anemonefish, 12 of 14
216 (Fig. 3). Figure 3 shows color preferences for MP ingestion in Japanese medaka (Fig.
217 3a), zebrafish (Fig. 3b), and Indian medaka (Fig. 3c); the results for clown anemonefish
218 are shown separately (Fig. 4b). Overall, the MP colors that were most frequently
219 ingested by Japanese medaka, zebrafish, Indian medaka, and clown anemonefish were
220 red, yellow, and green, and the least frequently ingested colors were blue and gray,
221 although significant differences could not analyze due to the number of MP particles
222 ingested were low level.

223 Next, we examined the presence or absence of color preferences (Fig. 4;
224 Experiment 2 and 3). In this experiment, we used clown anemonefish, which ingested
225 the most MP particles of the species examined in Experiment 1. Clown anemonefish
226 ingested fewer MP particles under dark conditions than under light conditions (Fig. 4a).
227 Moreover, whereas red, yellow, and green MP were preferred under light conditions
228 (Fig. 4b), no color preference was apparent under dark conditions (Fig. 4b). Results of
229 steel–Dwass multiple-comparison tests (significance level, 5%), showed that red MPs
230 was significantly more ingested in comparison to blue and gray MP under light
231 conditions (Fig. 4c), on the other hand, under dark conditions, no significant differences
232 were observed (Fig. 4c).

233

234 3.3. Time-course of MP retention in and excretion from the gastrointestinal tract
235 The results of the MP retention and excretion tests were as follows. Eight of 14
236 Japanese medaka (57%) excreted all of the MP contained in their gastrointestinal tracts
237 within 24 h (Figure 5); some residual MP remained in the gastrointestinal tracts after 24
238 h in the remaining six fish. Nine of 14 zebrafish (64%) excreted all of the MP contained
239 in their gastrointestinal tracts within 24 h (Figure 6); only one individual excreted less
240 than 90% of its ingested MP within the 24-h observation period. All 14 Indian medaka
241 excreted all of the MP contained in their gastrointestinal tracts within 24 h (Figure 7).
242 Nine of 13 clown anemonefish (69%) excreted all of the MP contained in their
243 gastrointestinal tracts within 24 h (Figure 8); only one individual excreted less than 20%
244 of its ingested MP within the 24-h observation period.

245 There was no consistent pattern of MP excretion among fish species, and the
246 excretion time varied widely among individuals of the same species. In addition, there
247 was no apparent difference in MP excretion time caused by the presence (clown
248 anemonefish) or absence (Japanese medaka, zebrafish, and Indian medaka) of a
249 stomach.

250

251 **4. Discussion**

252 This study is the first to evaluate whether fish differentiate among five distinct colors of
253 MP under laboratory conditions and can preferentially ingest MP on this basis.

254 Our results on MP ingestion highlight the magnitude of variability in MP
255 ingestion among species and individuals. Recently, Ohkubo et al (2020) reported that
256 the number of MP particles (diameter, 250–300 μm ; color, yellow) identified in the
257 gastrointestinal tracts of mummichog was 352 ± 240 at an exposure concentration of 3
258 mg/L, and the number identified in red seabream was 41.8 ± 14.9 at an exposure
259 concentration of 0.9 mg/L. In the present study, the average number of MP particles of
260 the type examined in the above reports (diameter, 250–300 μm ; color, yellow) identified
261 in gastrointestinal tracts was 0, 25 ± 18 , and 1 in Japanese medaka, zebrafish, and
262 Indian medaka, respectively, at an exposure concentration of 1.75 mg/L; in clown
263 anemonefish the number was 215 ± 198 at an exposure concentration of 17.5 mg/L.
264 Some of the species (Japanese medaka, zebrafish, and Indian medaka) do not possess
265 stomachs, and clown anemonefish do. However, the number of ingested MP particles
266 does not appear to depend strongly on the presence or absence of a stomach. Further
267 studies on other fish species could help identify commonalities between fish that are or
268 are not prone to accidental MP ingestion.

269 Our results on color preferences contrast with reports from the field. To date,
270 there have been numerous reports of plastics in the gastrointestinal tracts of wild fishes.
271 For example, blue and black MP filaments and blue, green, and black MP fragments
272 were frequently identified in Atlantic horse mackerel (*Trachurus trachurus*) from the
273 central Mediterranean Sea (Chenet et al., 2021). Along the southwest coast of the United

274 Kingdom, grey/transparent MP were the most frequently ingested by small-spotted
275 catshark (*Scyliorhinus canicula*), and green was the least frequently ingested (Morgan et
276 al., 2021). In the Adriatic Sea in Italy, black, tan, and blue MP fragments were
277 frequently found in the stomachs of *Sardina pilchardus*, and black and blue MP
278 fragments were frequently identified in the stomachs of *Engraulis encrasicolus* (Renzi
279 et al., 2019). By contrast, in our laboratory study, red, yellow, and green MP particles
280 were frequently observed in the gastrointestinal tracts of zebrafish, Indian medaka, and
281 clown anemonefish, and blue and gray particles were less common. This may indicate
282 that color preferences differ between field and laboratory conditions. Future studies on
283 the same species in the field and laboratory will clarify whether this is the case.

284 In general, vertebrates possess two types of photoreceptor cells: cones and rods.
285 Cone cells enable color vision during light conditions, and rod cells, which do not
286 distinguish among colors, are more sensitive under low-light conditions, meaning that
287 they are used most heavily in the dark. Bony fishes such as goldfish and zebrafish have
288 four spectral cone types comprising alternating rows of double cones with red (LWS)
289 and green (RH2) members and single blue (SWS2) and UV (SWS1) cones (Allison et
290 al., 2010; Baden and Osorio, 2019; Engström, 1960; Raymond et al., 1993). On the
291 other hand, the few shark species that have been studied only possess a single spectral
292 class of cone with LWS (Hart et al., 2011; Hart et al., 2020; Theiss et al., 2012),
293 indicating that they are almost certainly cone monochromats and do not possess color

294 vision (Schluessel et al., 2014). Recently, Mitchell et al (2021) reported that clown
295 anemonefish has four spectral cone types including LWS, RH2, SWS2, and SWS1, and
296 rhodopsin 1 (rod opsin, RH1). Therefore, this report and our results indicate that clown
297 anemonefish are able to recognize MP colors. Under light conditions, the species
298 showed a preference for red, yellow, and green MP, but under dark conditions, the
299 number of MP particles ingested declined and color preferences were not observed.
300 Various colors of MP have been identified in the field (Chenet et al., 2021; Morgan et
301 al., 2021; Renzi et al., 2019), and our study identified color preferences for MP
302 ingestion in clown anemonefish (a marine species). Further studies are needed to
303 determine the presence or absence of color preferences in other fish species to further
304 our understanding of MP ingestion in the field. In addition, color preference data could
305 help clarify whether fish are misidentifying MP as food.

306 To our knowledge, reports on MP retention and excretion times in fish in the
307 laboratory remain scarce, although there have been many reports of MP being identified
308 in the gastrointestinal tracts of wild fishes (Bessa et al., 2018; Jawad et al., 2021;
309 Kazour et al., 2020). Excretion times observed in the present study can be divided into
310 three patterns: (1) immediate excretion, in which most MP is excreted within the first 4
311 h; (2) gradual excretion, in which MP is excreted gradually over 24 h; and (3) delayed
312 excretion, in which most MP is excreted after 16 h. A previous study (Ohkubo et al.,
313 2020) on mummichog and red seabream also identified the above three patterns. In the

314 future, it will be necessary to investigate whether these three patterns apply to other fish
315 species. In addition, why were there observed the difference of excretion patterns in
316 same fish species? Various factors such as gender differences, health conditions, or
317 fitness differences, etc. can be considered, therefore, further research is needed in the
318 near future.

319 In contrast to previous reports, our results show that MP can remain in fish
320 gastrointestinal tracts for a prolonged period following ingestion. Ohkubo et al (2020)
321 reported that over 95% of ingested MP was excreted within 25 h in mummichog and red
322 seabream. Similarly, Naidoo and Glassom (2019) reported that only a few particles of
323 negligible mass remained 24 h after MP exposure in *Ambassis dussumieri*. By
324 comparison, we found that some Japanese medaka, zebrafish, and clown anemonefish
325 retained MP in the gastrointestinal tract even 24 h after MP ingestion. This indicates
326 that the time required for MP excretion does not depend on fish species, but on the
327 individual. In the future, we intend to examine excretion rates across a longer time
328 horizon to clarify the time required to discharge all ingested MP particles.

329

330 **5. Conclusion**

331 To assess the impact of MP on fish, it is essential to first understand accidental MP
332 ingestion and gastrointestinal-tract retention times. Ours is the first study to identify
333 color preferences among teleost fishes for MP ingestion, and also provides valuable data

334 on MP excretion times for both freshwater and marine fishes. The freshwater zebrafish
335 and marine clown anemonefish both ingested large numbers of MP particles compared
336 to Japanese and Indian medaka. Also, red, yellow, and green MP was more frequently
337 ingested than grey and blue MP overall. The number of MP particles ingested was
338 reduced, and color preferences were not observed under dark conditions in clown
339 anemonefish. These findings suggest that clown anemonefish rely on color vision to
340 recognize MP colors and express color preferences. MP excretion times varied widely
341 among fish of the same species, and some individuals still had MP particles remaining
342 in their gastrointestinal tracts more than 24 h after exposure. These findings provide
343 new insights into accidental MP ingestion by fishes.

344

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350

351

352 **References**

- 353 Alfaro-Núñez, A., Astorga, D., Cáceres-Farías, L., Bastidas, L., Soto, Villegas, C.,
354 Macay, K., Christensen, J.H., 2021. Microplastic pollution in seawater and marine
355 organisms across the Tropical Eastern Pacific and Galápagos. *Sci. Rep.* 11, 6424.
- 356 Allison, W.T., Barthel, L.K., Skebo, K.M., Takechi, M., Kawamura, S., Raymond, P.A.,
357 2010. Ontogeny of cone photoreceptor mosaics in zebrafish. *J. Comp. Neurol.* 518,
358 4182–4195.
- 359 Andrade, M.C., Winemiller, K.O., Barbosa, P.S., Fortunati, A., Chelazzi, D., Cincinelli,
360 A., Giarrizzo, T., 2019. First account of plastic pollution impacting freshwater
361 fishes in the Amazon: Ingestion of plastic debris by piranhas and other serrasalmids
362 with diverse feeding habits. *Environ. Pollut.* 244, 766–773.
- 363 Baden, T., Osorio, D., 2019. The Retinal Basis of Vertebrate Color Vision. *Annu. Rev.*
364 *Vis. Sci.* 5, 177–200.
- 365 Bagaev, A., Esiukova, E., Litvinyuk, D., Chubarenko, I., Veerasingam, S.,
366 Venkatachalapathy, R., Verzhevskaya, L., 2021. Investigations of plastic
367 contamination of seawater, marine and coastal sediments in the Russian seas: a
368 review. *Environ. Sci. Pollut. Res. Int.* doi: 10.1007/s11356-021-14183-z.
- 369 Bessa, F., Barría, P., Neto, J.M., Frias, J.P.G.L., Otero, V., Sobral, P., Marques, J.C.,
370 2018. Occurrence of microplastics in commercial fish from a natural estuarine
371 environment. *Mar. Pollut. Bull.* 128, 575–584.

372 Camp, E.F., Hobbs, J.P., De Brauwer, M., Dumbrell, A.J., Smith, D.J., 2016.
373 Cohabitation promotes high diversity of clownfishes in the Coral Triangle. Proc.
374 Biol. Sci. 283, 20160277.

375 Chen, T.H., Hsieh, C.Y., 2017. Fighting Nemo: Effect of 17 α -ethinylestradiol (EE2) on
376 aggressive behavior and social hierarchy of the false clown anemonefish
377 *Amphiprion ocellaris*. Mar. Pollut. Bull. 124, 760-766.

378 Chen, T.H., Hsieh, C.Y., Ko, F.C., Cheng, J.O., 2018. Effect of the UV-filter
379 benzophenone-3 on intra-colonial social behaviors of the false clown anemonefish
380 (*Amphiprion ocellaris*). Sci. Total. Environ. 644, 1625-1629.

381 Chenet, T., Mancina, A., Bono, G., Falsone, F., Scannella, D., Vaccaro, C., Baldi, A.,
382 Catani, M., Cavazzini, A., Pasti, L., 2021. Plastic ingestion by Atlantic horse
383 mackerel (*Trachurus trachurus*) from central Mediterranean Sea: A potential cause
384 for endocrine disruption. Environ. Pollut. 284, 117449.

385 Christian, L., 2007. The husbandry of zebrafish (*Danio rerio*): A review. Aquaculture.
386 269, 1–20 .

387 Courtene-Jones, W., Quinn, B., Gary, S.F., Mogg, A.O.M., Narayanaswamy, B.E.,
388 2017. Microplastic pollution identified in deep-sea water and ingested by benthic
389 invertebrates in the Rockall Trough, North Atlantic Ocean. Environ. Pollut. 231,
390 271–280.

- 391 Dai, Z., Zhang, H., Zhou, Q., Tian, Y., Chen, T., Tu, C., Fu, C., Luo, Y., 2018.
392 Occurrence of microplastics in the water column and sediment in an inland sea
393 affected by intensive anthropogenic activities. *Environ. Pollut.* 242, 1557–1565.
- 394 Dong, S., Kang, M., Wu, X., Ye, T., 2014. Development of a promising fish model
395 (*Oryzias melastigma*) for assessing multiple responses to stresses in the marine
396 environment. *Biomed. Res. Int.* 2014, 563131.
- 397 Engström, K. 1960. Cone types and cone arrangements in the retina of some cyprinids.
398 *Acta. Zool.* 41, 277–295.
- 399 Galarpe, V.R.K.R., Jaraula, C.M.B., Paler, M.K.O., 2021. The nexus of macroplastic
400 and microplastic research and plastic regulation policies in the Philippines marine
401 coastal environments. *Mar. Pollut. Bull.* 167, 112343.
- 402 Galloway, T.S., Lewis, C.N., 2016. Marine microplastics spell big problems for future
403 generations. *Proc. Natl. Acad. Sci. U. S. A.* 113, 2331–3.
- 404 Grigorakis, S., Mason, S.A., Drouillard, K.G., 2017. Determination of the gut retention
405 of plastic microbeads and microfibers in goldfish (*Carassius auratus*).
406 *Chemosphere.* 169, 233–238.
- 407 Hart, N.S., Theiss, S.M., Harahush, B.K., Collin, S.P., 2011. Microspectrophotometric
408 evidence for cone monochromacy in sharks. *Naturwissenschaften.* 98, 193–201.

409 Hart, N.S., Lamb, T.D., Patel, H.R., Chuah, A., Natoli, R.C., Hudson, N.J., Cutmore,
410 S.C., Davies, W.I.L., Collin, S.P., Hunt, D.M., 2020. Visual Opsin Diversity in
411 Sharks and Rays. *Mol. Biol. Evol.* 37, 811–827.

412 He, S., Yu, D., Li, P., Zhang, M., Xing, S., Sun, C., Li, Z.H., 2021. Triphenyltin
413 exposure causes changes in health-associated gut microbiome and metabolites in
414 marine medaka. *Environ. Pollut.* 288, 117751.

415 Hidalgo-Ruz, V., Gutow, L., Thompson, R.C., Thiel, M., 2012. Microplastics in the
416 marine environment: a review of the methods used for identification and
417 quantification. *Environ. Sci. Technol.* 46, 3060–3075.

418 Horie, Y., Kanazawa, N., Suzuki, A., Yonekura, K., Chiba, T., 2019. Influences of
419 Salinity and Organic Compounds on Embryo Development in Three Medaka
420 *Oryzias* Congeners with Habitats Ranging from Freshwater to Marine. *Bull.*
421 *Environ. Contam. Toxicol.* 103, 411–415.

422 Hossain, M.S., Sarker, S., Sharifuzzaman, S.M., Chowdhury, S.R., 2020. Primary
423 productivity connects hilsa fishery in the Bay of Bengal. *Sci. Rep.* 10, 5659.

424 Kim, S.W., Chae, Y., Kim, D., An, Y.J., 2019. Zebrafish can recognize microplastics as
425 inedible materials: Quantitative evidence of ingestion behavior. *Sci. Total. Environ.*
426 649, 156–162.

427 Jawad, L.A., Adams, N.J., Nieuwoudt, M.K., 2021. Ingestion of microplastics and
428 mesoplastics by *Trachurus declivis* (Jenyns, 1841) retrieved from the food of the

429 Australasian gannet *Morus serrator*: First documented report from New Zealand.
430 Mar. Pollut. Bull. 170, 112652.

431 Kazour, M., Jemaa, S., El, Rakwe, M., Duflos, G., Hermabassiere, L., Dehaut, A., Le,
432 Bihanic, F., Cachot, J., Cornille, V., Rabhi, K., Khalaf, G., Amara, R., 2020.
433 Juvenile fish caging as a tool for assessing microplastics contamination in estuarine
434 fish nursery grounds. Environ. Sci. Pollut. Res. Int. 27, 3548–3559.

435 Lawrence, C., 2007. The husbandry of zebrafish (*Danio rerio*): A review. Aquaculture.
436 269, 1-29.

437 Li, W.C., Tse, H.F., Fok, L., 2016. Plastic waste in the marine environment: A review
438 of sources, occurrence and effects. Sci. Total. Environ. 566-567, 333-349.

439 Li, X., Chen, L., Mei, Q., Dong, B., Dai, X., Ding, G., Zeng, E.Y., 2018. Microplastics
440 in sewage sludge from the wastewater treatment plants in China. Water. Res. 142,
441 75-85.

442 Lusher, A.L., Burke, A., O'Connor, I., Officer, R., 2014. Microplastic pollution in the
443 Northeast Atlantic Ocean: validated and opportunistic sampling. Mar. Pollut. Bull.
444 88, 325–333.

445 Martin, J., Lusher, A., Thompson, R.C., Morley, A., 2017. The Deposition and
446 Accumulation of Microplastics in Marine Sediments and Bottom Water from the
447 Irish Continental Shelf. Sci. Rep. 7, 10772.

448 McNeish, R.E., Kim, L.H., Barrett, H.A., Mason, S.A., Kelly, J.J., Hoellein, T.J., 2018.
449 Microplastic in riverine fish is connected to species traits. *Sci. Rep.* 8, 11639.

450 Mitchell, L.J., Cheney, K.L., Lührmann, M., Marshall, J., Michie, K., Cortesi, F., 2021.
451 Molecular Evolution of Ultraviolet Visual Opsins and Spectral Tuning of
452 Photoreceptors in Anemonefishes (Amphiprioninae). *Genome. Biol. Evol.* 13(10),
453 evab184.

454 Morgan, E., Hutchinson, D., Gaion, A., 2021. Plastic Ingestion by the Small-Spotted
455 Catshark (*Scyliorhinus canicula*) from the South West Coast of the United
456 Kingdom. *Bull. Environ. Contam. Toxicol.* 106, 910–915.

457 Naidoo, T., Glassom, D., 2019. Decreased growth and survival in small juvenile fish,
458 after chronic exposure to environmentally relevant concentrations of microplastic.
459 *Mar. Pollut. Bull.* 145, 254–259.

460 OECD., 2012. Guidelines for the testing of chemicals, test no. 229: Fish Short Term
461 Reproduction Assay. OECD Publishing, Paris.

462 OECD., 2013. Guidelines for the testing of chemicals, test no. 210: Fish, Early-life
463 Stage Toxicity Test. OECD Publishing, Paris.

464 OECD., 2019. Guidelines for the testing of chemicals, test no. 203: Fish, Acute Toxicity
465 Test. OECD Publishing, Paris.

466 Ohkubo, N., Ito, M., Hano, T., Kono, K., Mochida, K., 2020. Estimation of the uptake
467 and gut retention of microplastics in juvenile marine fish: Mummichogs

468 (Fundulusheteroclitus) and red seabreams (*Pagrus major*). *Mar. Pollut. Bull.* 160,
469 111630.

470 Pinheiro, L.M., Agostini, V.O., Lima, A.R.A., Ward, R.D., Pinho, G.L.L., 2021. The
471 fate of plastic litter within estuarine compartments: An overview of current
472 knowledge for the transboundary issue to guide future assessments. *Environ.*
473 *Pollut.* 279, 116908.

474 Raymond, P.A., Barthel, L.K., Rounsifer, M.E., Sullivan, S.A., Knight, J.K., 1993.
475 Expression of rod and cone visual pigments in goldfish and zebrafish: A rhodopsin-
476 like gene is expressed in cones. *Neuron.* 10, 1161–1174.

477 Renzi, M., Specchiulli, A., Blašković, A., Manzo, C., Mancinelli, G., Cilenti, L., 2019.
478 Marine litter in stomach content of small pelagic fishes from the Adriatic Sea:
479 sardines (*Sardina pilchardus*) and anchovies (*Engraulis encrasicolus*). *Environ. Sci.*
480 *Pollut. Res. Int.* 26, 2771–2781.

481 Sakaizumi, M., 1986. Genetic divergence in wild populations of the medaka *Oryzias*
482 *latipes* (Pisces: Oryziatidae) from Japan and China. *Genetica.* 69, 119–125.

483 Sakaizumi, M., Jeon, S.R., 1987. Two divergent groups in the wild populations of
484 medaka *Oryzias latipes* (Pisces: Oryziatidae) in Korea. *Kor. J. Limnol.* 20, 13–20.

485 Schluessel, V., Rick, I.P., Plischke, K., 2014. No rainbow for grey bamboo sharks:
486 evidence for the absence of colour vision in sharks from behavioural discrimination
487 experiments. *J. Comp. Physiol. A.* 200, 939–947.

488 Su L, Xue Y, Li L, Yang D, Kolandhasamy P, Li D, Shi H., 2016. Microplastics in
489 Taihu Lake, China. *Environ. Pollut.* 216, 711–719.

490 Theiss, S.M., Davies, W.I., Collin, S.P., Hunt, D.M., Hart, N.S., 2012. Cone mono-
491 chromacy and visual pigment spectral tuning in wobbegong sharks. *Biol. Lett.* 8,
492 1019–1022.

493 Wang, Z., Yeung, K.W.Y., Zhou, G.J., Yung, M.M.N., Schlekat, C.E., Garman, E.R.,
494 Gissi, F., Stauber, J.L., Middleton, E.T., Lin, Wang, Y.Y., Leung, K.M.Y., Acute
495 and chronic toxicity of nickel on freshwater and marine tropical aquatic organisms.
496 *Ecotoxicol. Environ. Saf.* 206, 111373.

497 Wu, P., Tang, Y., Dang, M., Wang, S., Jin, H., Liu, Y., Jing, H., Zheng, C., Yi, S., Cai,
498 Z., 2020. Spatial-temporal distribution of microplastics in surface water and
499 sediments of Maozhou River within Guangdong-Hong Kong-Macao Greater Bay
500 Area. *Sci. Total. Environ.* 717, 135187 .

501 Yang, L., Zhang, Y., Kang, S., Wang, Z., Wu, C., 2021. Microplastics in freshwater
502 sediment: A review on methods, occurrence, and sources. *Sci. Total. Environ.* 754,
503 141948.

504 Zheng, Y., Li, Y., Yue, Z., Samreen, Li, Z., Li, X., Wang, J., 2020. Teratogenic effects
505 of environmentally relevant concentrations of phenanthrene on the early
506 development of marine medaka (*Oryzias melastigma*). *Chemosphere.* 254, 126900.

507 Zhang, L., Xie, Y., Zhong, S., Liu, J., Qin, Y., Gao, P., 2021. Microplastics in
508 freshwater and wild fishes from Lijiang River in Guangxi, Southwest China. *Sci.*
509 *Total. Environ.* 755, 142428.
510

511 **Figures**

512 Figure 1. Experimental flow chart. In Experiment 1, fish from each of the four species
513 were each exposed to a single color (red, blue, yellow, green, or gray) of microplastic
514 (MP). In Experiment 2, fish were exposed to a mix of the five MP colors under light
515 conditions. In Experiment 3, clown anemonefish were exposed to a mix of the five MP
516 colors under dark conditions. In Experiment 4, we determined gastrointestinal tract
517 retention and excretion times in each of the four fish species using green MP.

518

519 Figure 2. The number of microplastic (MP) particles in the gastrointestinal tracts of **(a)**
520 Japanese medaka, **(b)** zebrafish, **(c)** Indian medaka, and **(d)** clown anemonefish after
521 exposure to various MP colors. Circles show values for each individual.

522

523 Figure 3. The number of microplastic (MP) particles of various colors observed in the
524 gastrointestinal tracts of **(a)** Japanese medaka, **(b)** zebrafish, and **(c)** Indian medaka after
525 exposure to a mix of five MP colors. *X*-axis values show ID numbers for each fish: i.e.,
526 exposure tests were conducted on a total of four Japanese medaka, six zebrafish, and
527 five Indian medaka.

528

529 Figure 4. Color preferences of clown anemonefish under light and dark conditions. The
530 number of fish that ingested MP was 12 of 14 in both light and dark conditions. **(a)** The

531 number of microplastic (MP) particles observed in clown anemonefish gastrointestinal
532 tracts under light (circles) and dark (triangles) conditions. **(b)** The number of MP
533 particles of various colors observed in the gastrointestinal tracts of clown anemonefish
534 under light conditions and dark conditions after exposure to a mix of five MP colors. *X*-
535 axis values show ID numbers for each fish: i.e., exposure tests were conducted on a
536 total of twelve individuals under light and dark conditions. **(c)** Box plot of the number
537 of MP particles of various colors observed in the gastrointestinal tracts of clown
538 anemonefish. *Values that are significantly different (Steel's test or Steel–Dwass
539 multiple-comparison tests; * $P < 0.05$)

540

541 Figure 5. Microplastic (MP) excretion in Japanese medaka. Bars show numbers of
542 ingested MP particles remaining in fish gastrointestinal tracts at each sampling time,
543 and lines show % of total excreted MPs at each sampling time.

544

545 Figure 6. Microplastic (MP) excretion in zebrafish. Bars show numbers of ingested MP
546 particles remaining in fish gastrointestinal tracts at each sampling time, and lines
547 show % of total excreted MPs at each sampling time.

548

549 Figure 7. Microplastic (MP) excretion in Indian medaka. Bars show numbers of
550 ingested MP particles remaining in fish gastrointestinal tracts at each sampling time,
551 and lines show % of total excreted MPs at each sampling time.

552

553 Figure 8. Microplastic (MP) excretion in clown anemonefish. Bars show numbers of
554 ingested MP particles remaining in fish gastrointestinal tracts at each sampling time,
555 and lines show % of total excreted MPs at each sampling time.

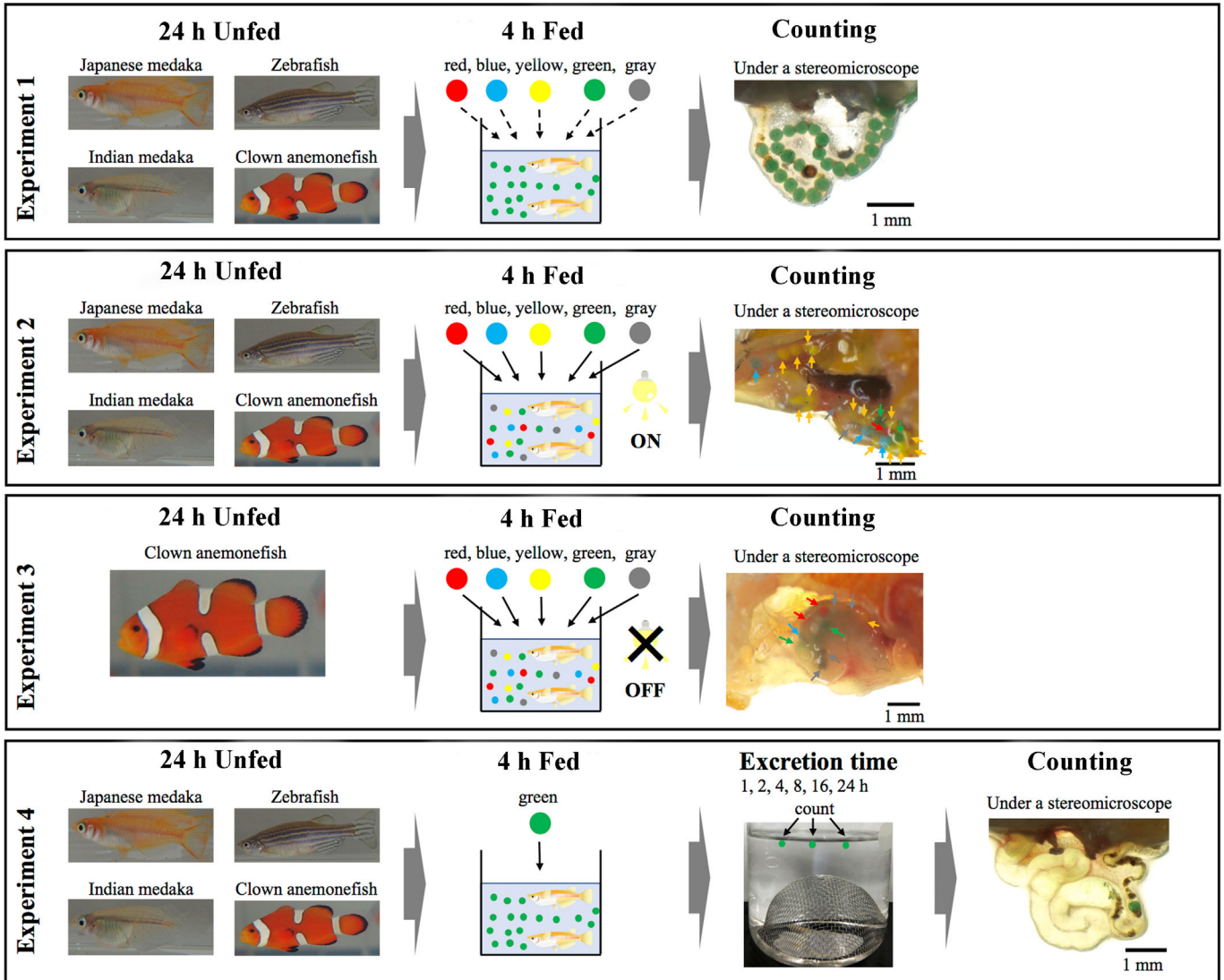
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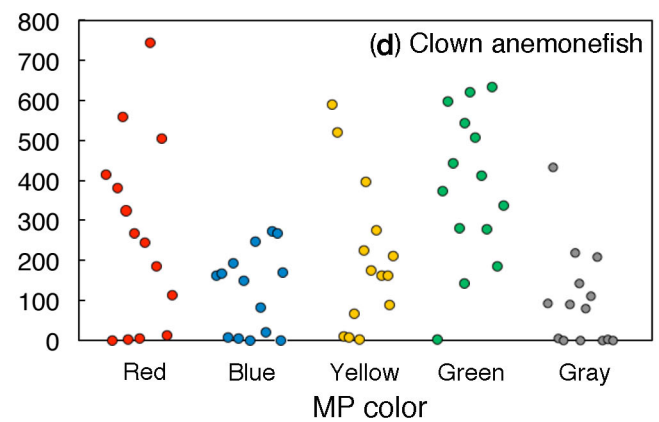
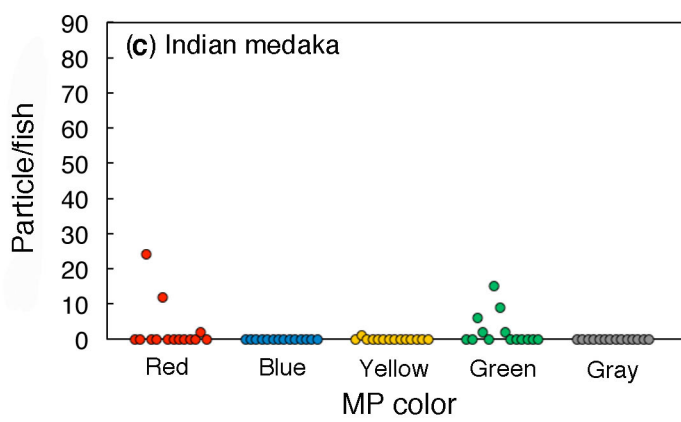
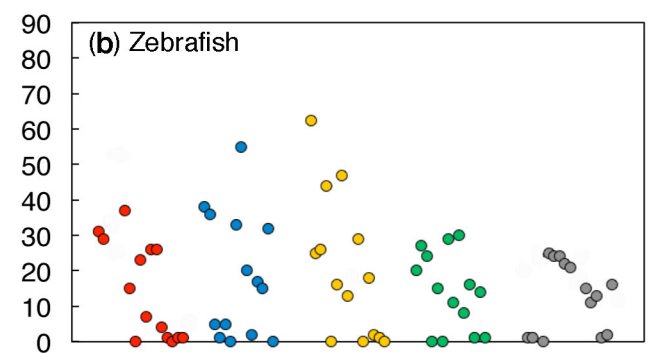
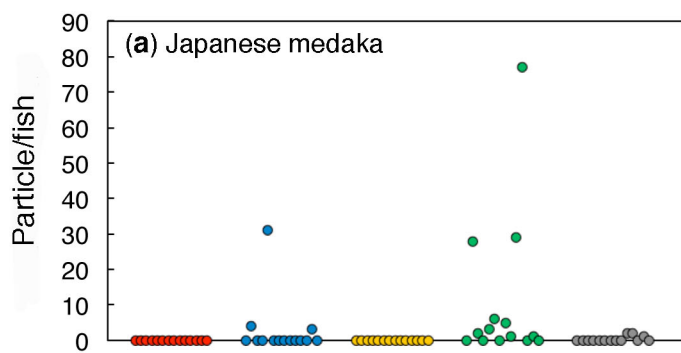
557 Supplementary Figure 1. Photographs of polyethylene microspheres in the five colors
558 used in the study: (a) red, (b) blue, (c) yellow, (d) green, and (e) gray. Average particle
559 diameters for each color (f) were measured by using a stereomicroscope (SZ61,
560 Olympus). Error bars show \pm standard deviation ($n = 100$ per color).

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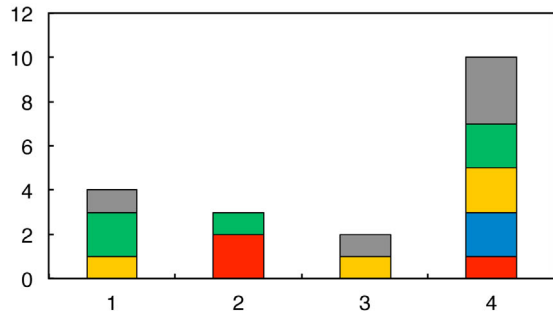
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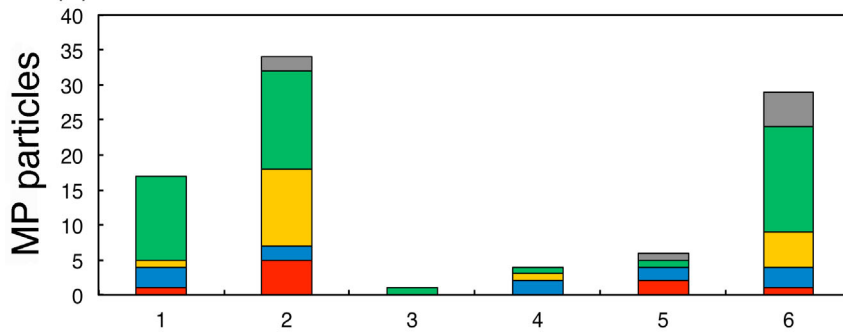




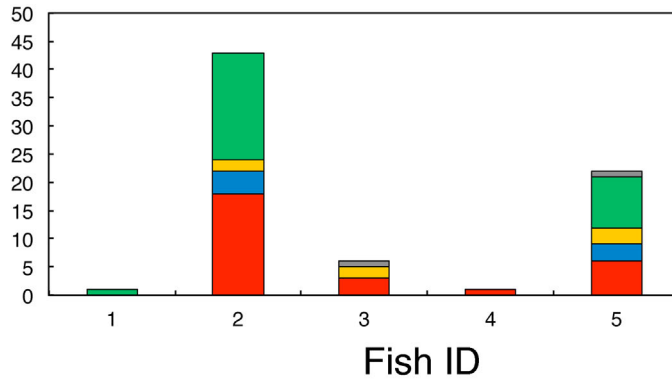
(a) Japanese medaka

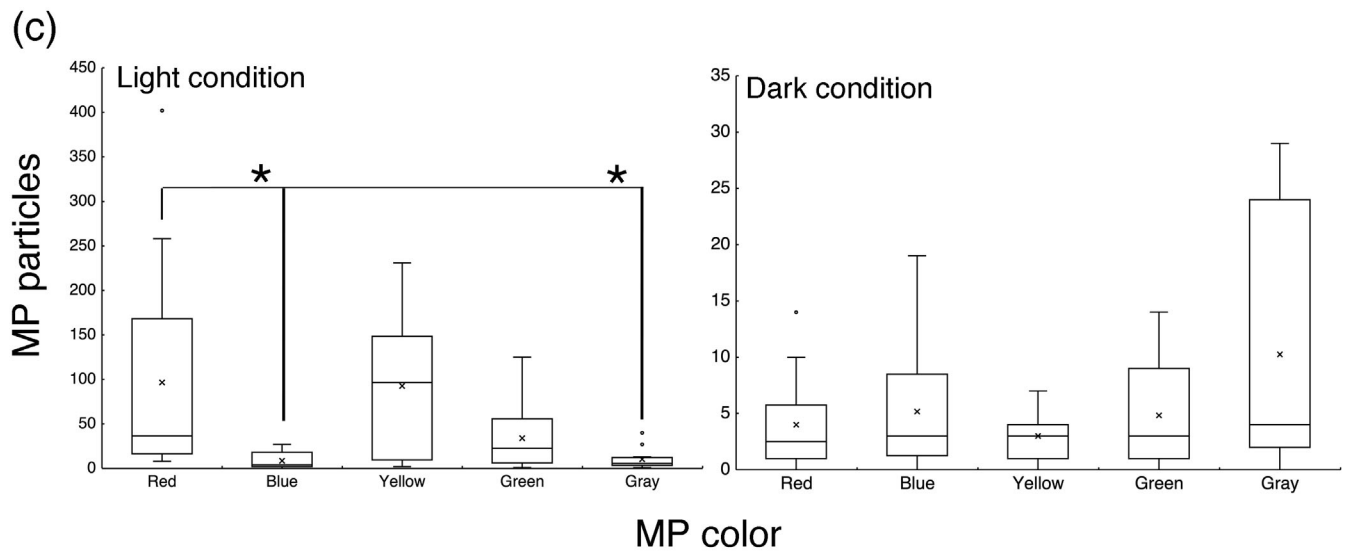
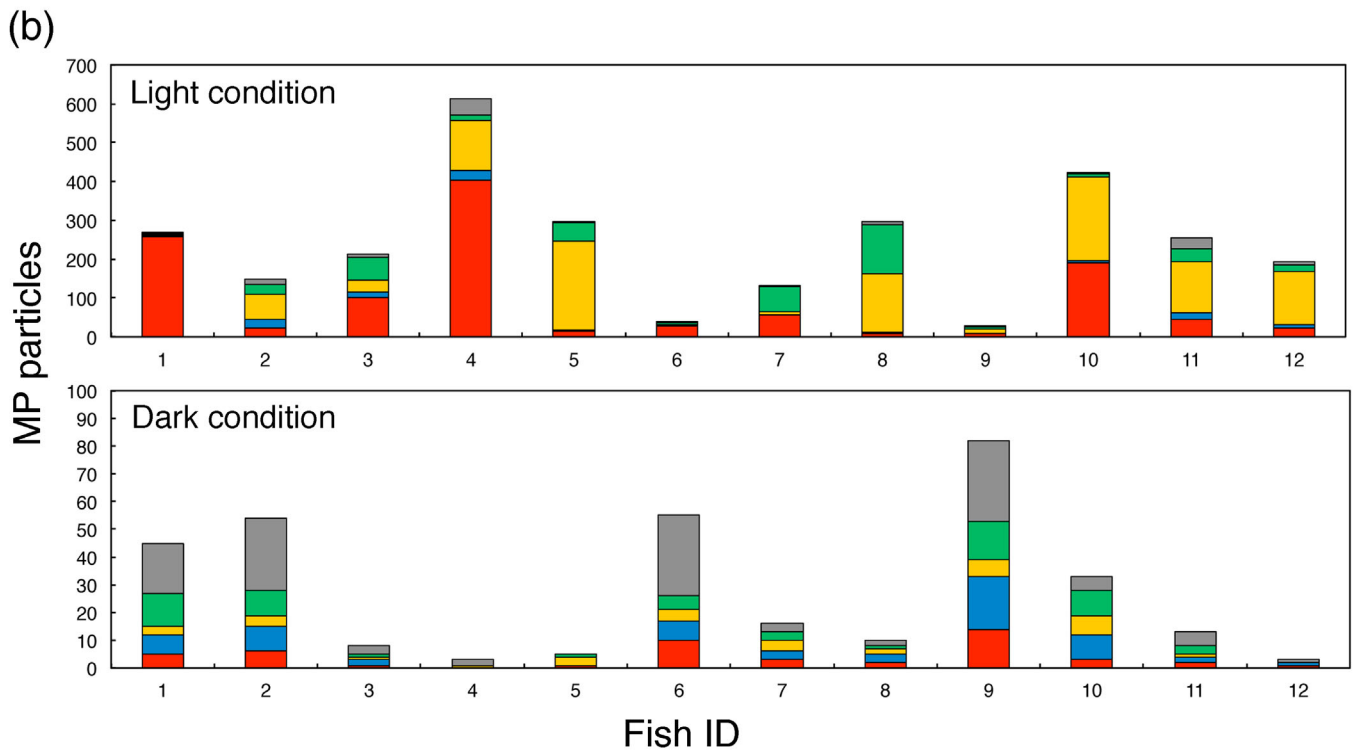
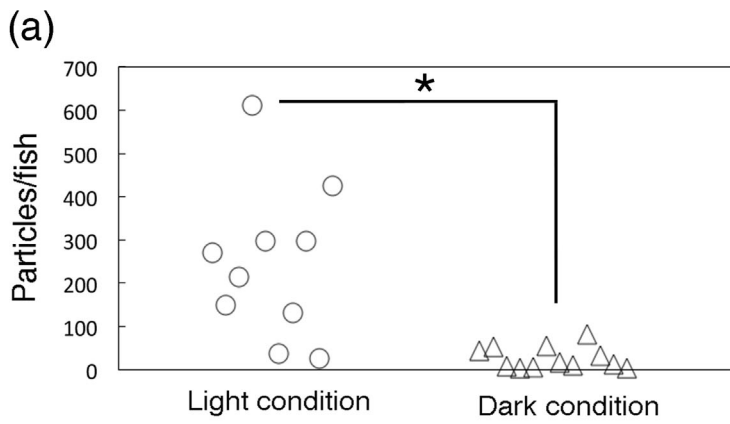


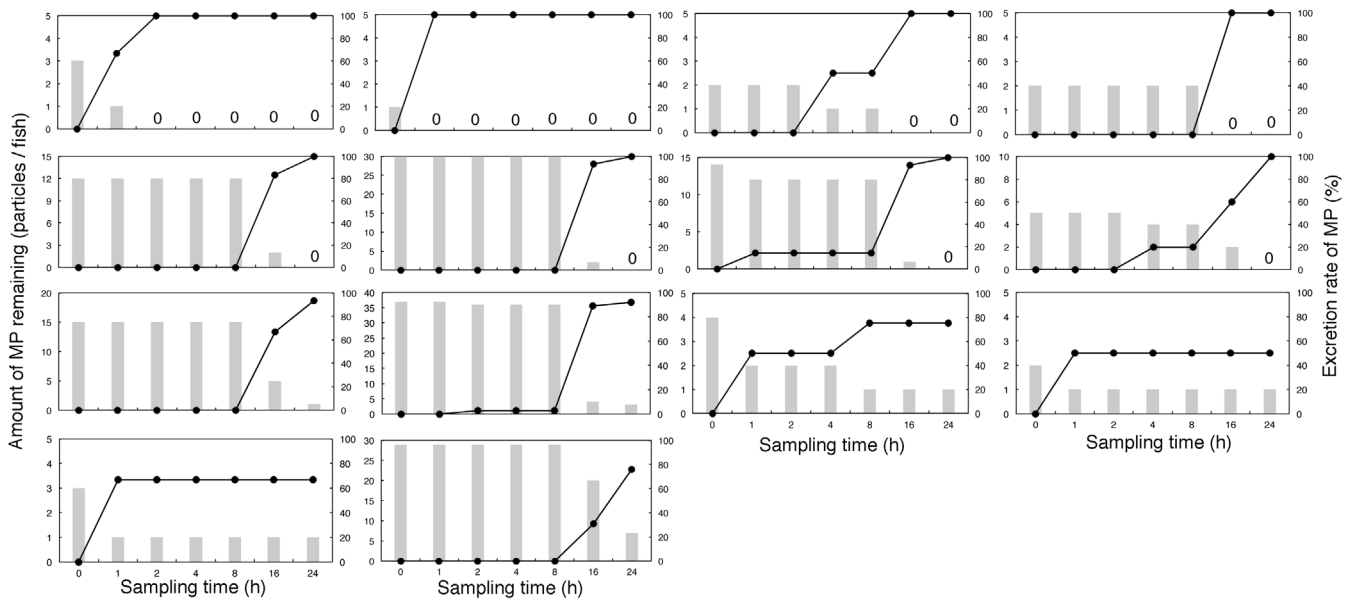
(b) Zebrafish

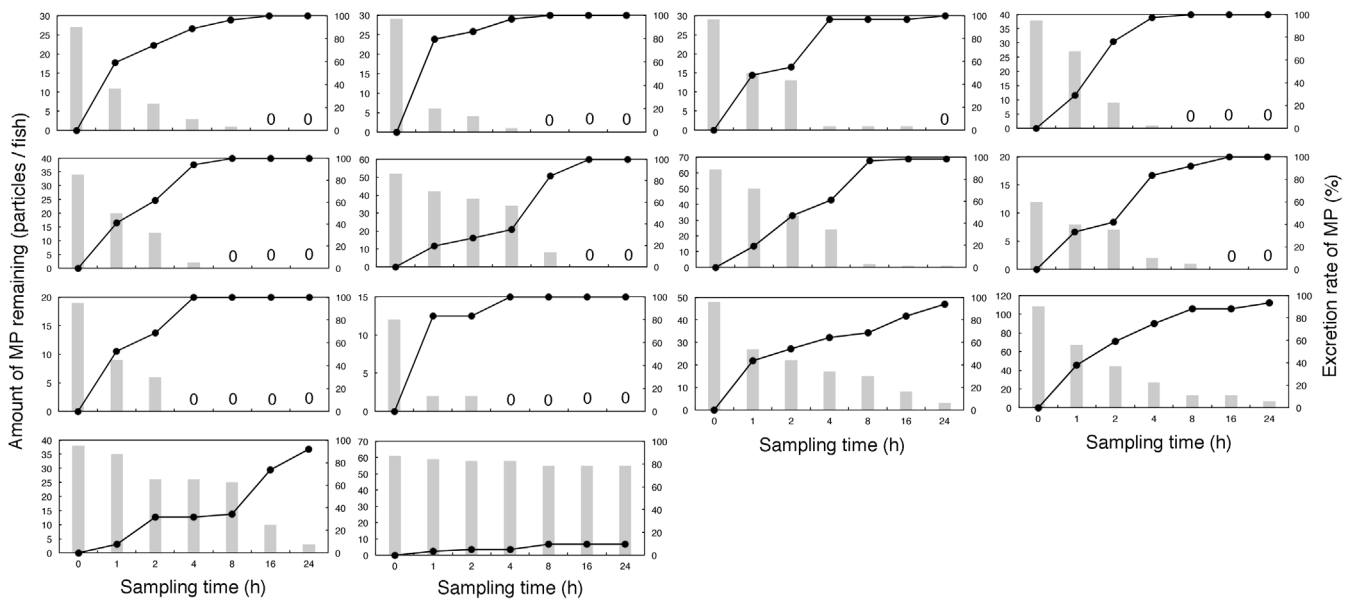


(c) Indian medaka

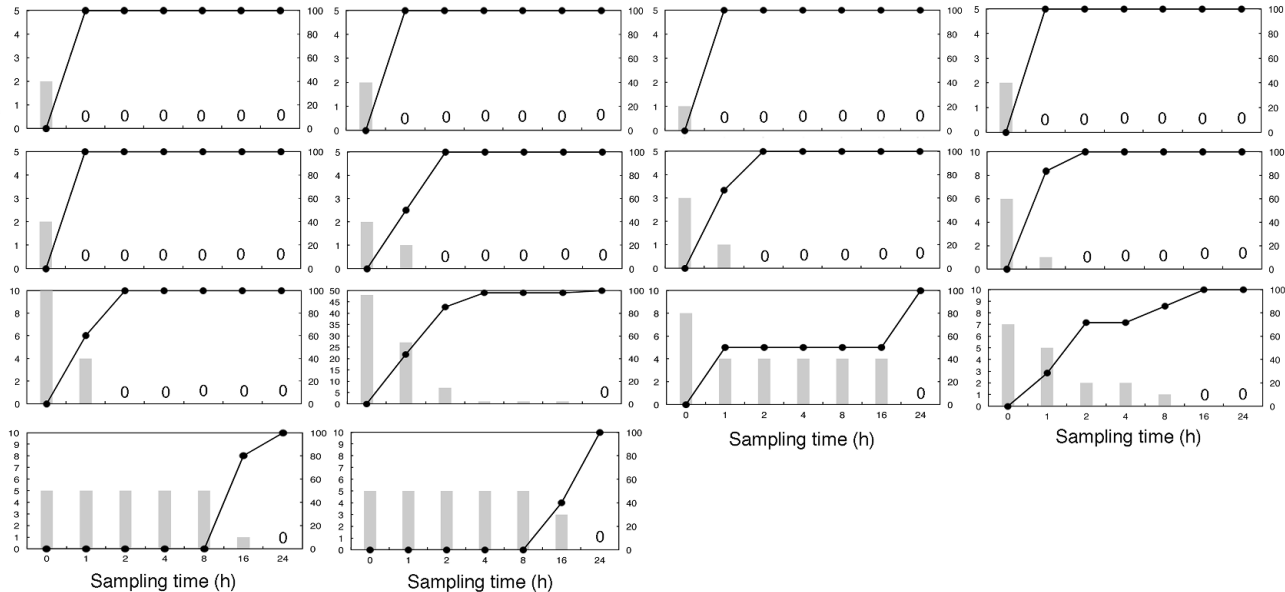




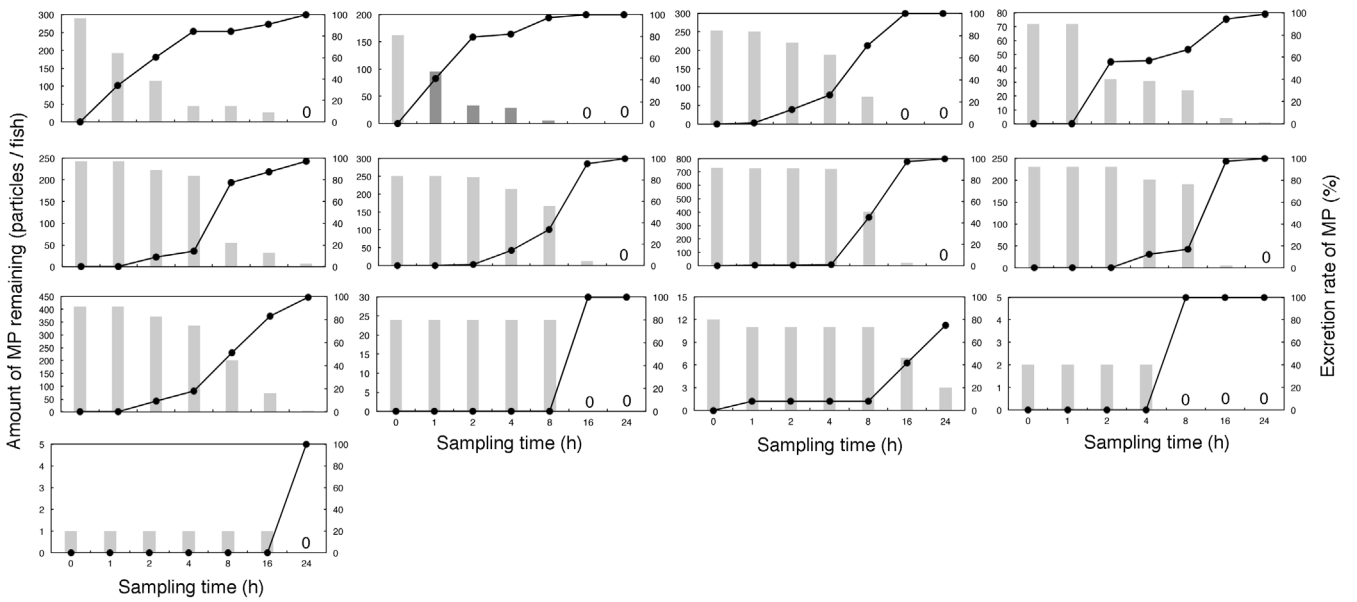




Amount of MP remaining (particles / fish)



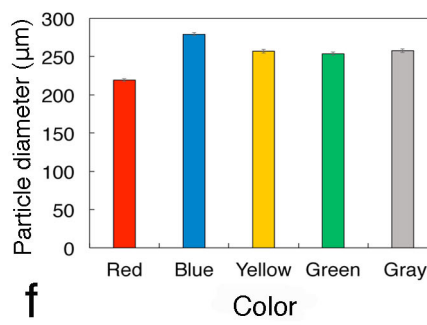
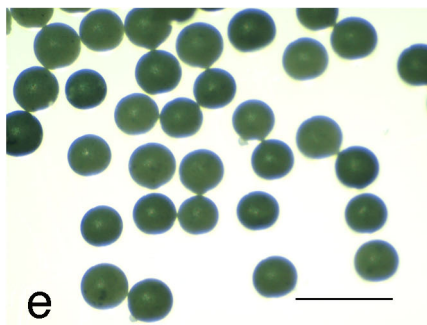
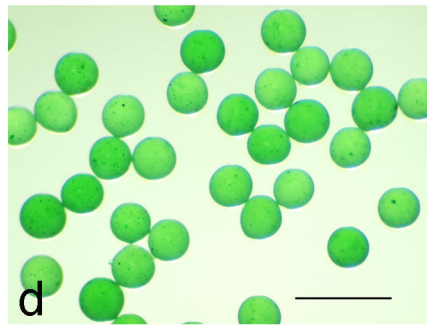
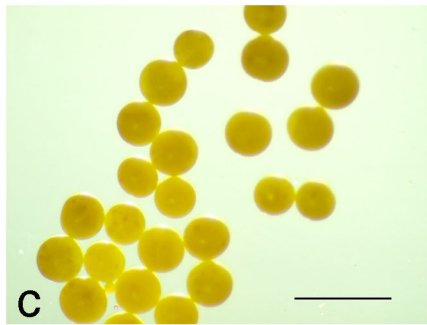
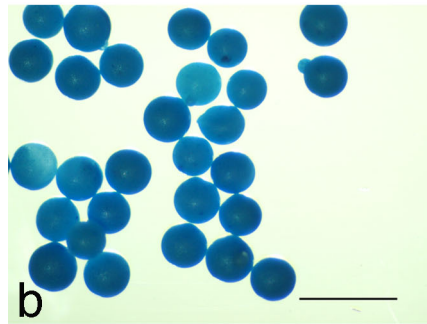
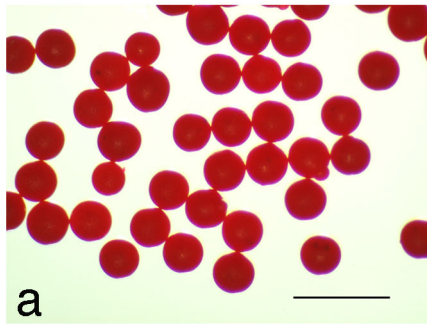
Excretion rate of MP (%)



Table

Table 1. The number of fish that ingested microplastic (MP) of various colors for the species Japanese medaka, zebrafish, Indian medaka, and clown anemonefish.

Species	MP color	Number of fish (percent of total)
Japanese medaka	Red	0 of 14 (0%)
	Blue	3 of 14 (21%)
	Yellow	0 of 14 (0%)
	Green	9 of 14 (64%)
	Gray	3 of 14 (21%)
Zebrafish	Red	12 of 14 (85%)
	Blue	12 of 14 (85%)
	Yellow	12 of 14 (85%)
	Green	12 of 14 (85%)
	Gray	13 of 14 (92%)
Indian medaka	Red	3 of 14 (21%)
	Blue	0 of 14 (0%)
	Yellow	1 of 14 (7%)
	Green	5 of 14 (35%)
	Gray	0 of 14 (0%)
Clown anemonefish	Red	13 of 14 (92%)
	Blue	12 of 14 (85%)
	Yellow	13 of 14 (92%)
	Green	14 of 14 (100%)
	Gray	10 of 14 (71%)



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Declaration of interests

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

The authors declare the following financial interests/personal relationships which may be considered as potential competing interests: